

# BMS

## Next generation sequencing

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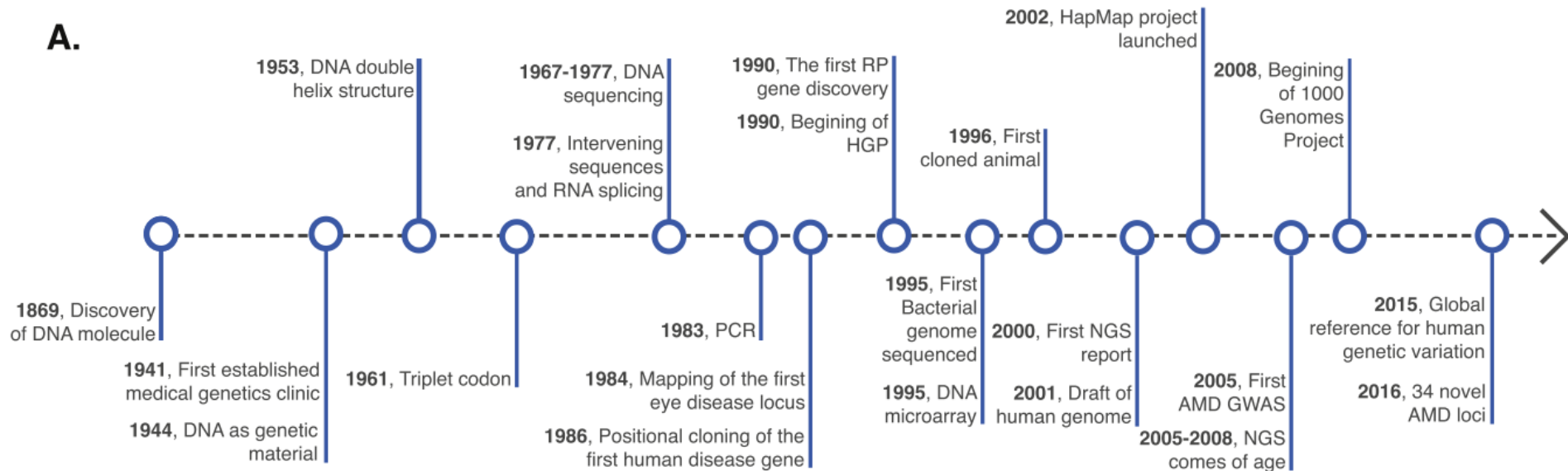
# Human Genome Project



[https://www.researchgate.net/figure/Human-genome-Project-published-in-Nature-http-wwwnaturecom-nature-journal\\_fig2\\_329830959](https://www.researchgate.net/figure/Human-genome-Project-published-in-Nature-http-wwwnaturecom-nature-journal_fig2_329830959)

# Timeline for Human Genome Project

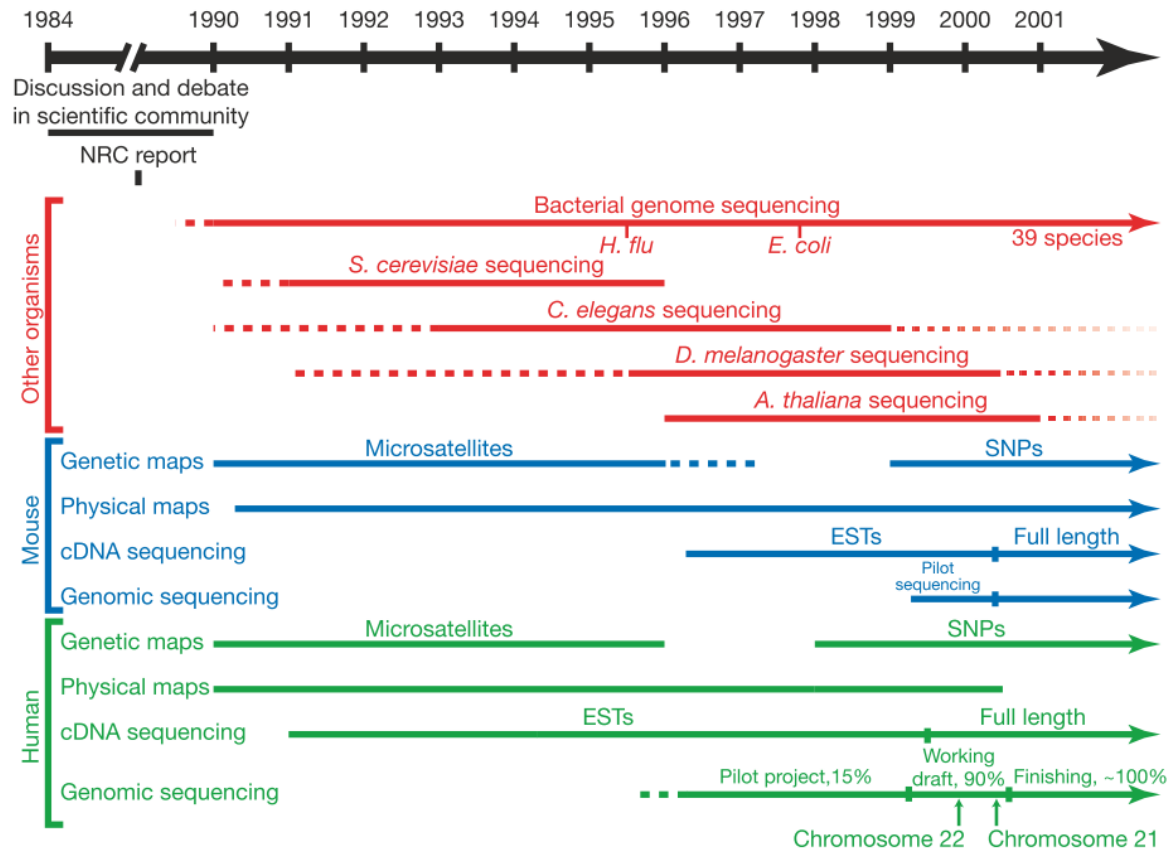
A.



<https://www.oreilly.com/ideas/genomics-and-the-role-of-big-data-in-personalizing-the-healthcare-experience>

# Timeline until HGP

- Human genome project formally launched in 1990 and was declared complete on April 14, 2003



# White House on 26 June 2000

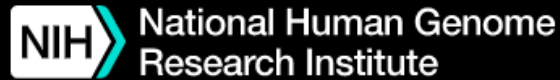
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<https://www.theguardian.com/science/gallery/2010/jun/23/human-genome-project>

# Cost for Human Genome Project

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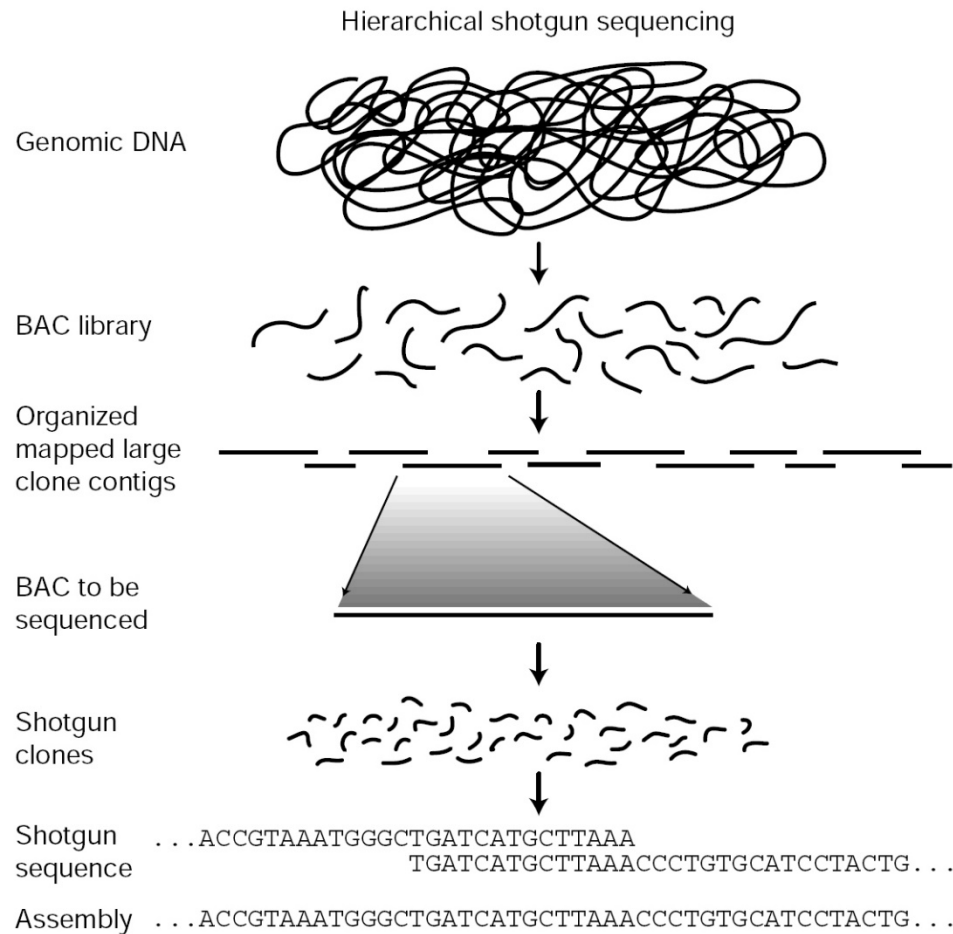


## How much did it cost?

In 1990, Congress established funding for the Human Genome Project and set a target completion date of 2005. Although estimates suggested that the project would cost a total of \$3 billion over this period, the project ended up costing less than expected, about \$2.7 billion in FY 1991 dollars. Additionally, the project was completed more than two years ahead of schedule.

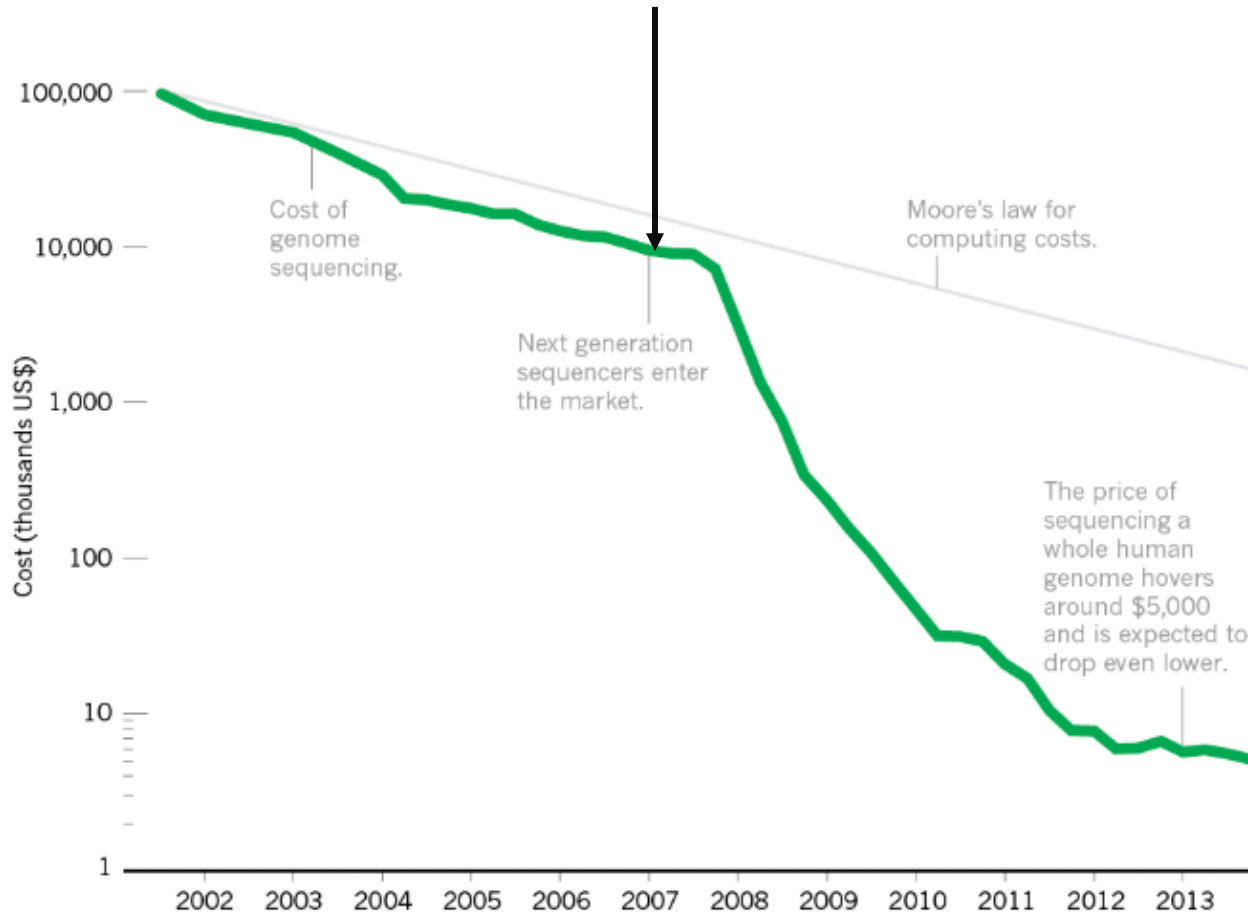
[https://www.researchgate.net/figure/Human-genome-Project-published-in-Nature-http-wwwnaturecom-nature-journal\\_fig2\\_329830959](https://www.researchgate.net/figure/Human-genome-Project-published-in-Nature-http-wwwnaturecom-nature-journal_fig2_329830959)

# Strategy for sequencing



# Cost for genome sequencing

Next Generation Sequencing platform was appeared as game exchanger in sequencing field



[https://www.researchgate.net/figure/a-Cost-per-base-of-the-different-sequencing-techniques-as-a-function-of-time-The-gray\\_fig2\\_271772842](https://www.researchgate.net/figure/a-Cost-per-base-of-the-different-sequencing-techniques-as-a-function-of-time-The-gray_fig2_271772842)

# Next Generation Sequencing Platform

- Roche/454 (GS FLX+/GS Junior)
- Illumina Genome Analyzer (HiSeq/MiSeq/NextSeq)
- Life Technologies (3500 Genetic Analyzer)
- Ion Torrent Proton/PGM)
- Applied Biosystems (SOLiD, 3730xl DNA Analyzer )



illumina®



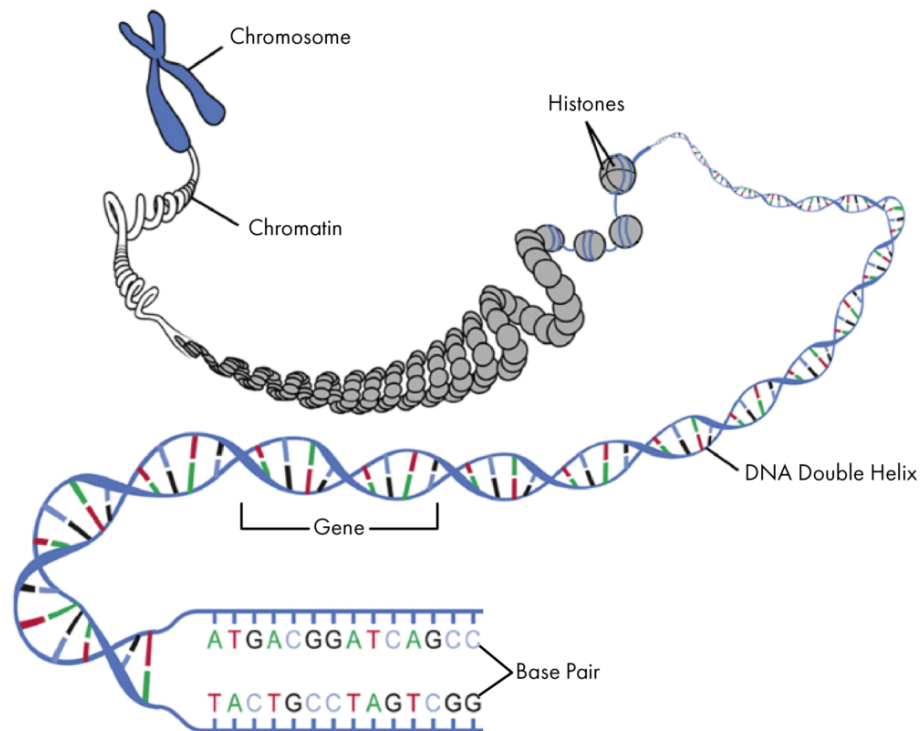
ion torrent



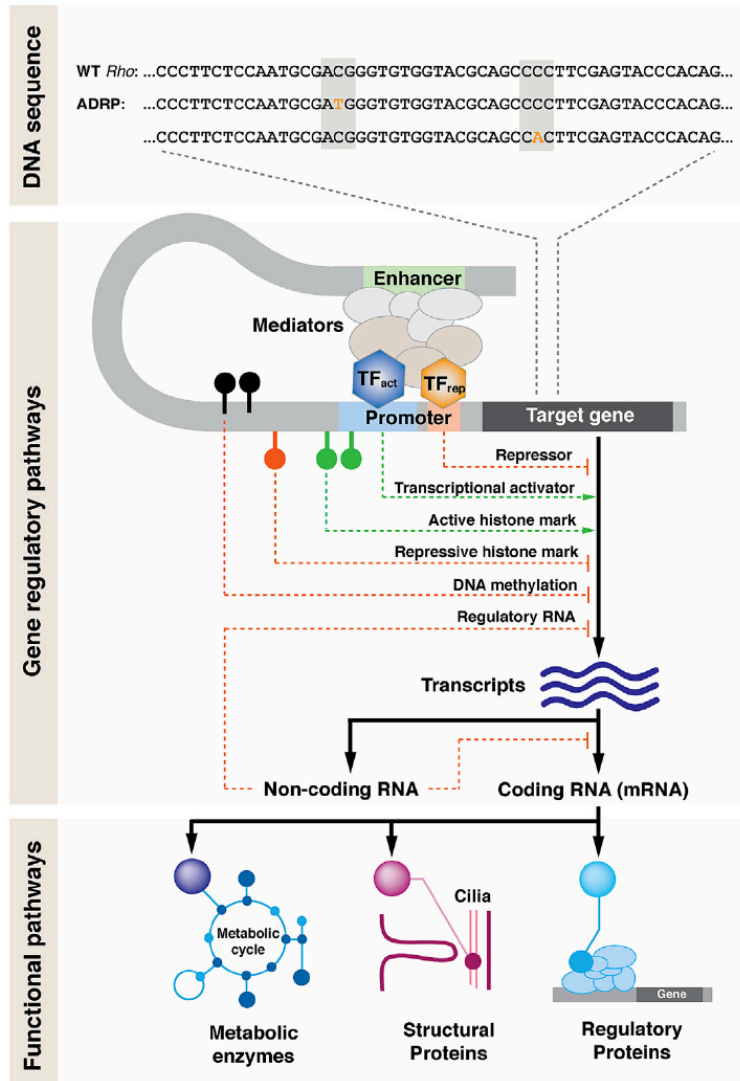
# “DNA Sequencing”

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- DNA sequencing involves the use of various methods for determining the order of the nucleotide bases — **adenine**, **cytosine**, **guanine**, and **thymine** — in a molecule of DNA

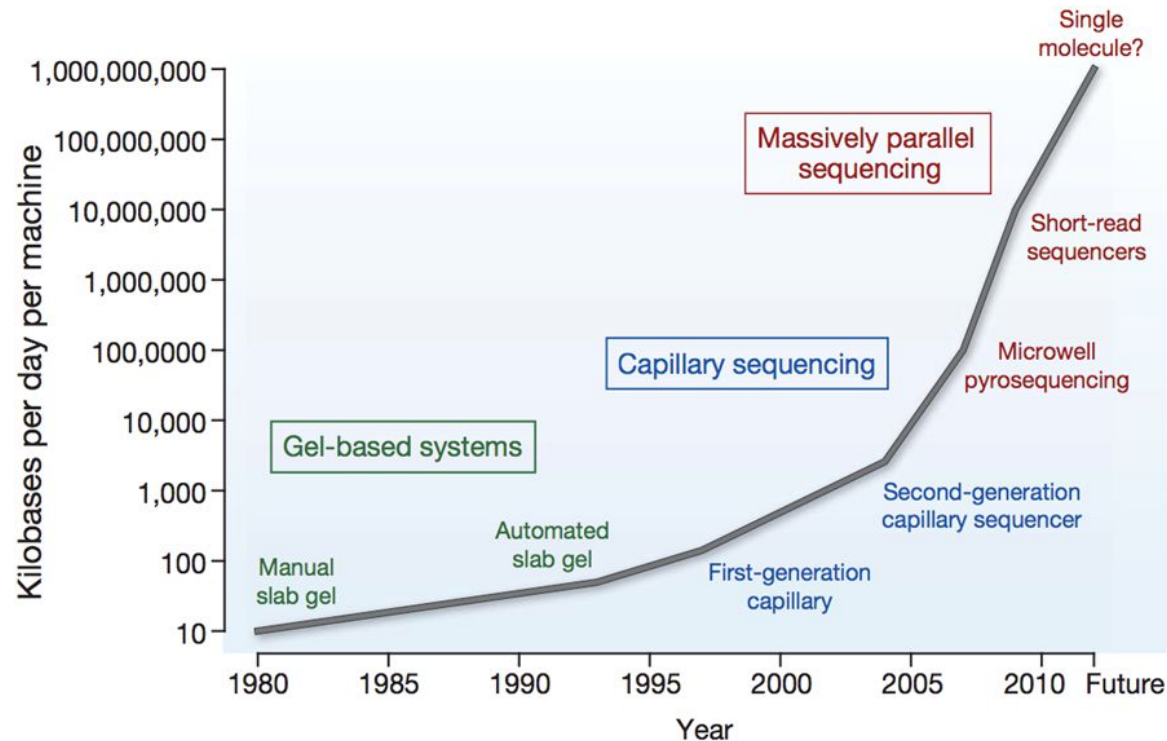


# Information including in sequence

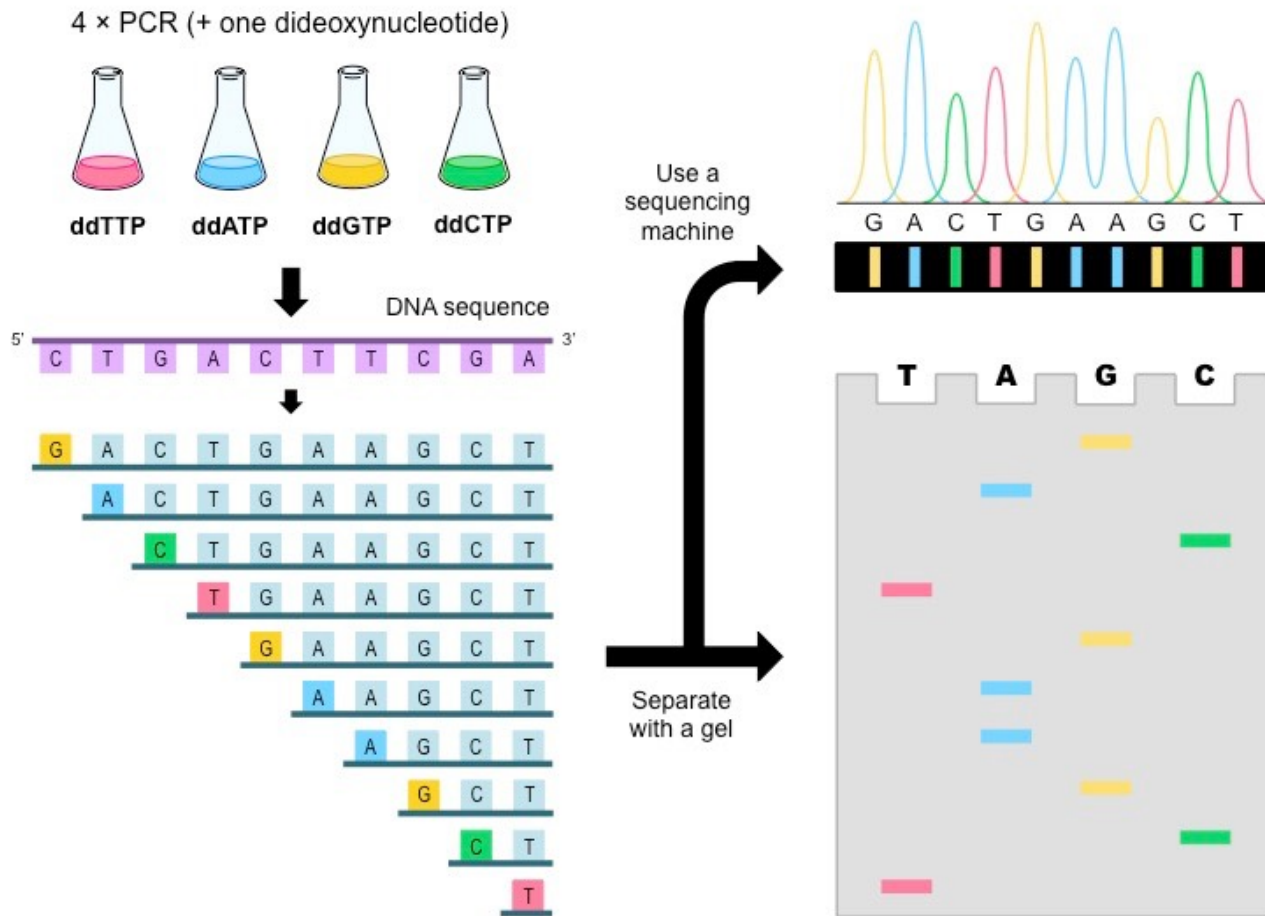


- DNA sequence includes not only template for transcription, but also act as regulation factors

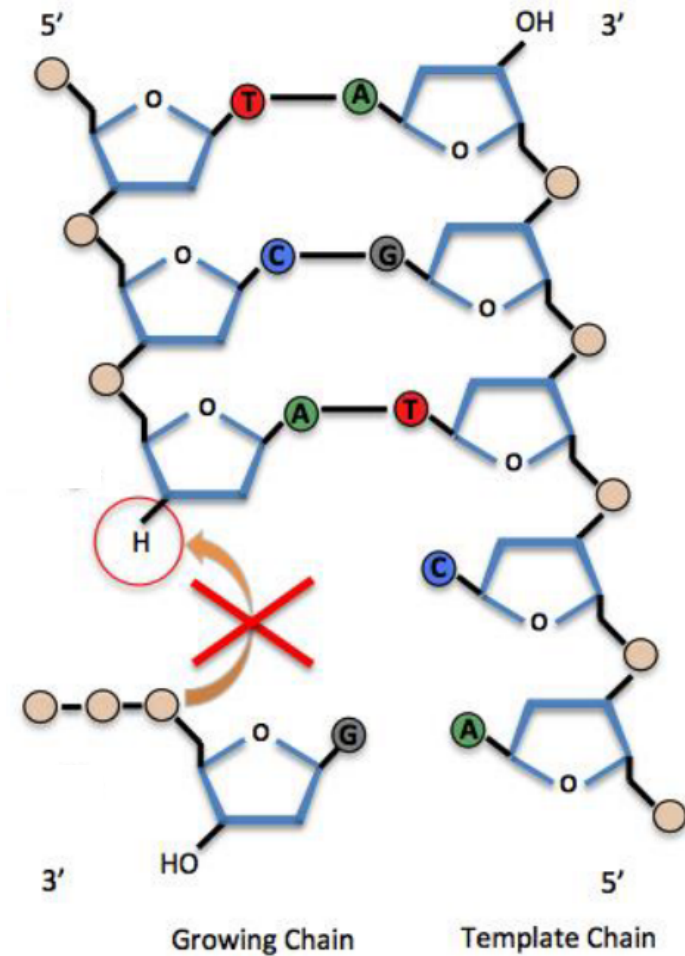
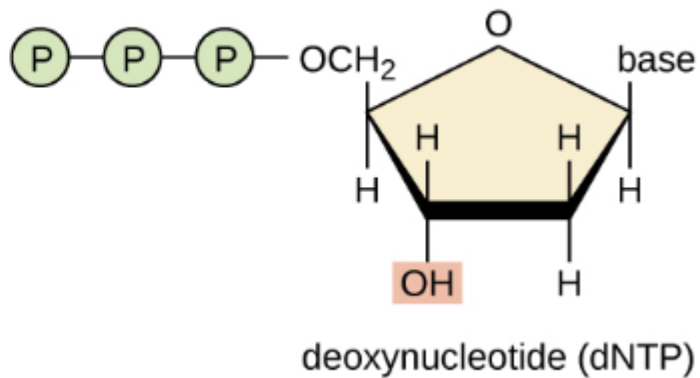
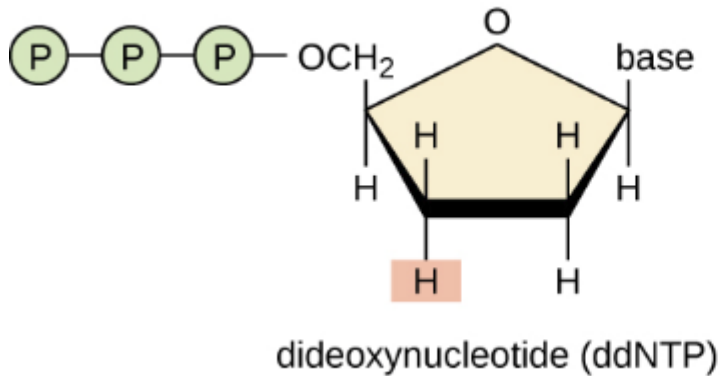
# History of Sequencing technology



# Sanger Sequencing



# Sanger Sequencing



<https://open.library.ubc.ca/cIRcle/collections/ubctheses/24/items/1.0166738>

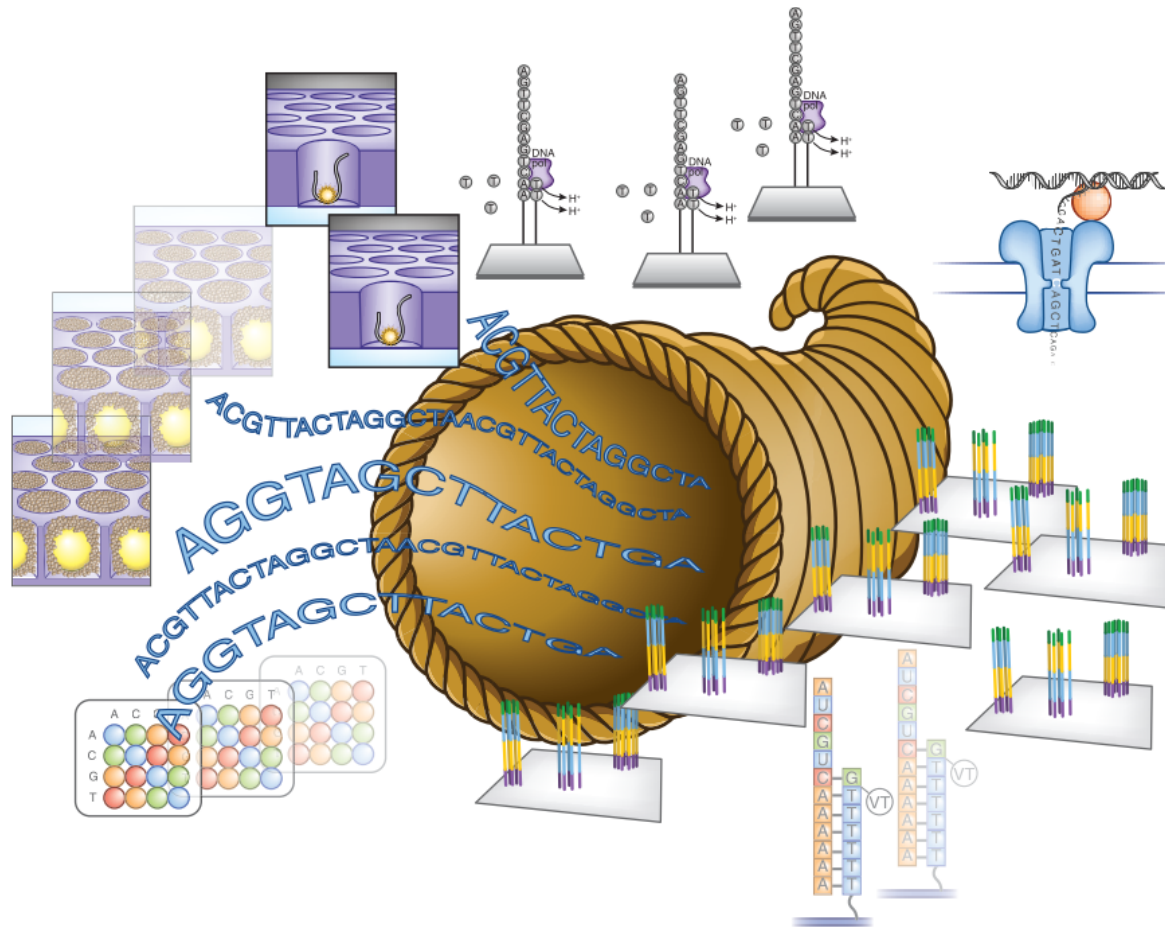
# Sanger Sequencing

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Technology	Analysis time	Average read length	Throughput (Mb/ <a href="#">h</a> )
Slab gel	6–8 hours	700 bp	0.0672
Capillary array electrophoresis	1–3 hours	700 bp	0.166

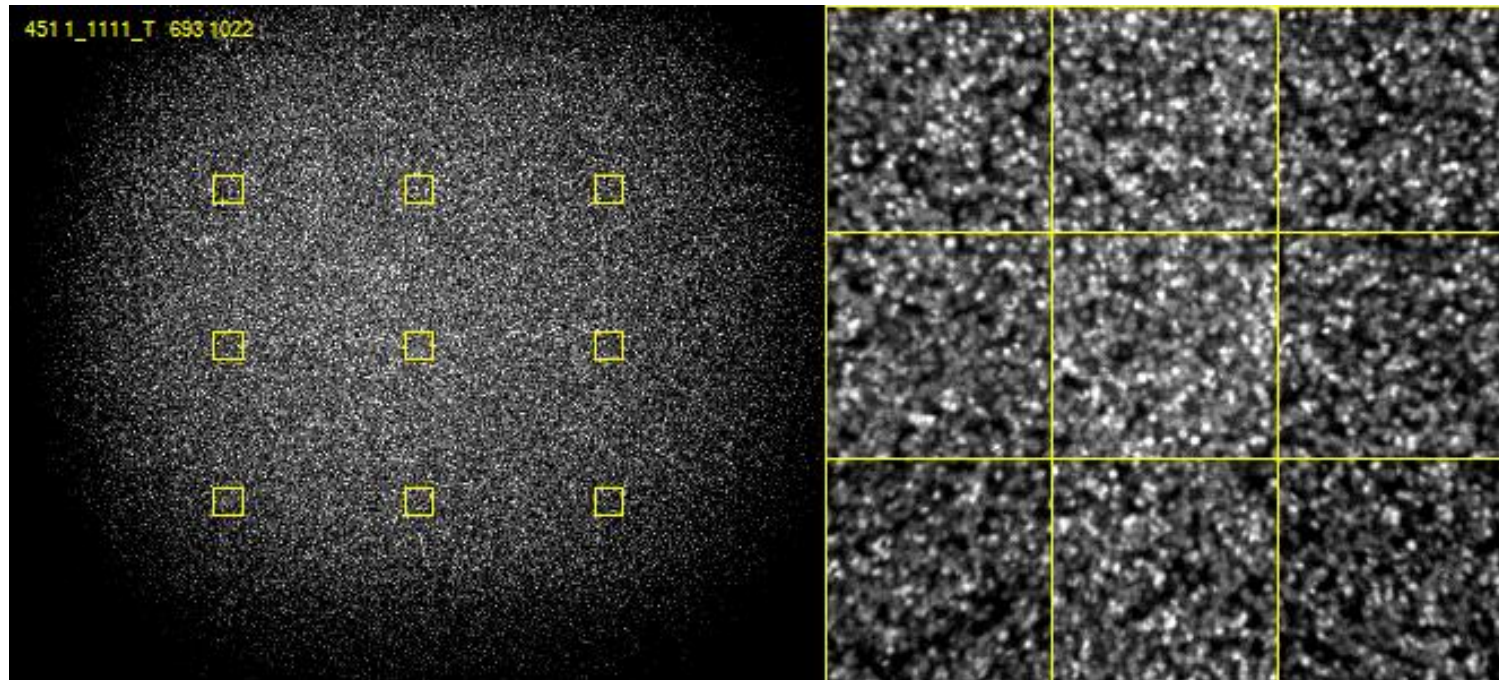
[https://en.wikipedia.org/wiki/Sanger\\_sequencing](https://en.wikipedia.org/wiki/Sanger_sequencing)

# Next generation Sequencing



# Next generation Sequencing

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# Definition of “Massive parallel”

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- **Massive**  
[형용사] (육중하면서) 거대한
- **Parallel**  
[형용사] 병행[병렬]의
- **Massive parallel sequencing**  
The DNA is sequenced via spatially separated, clonally amplified DNA templates or single DNA molecules in a flow cell.

**Massive parallel sequencing = Next Generation Sequencing**

# Next Generation Sequencing Platform

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- Roche/454 (GS FLX+/GS Junior)
- Illumina Genome Analyzer (HiSeq/MiSeq/NextSeq)
- Life Technologies (3500 Genetic Analyzer)
- Ion Torrent Proton/PGM)
- Applied Biosystems (SOLiD, 3730xl DNA Analyzer )



illumina



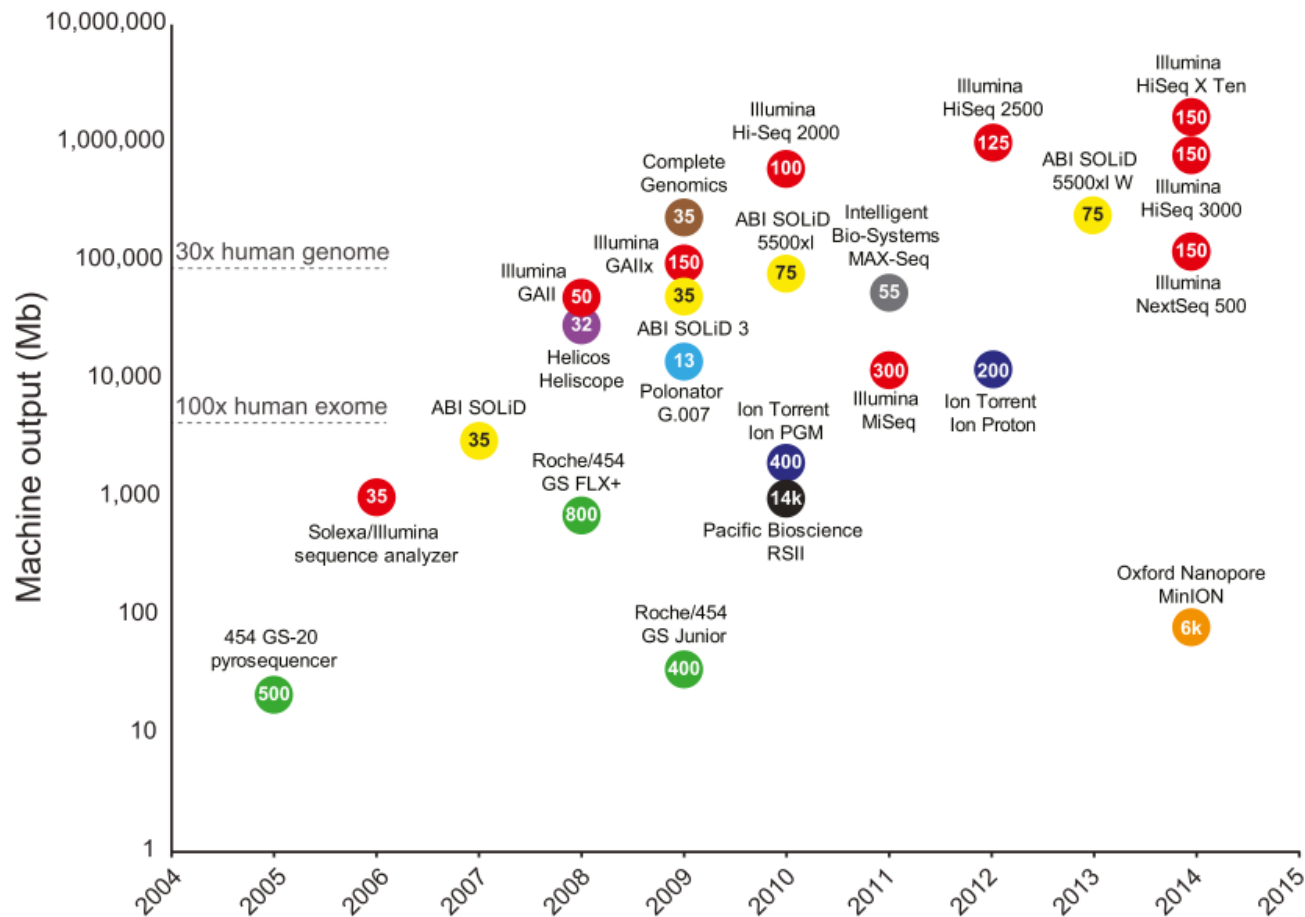
ion torrent  
♦ ★ △ ○ × □ + ∞



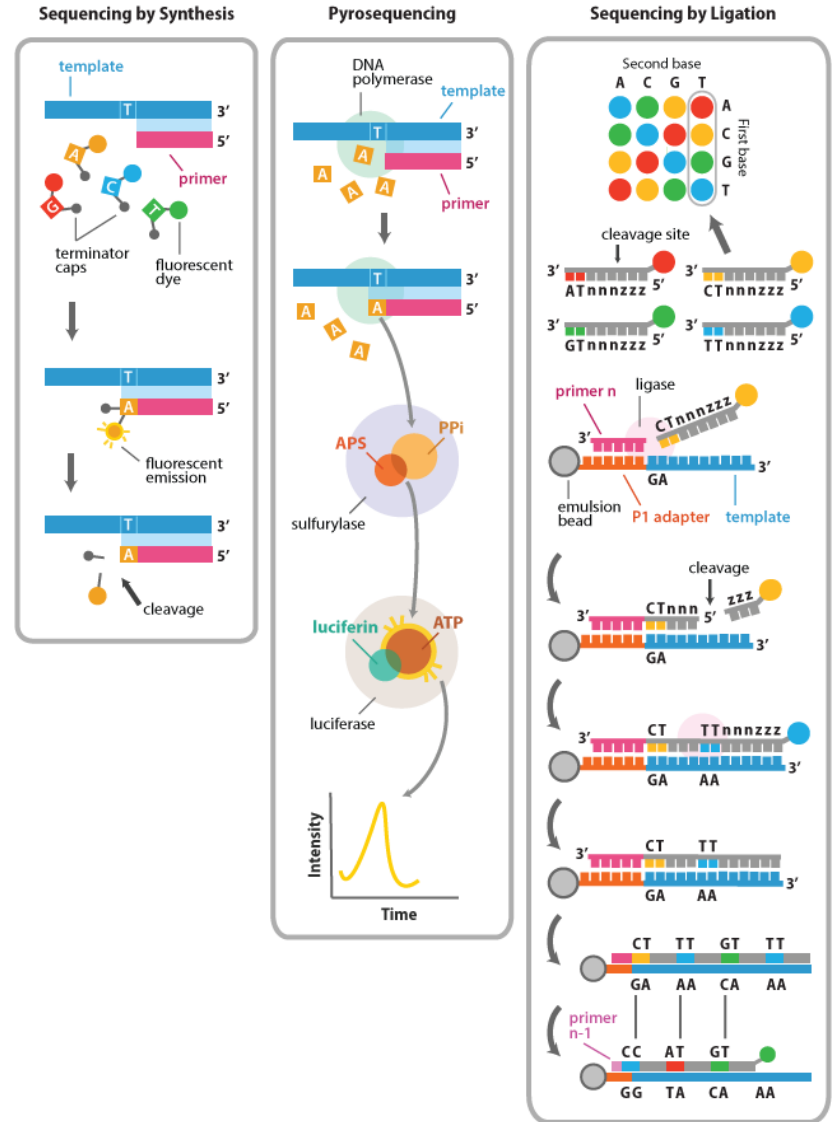
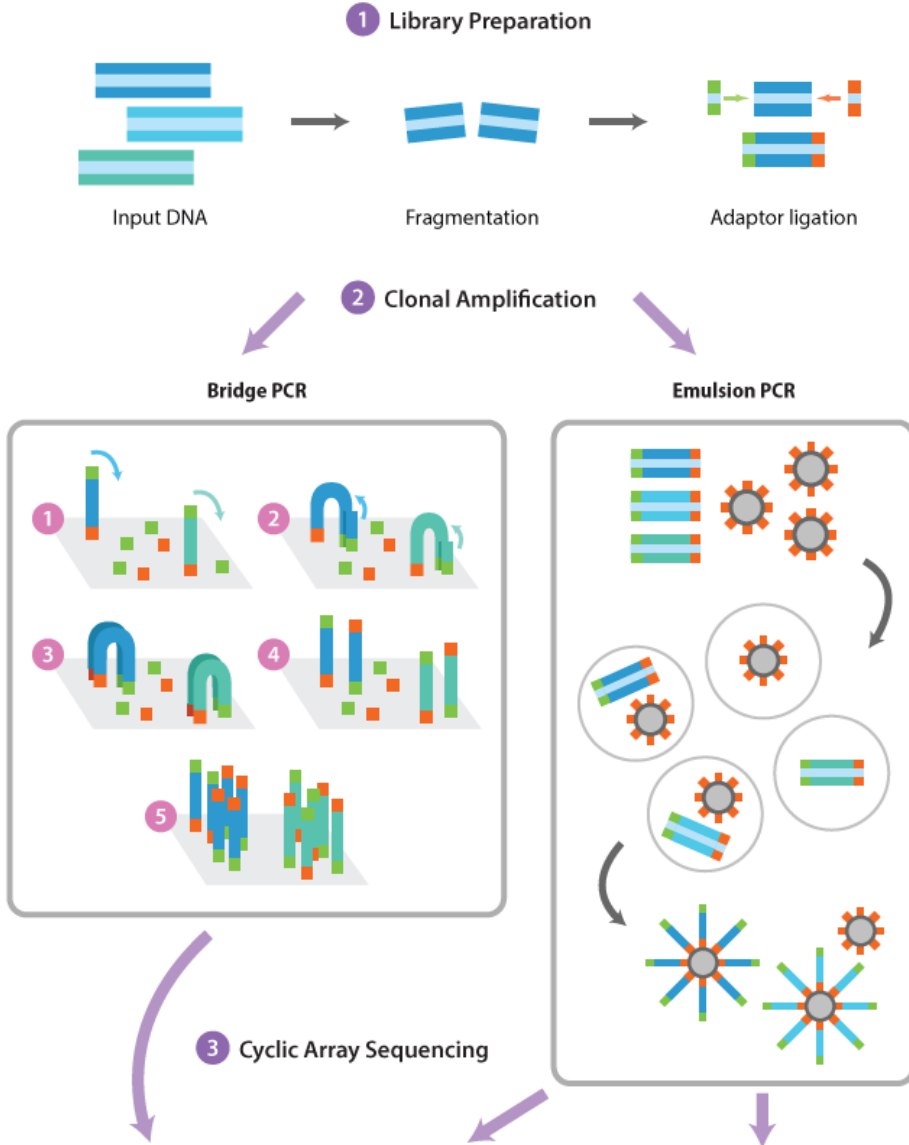
Roche

<http://www.vib.be/en/about-vib/annual-report/2012/research/activities/Pages/Service%20Facilities.aspx>

# Next Generation Sequencing Machine

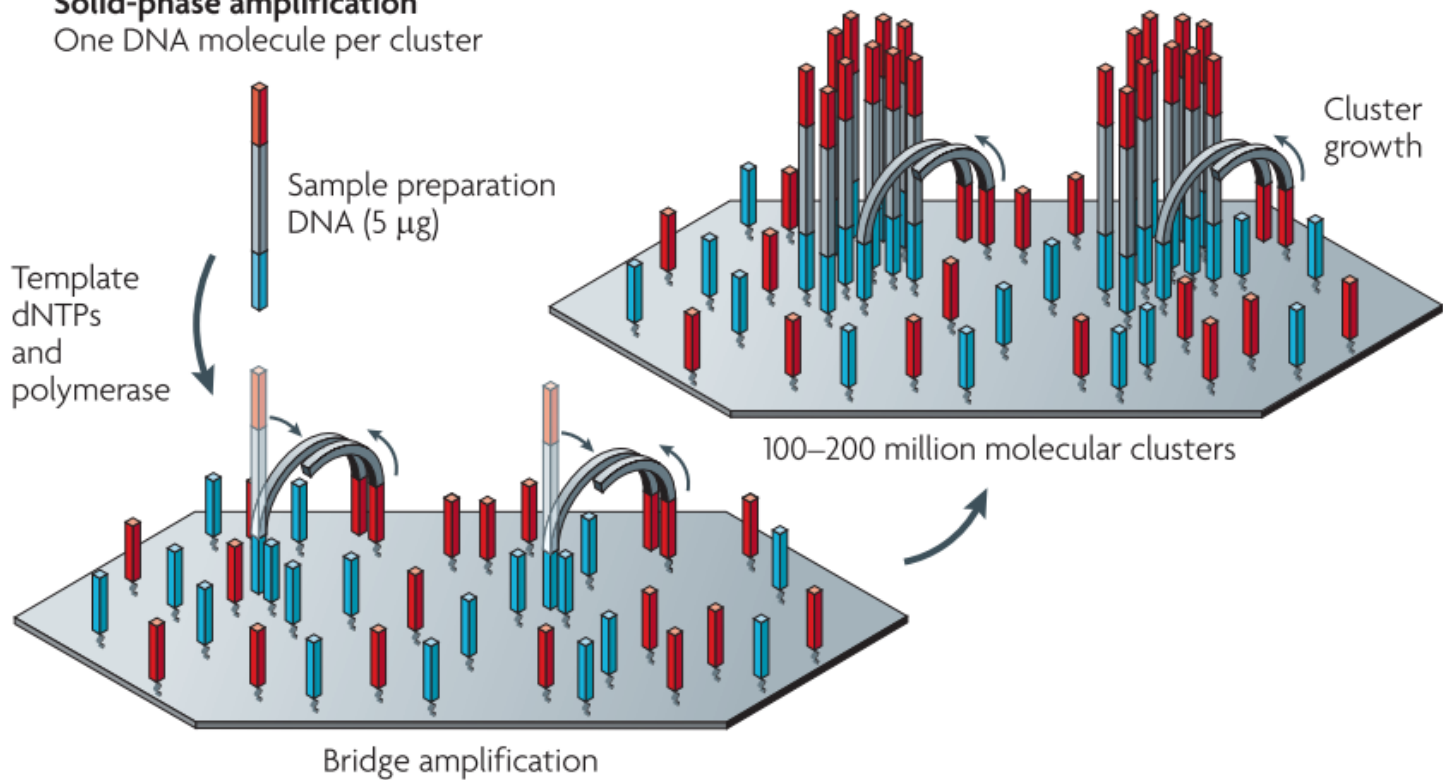


# Steps of NGS



# Clonal amplification - Bridge PCR

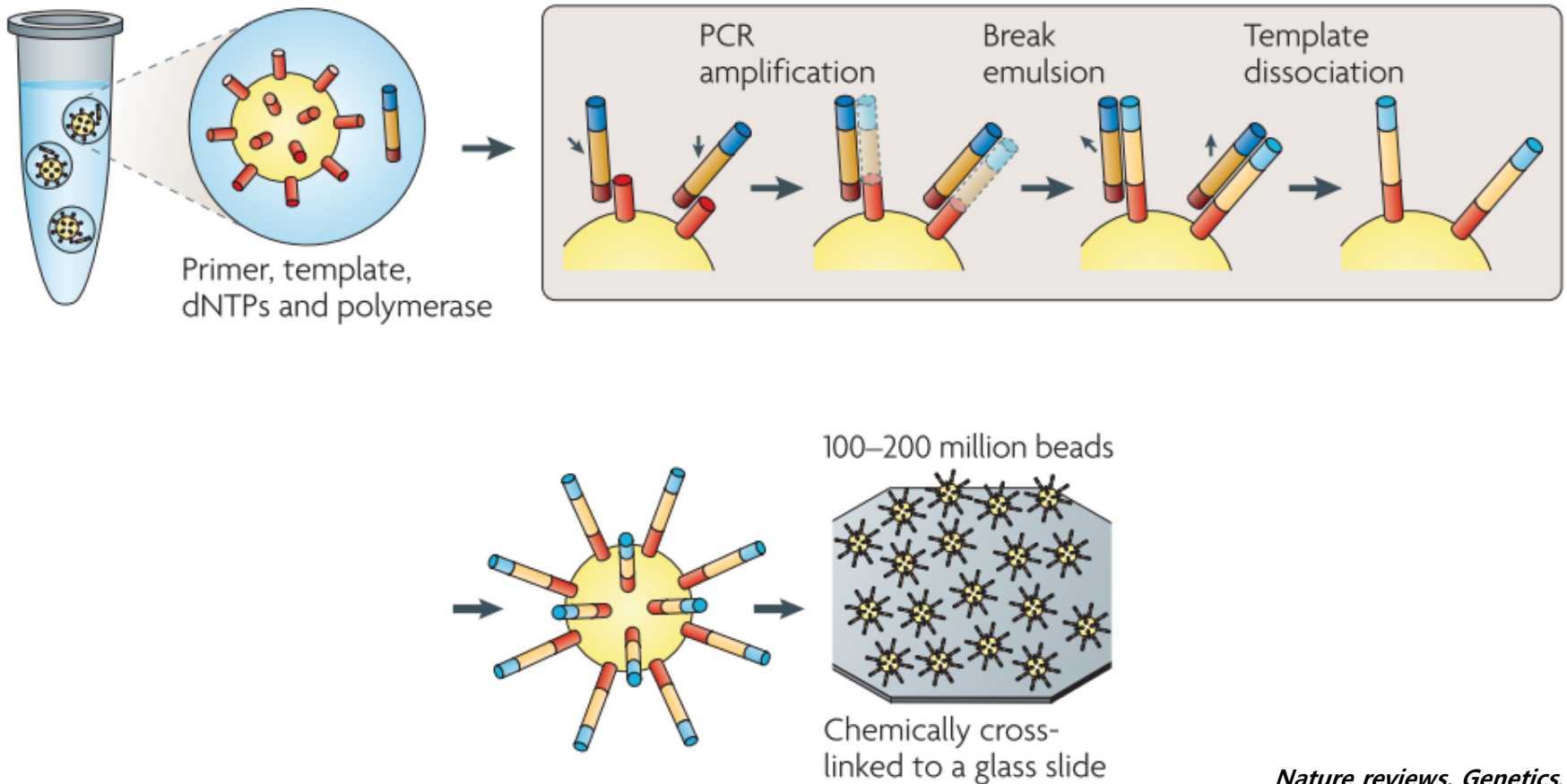
## **b** Illumina/Solexa Solid-phase amplification One DNA molecule per cluster



# Clonal amplification – Emulsion PCR

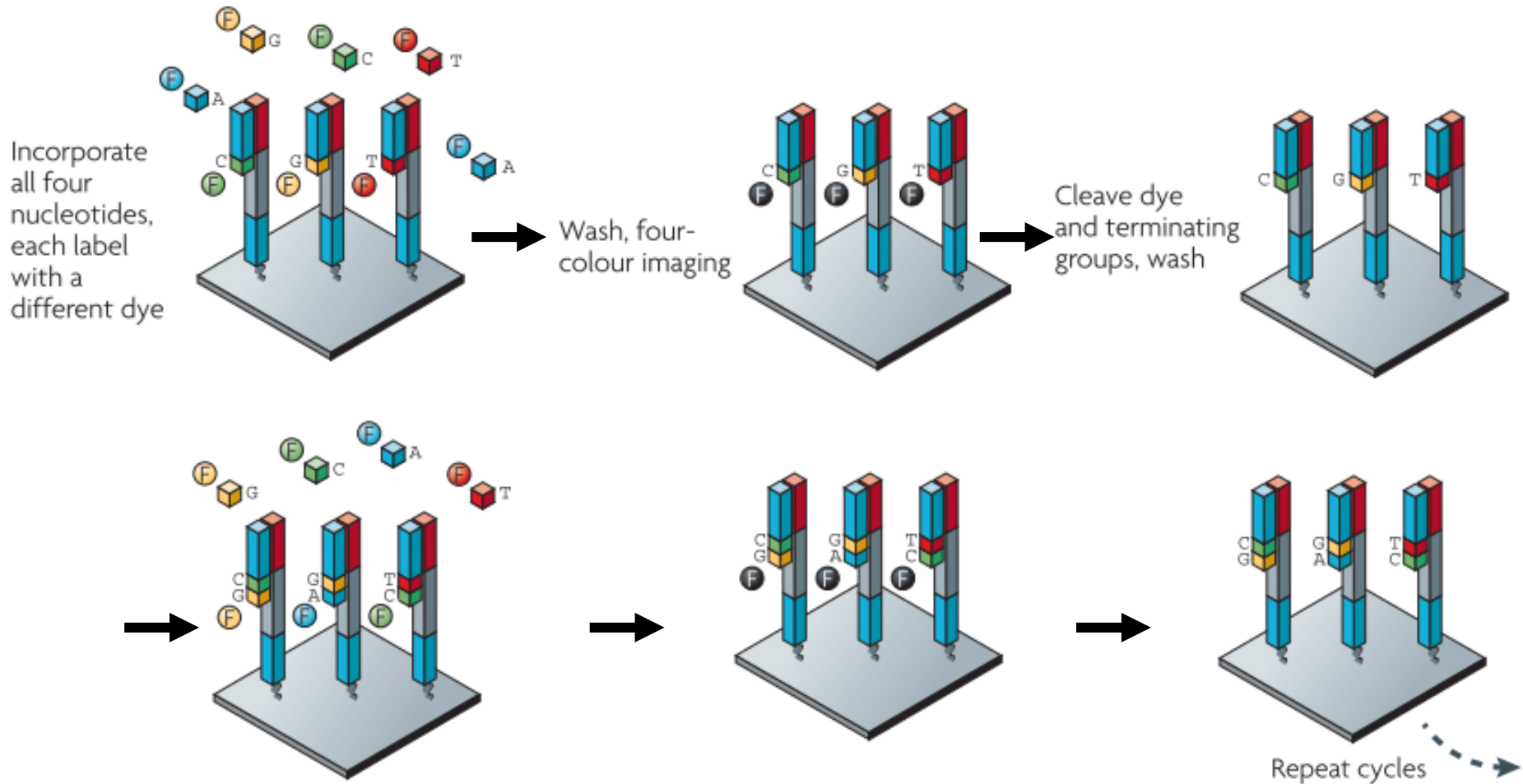
## a Roche/454, Life/APG, Polonator Emulsion PCR

One DNA molecule per bead. Clonal amplification to thousands of copies occurs in microreactors in an emulsion



*Nature reviews. Genetics*, 2009

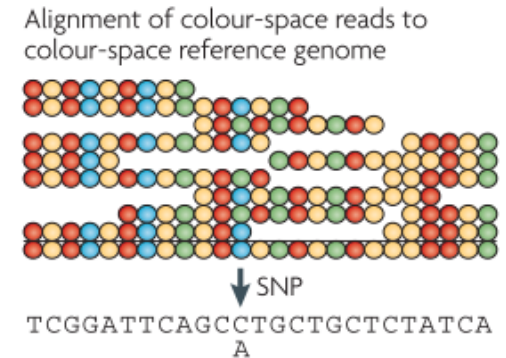
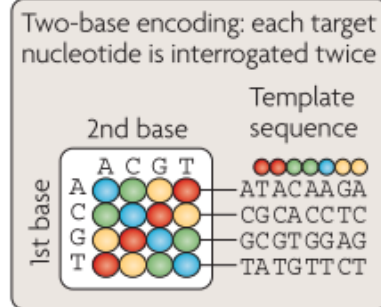
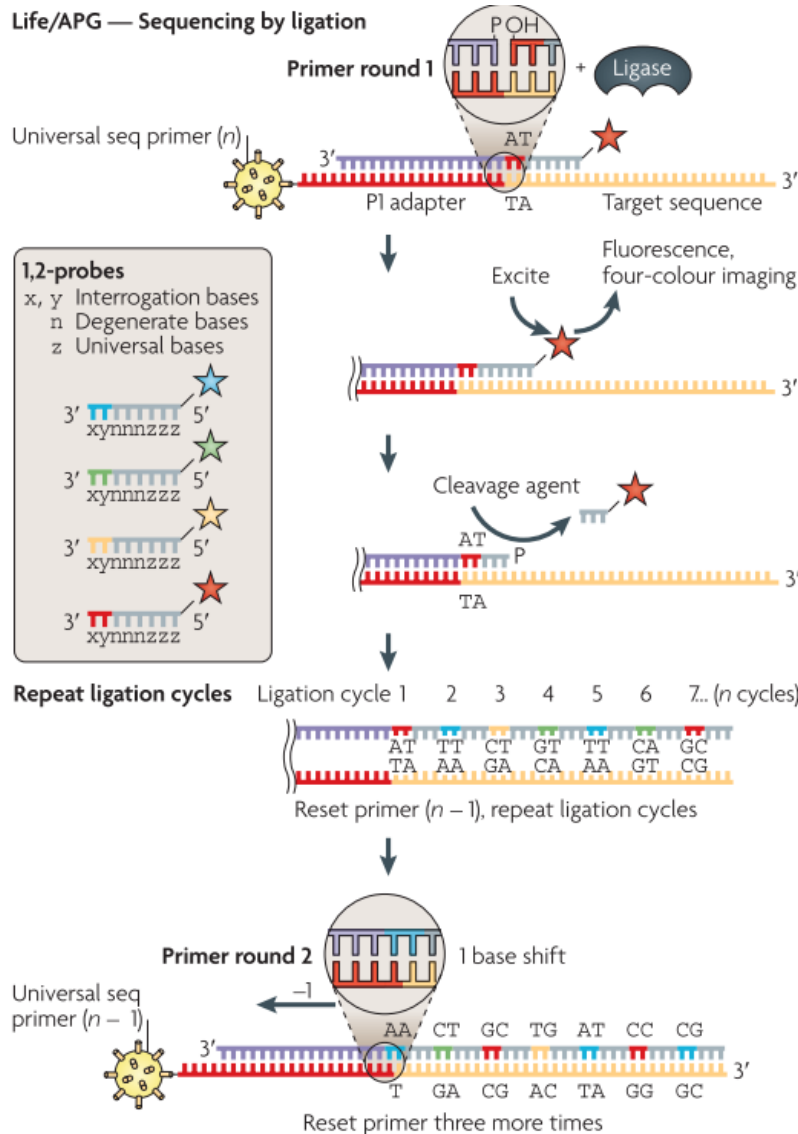
# Sequencing by sequencing





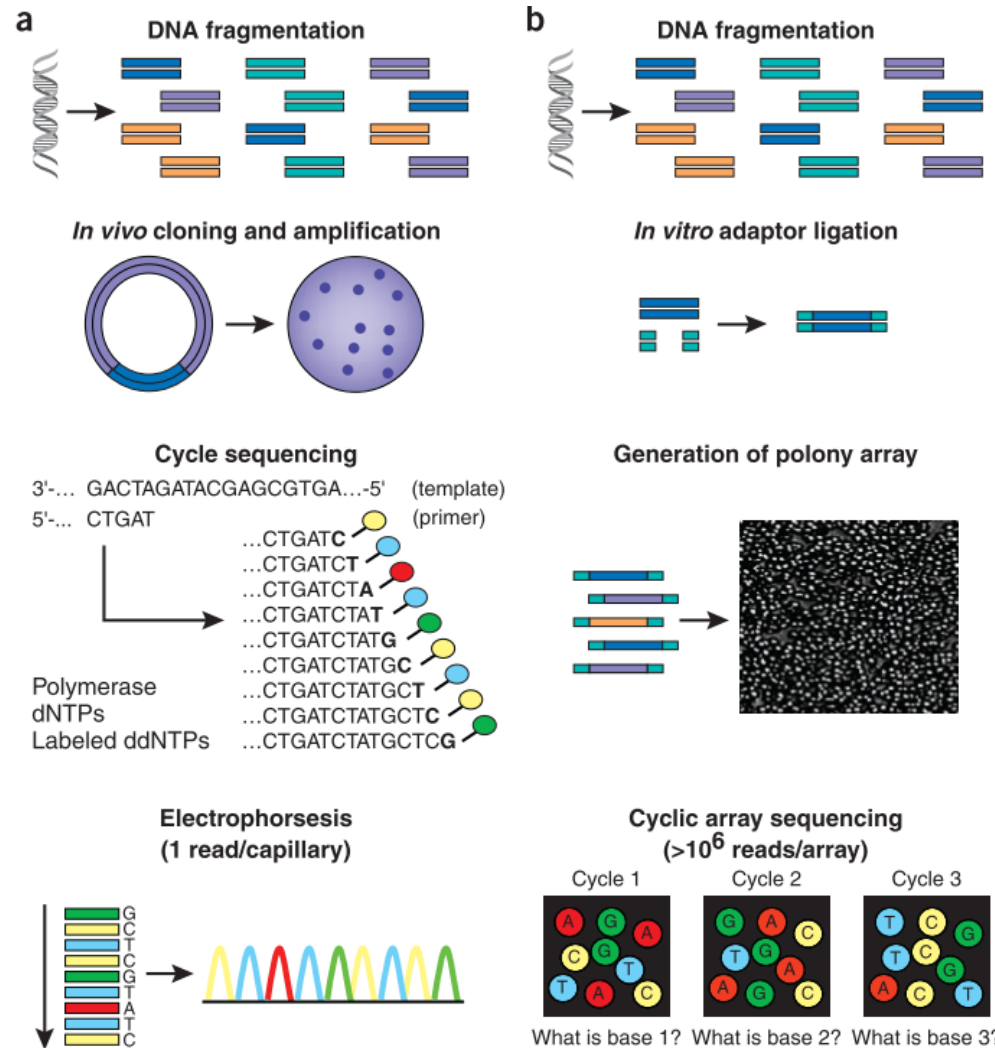
# Sequencing by ligation

Life/APG — Sequencing by ligation



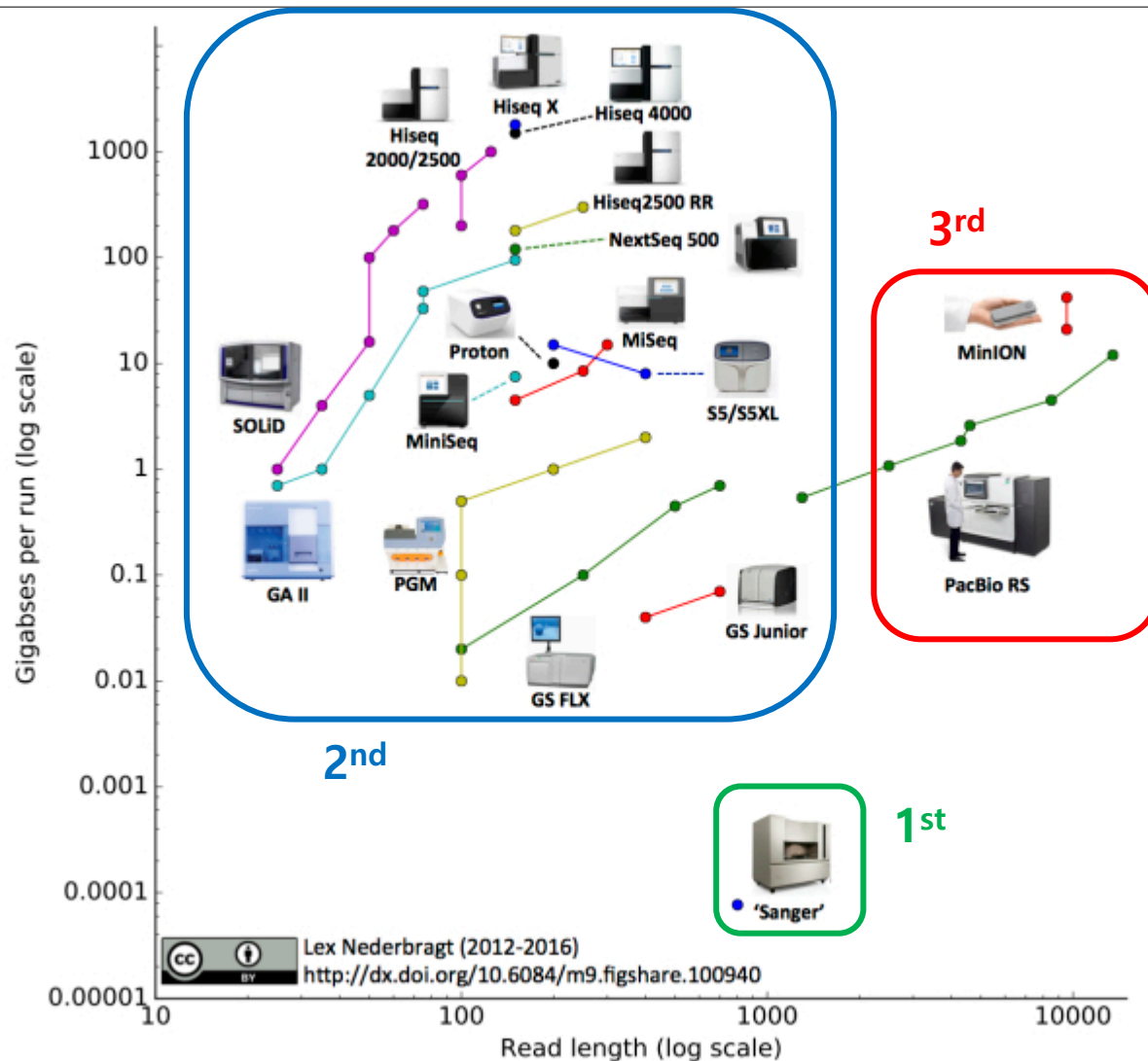
*Nature reviews. Genetics*, 2009

# Comparison between Sanger and NGS



*Nature biotechnology*, 2008

# History of Sequencing technology



<https://flxlexblog.wordpress.com/tag/illumina/>

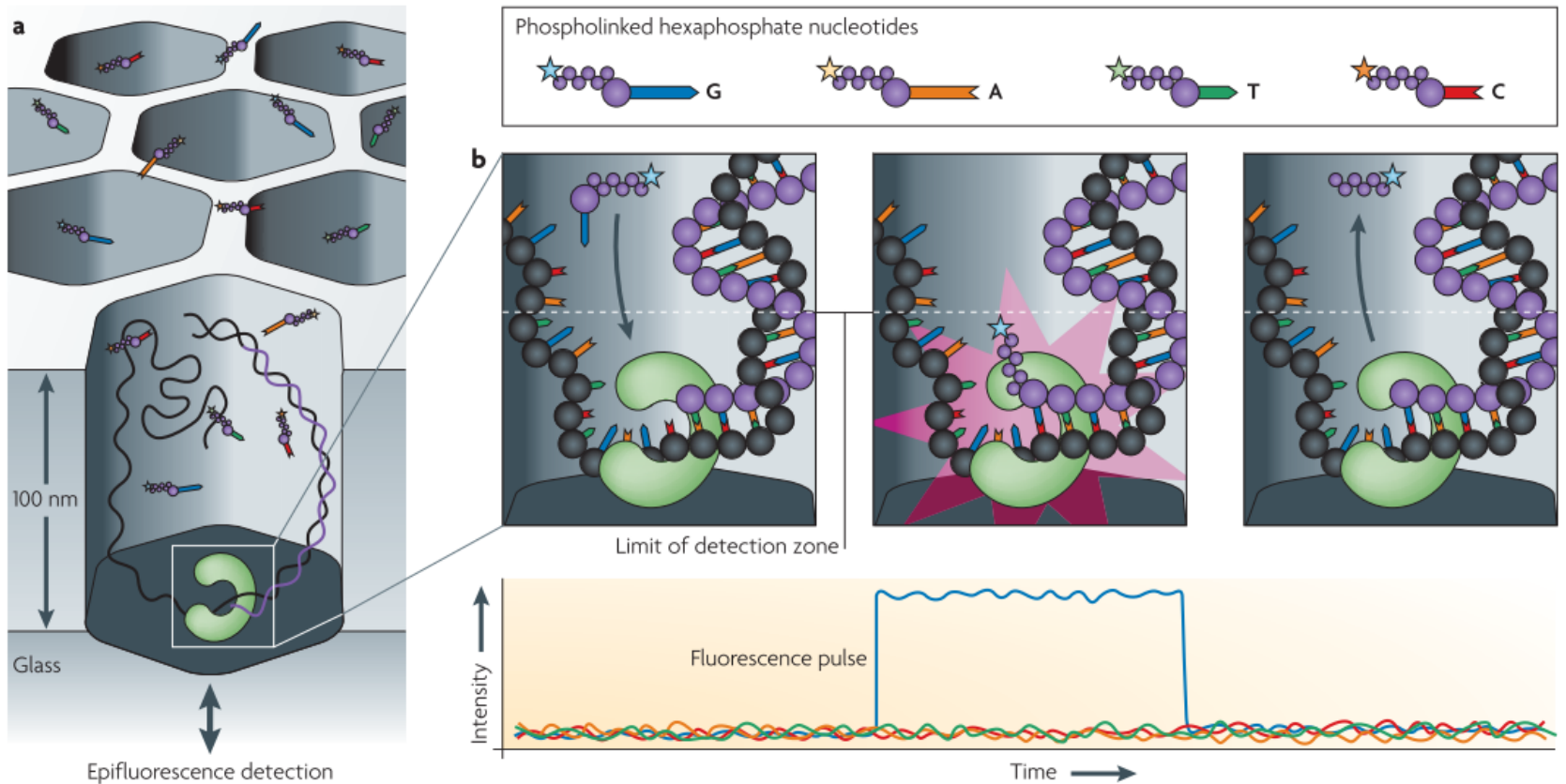
# Third Generation Sequencing

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- Third generation sequencing works by reading the nucleotide sequences at the single molecule level.
- Existing methods require breaking long strands of DNA into small segments then inferring nucleotide sequences by amplification and synthesis.
- Third Generation Sequencing = long-read sequencing
- PacBio and Oxford Nanopore provide manufactures



# PacBio



# Steps in PacBio

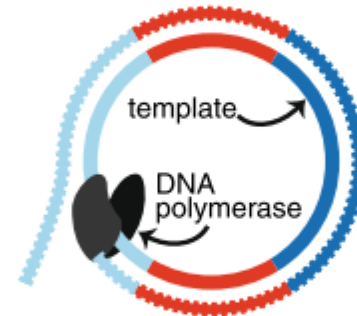
1. generate amplicon

5' forward strand 3'  
3' reverse strand 5'

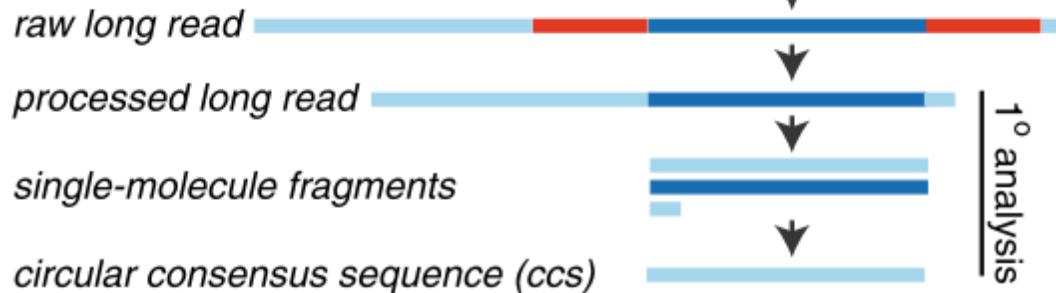
2. ligate adaptors



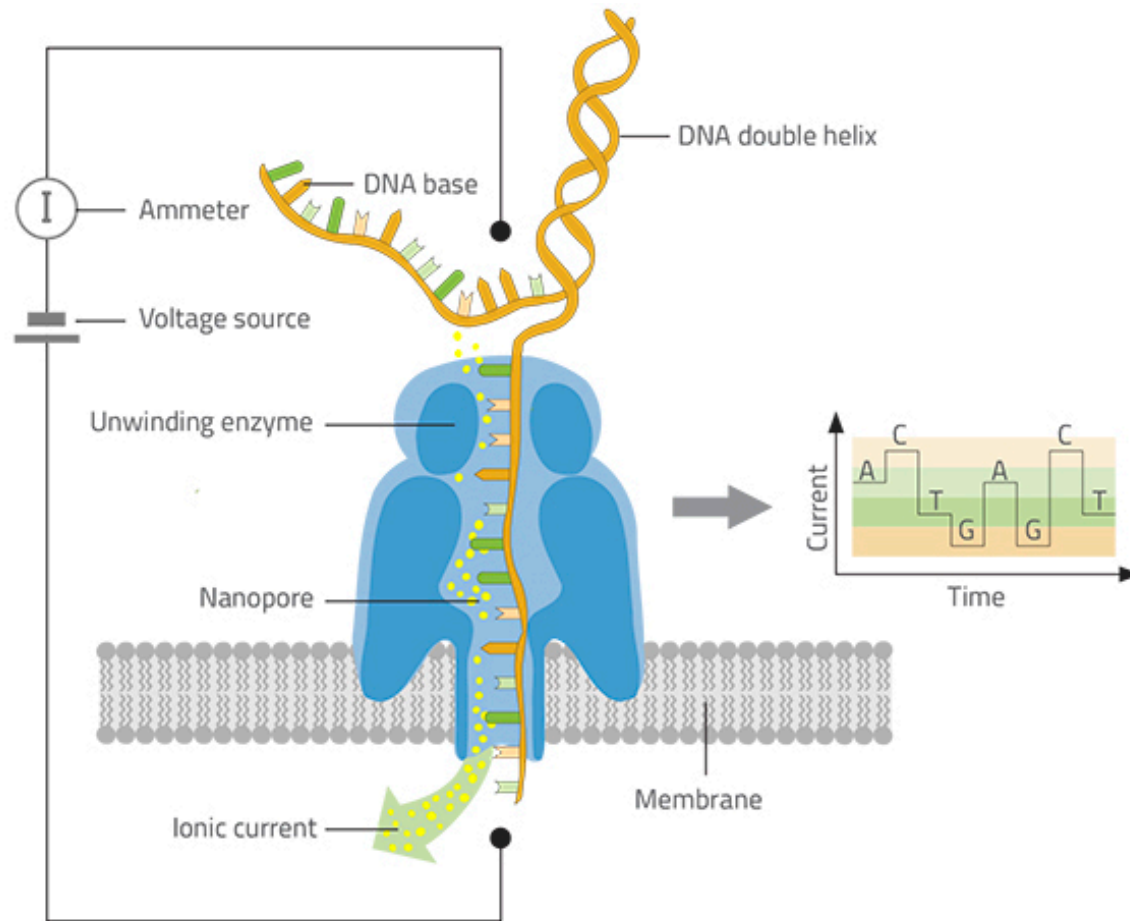
3. sequence



4. data analysis

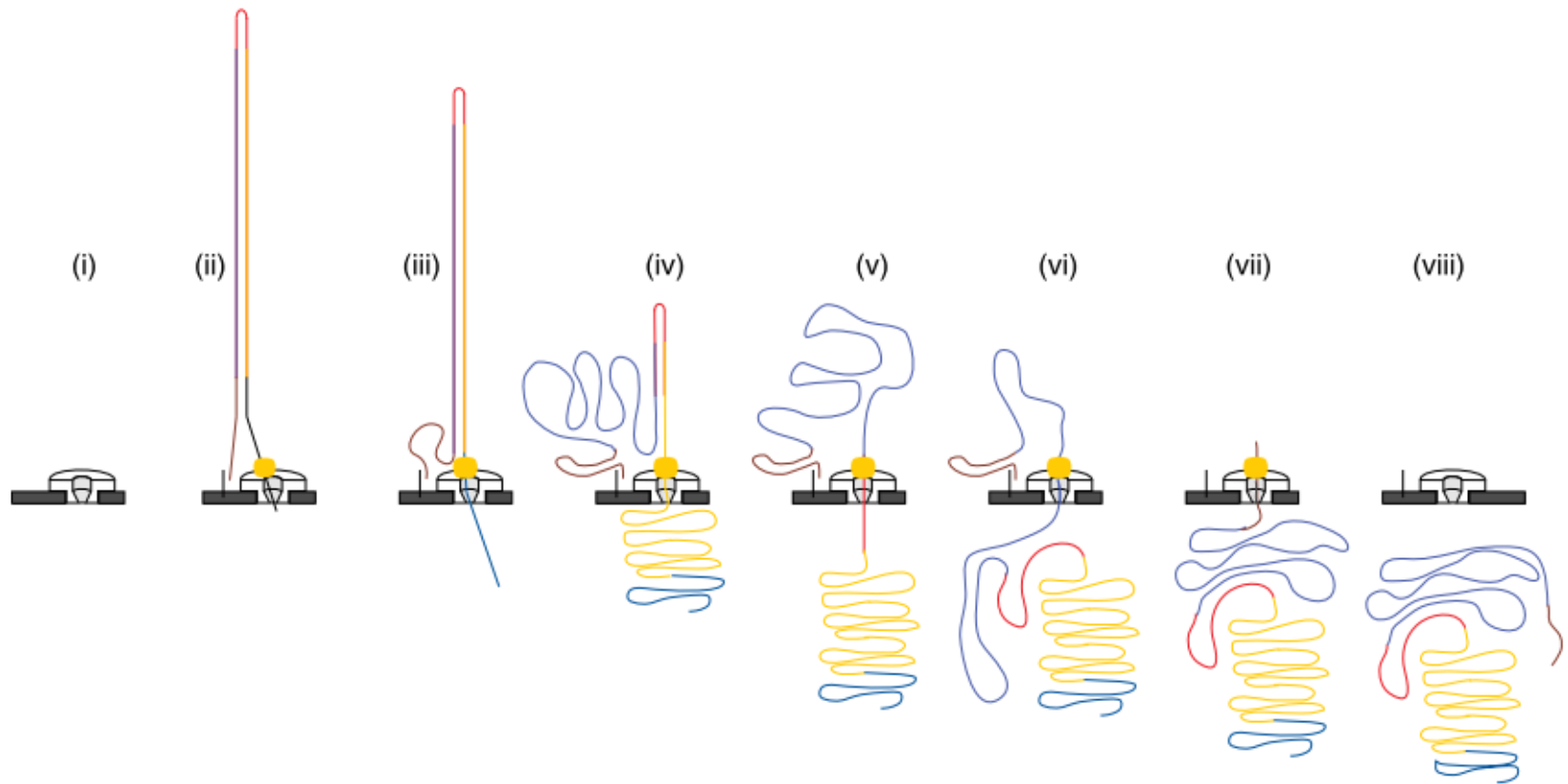


# Nanopore

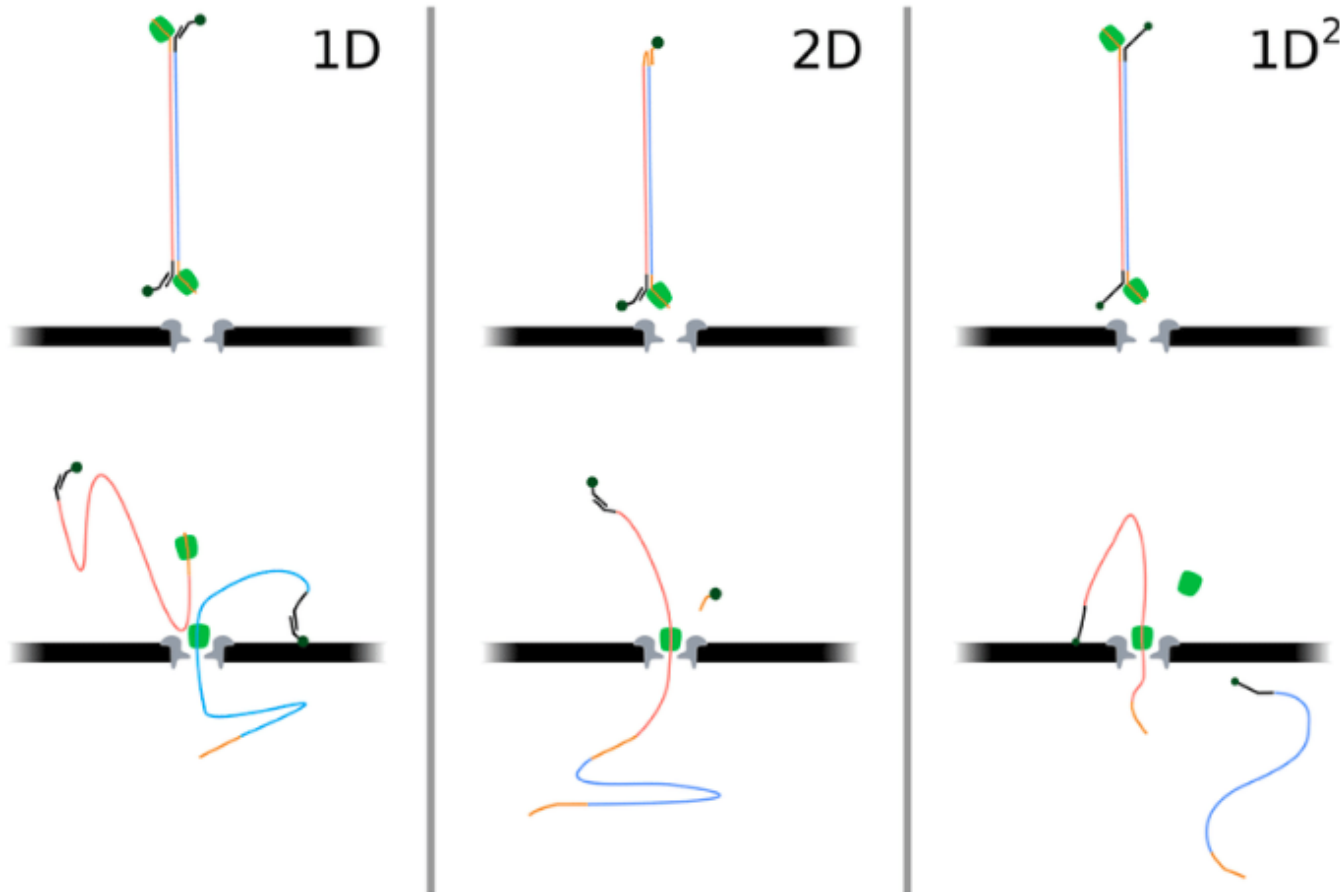


<https://www.scienceinschool.org/content/decoding-dna-pocket-sized-sequencer>

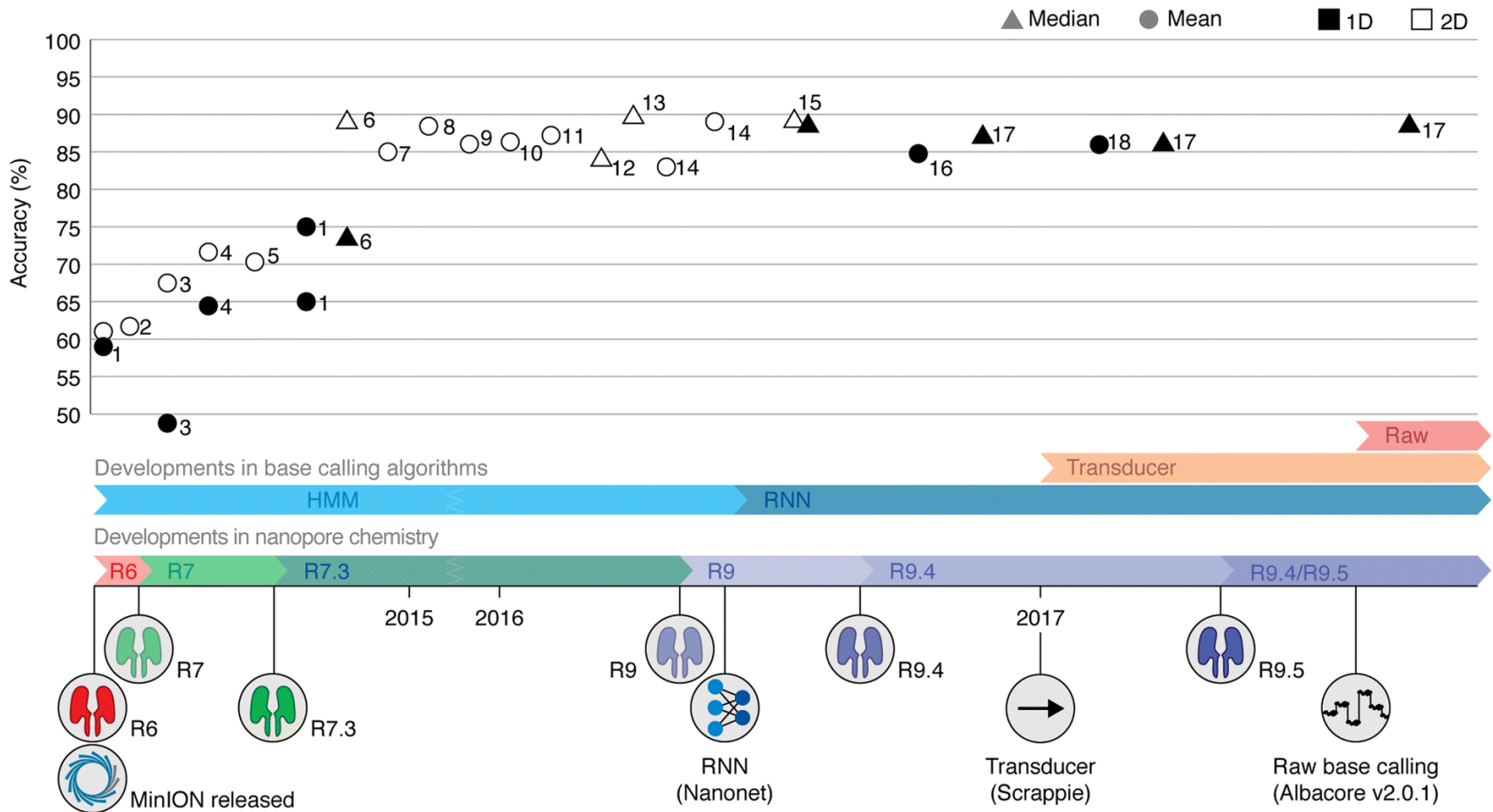
# Steps in Nanopore



# Strategy to improve sequencing accuracy



# Accuracy of Nanopore

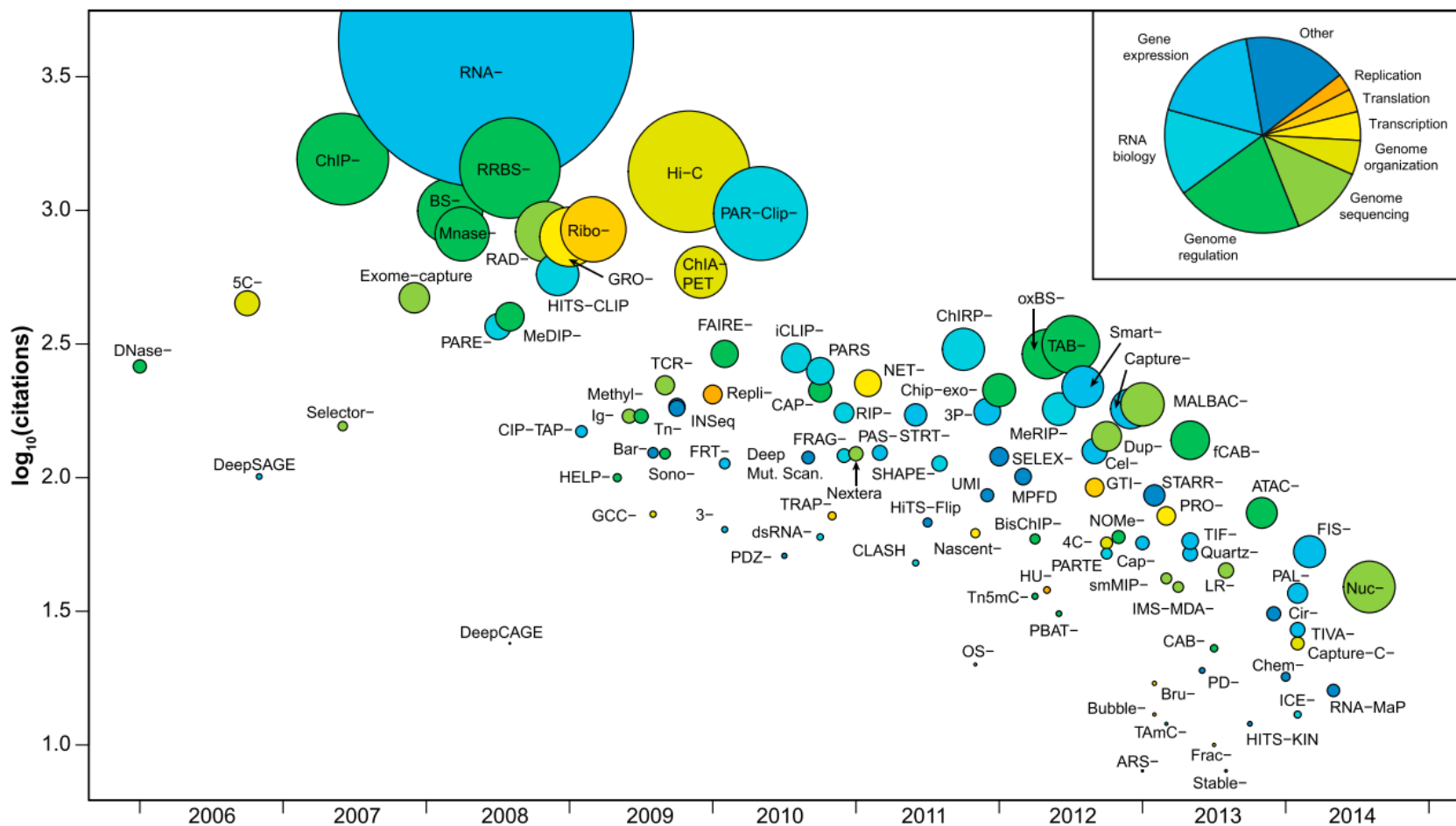


*Nature biotechnology*, 2008

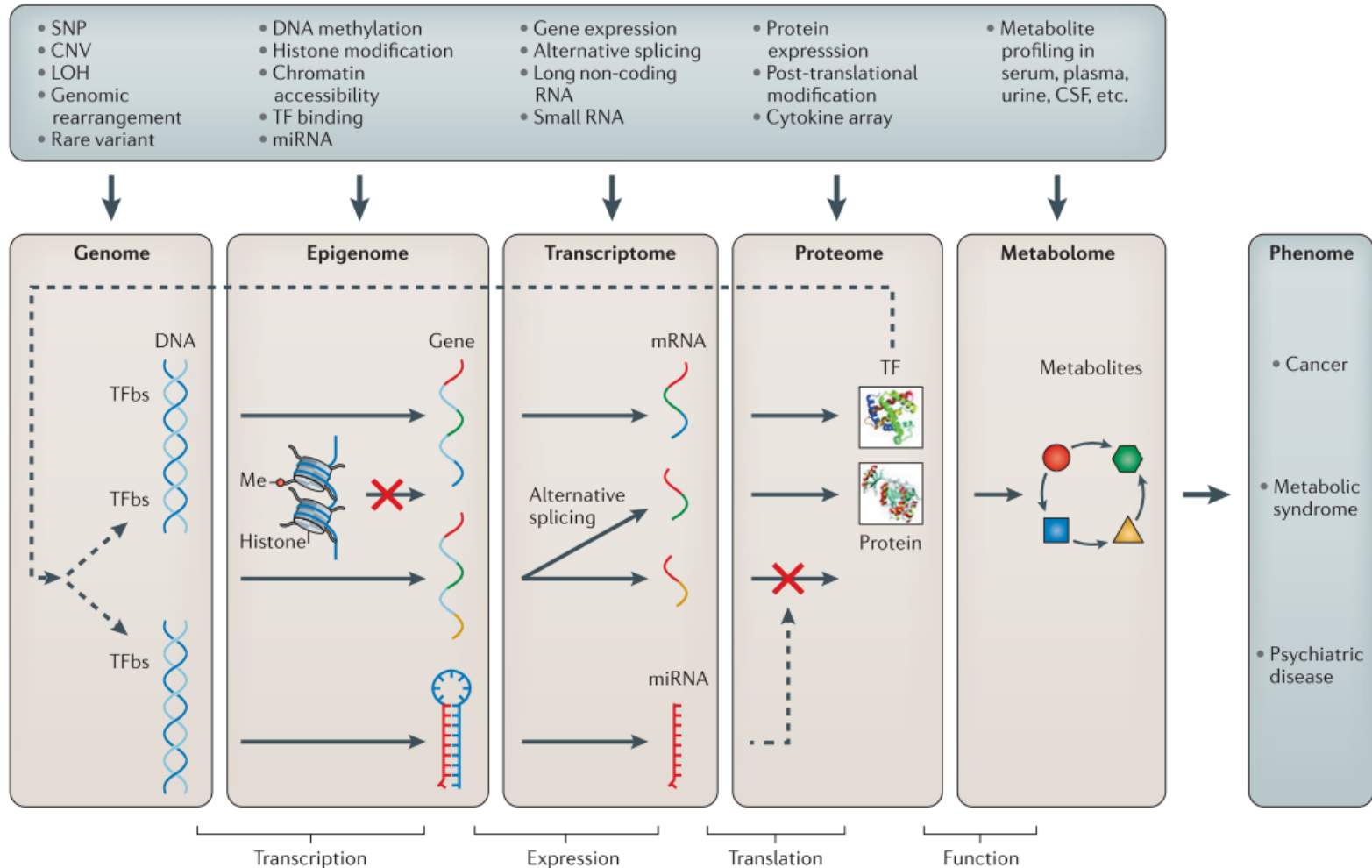
# Comparison of platform between 2<sup>nd</sup> and 3<sup>rd</sup>

Sequencing platform		Total output (bases per run)	Total reads (million per run)	Read length (bases)	Run time (days)	Purpose/definition
Illumina	HiSeq X	1.6–1.8 Tb	6000 M	2 × 150 bp	<3	Allows sequencing of larger genomes (e.g., mammalian genomes) at population level
	MiSeq	300 Mb–15 Gb	50 M	2 × 300 bp	0.2–2.7	Designed for particularly small genomes (e.g., bacterial genomes) and amplicon sequencing
Life technologies	Solid 5500 Systems	80 Gb–320 Gb	1200 M–2400 M	50–2 × 50 bp	7	Offers application-per-lane sequencing that allows transcriptome, exome and genome sequencing concurrently in a single run. Additionally, pay-per-lane sequencing feature makes Solid 5500 Systems cost-effective because reagents are not required for unused lanes.
	Ion Torrent 520 Chip	600 Mb–2 Gb	3–5 M	200–400 bp	0.1	Ion S5 System allows generation of diverse sequencing data ranging from targeted re-sequencing to genome sequencing with as little as 10 ng sample.
	Ion Torrent 540 Chip	10–15 Gb	60–80 M	200–400 bp	0.1	
PacBio	Sequel System	500 Mb–16 Gb	55–880 M	up to 60 kb	<0.1–0.3	Useful in the studies of <i>de novo</i> assembly of large genomes. Sequel System can be utilized for generating variation, expression and/or regulation related sequencing data.
	PacBio RS II	500 Mb–16 Gb	55–880 M	up to 60 kb	<0.1–0.3	Much more suitable for sequencing small genomes although animal and plant genomic studies is also possible.
Nanopore	PromethION	up to 12 Tb <sup>a</sup>	1250 M <sup>a</sup>	230–300 kb <sup>a</sup>	2	Ideal for large sample numbers. PromethION can sequence up to 48 samples in a single run
	MinION	up to 42 Gb <sup>a</sup>	up to 4.4 M <sup>a</sup>	230–300 kb <sup>a</sup>	2	Portable sequencing instrument. MinION can be run with a desktop or laptop computer and data can be performed in real time.

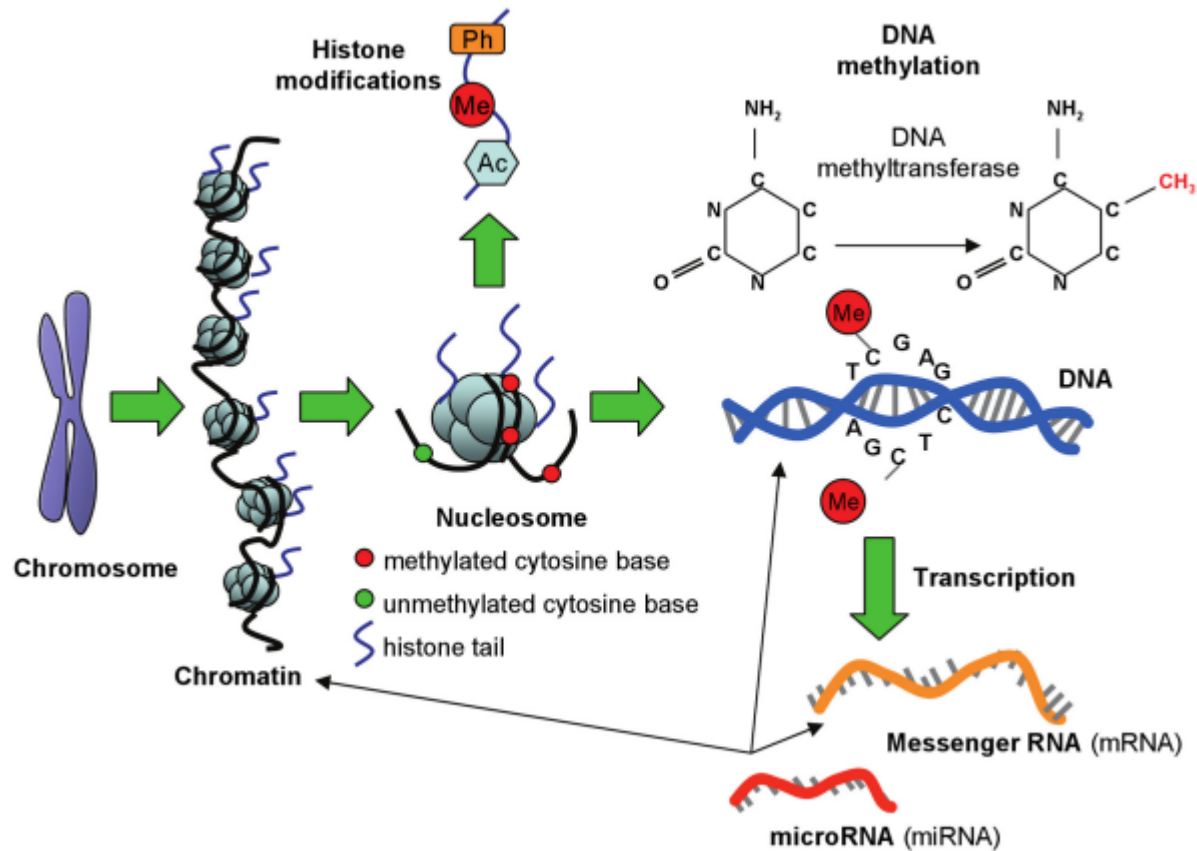
# Application of NGS



# Biological layer

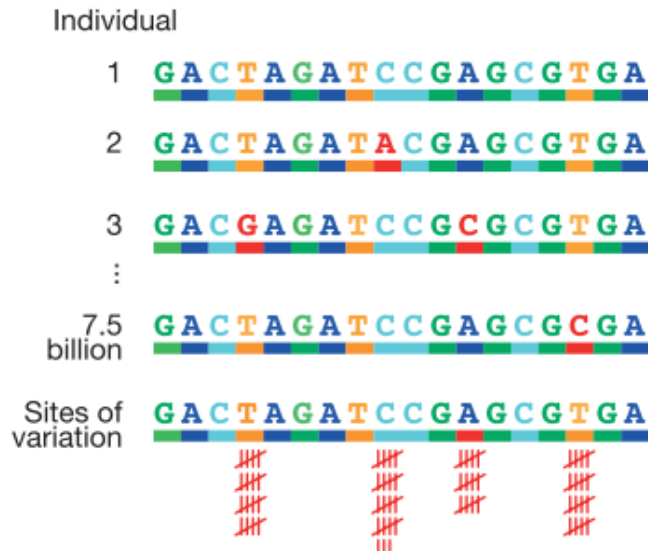


# Central dogma with epigenome

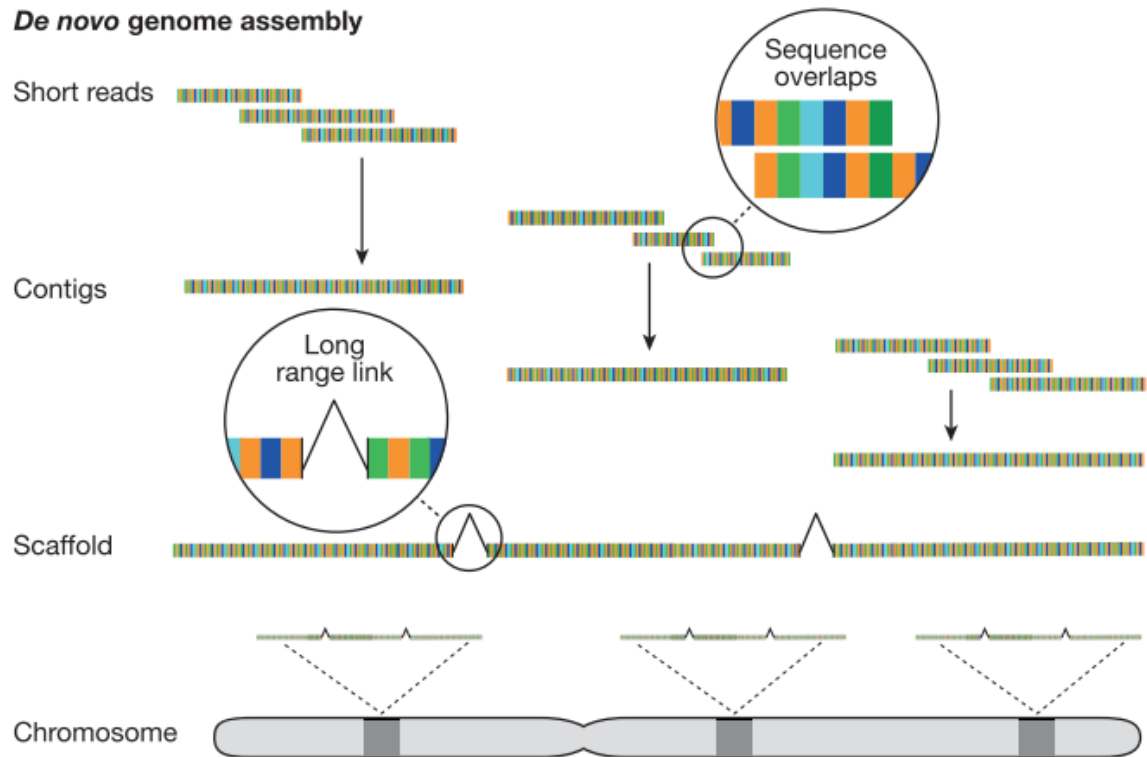


# Sequencing using whole genome

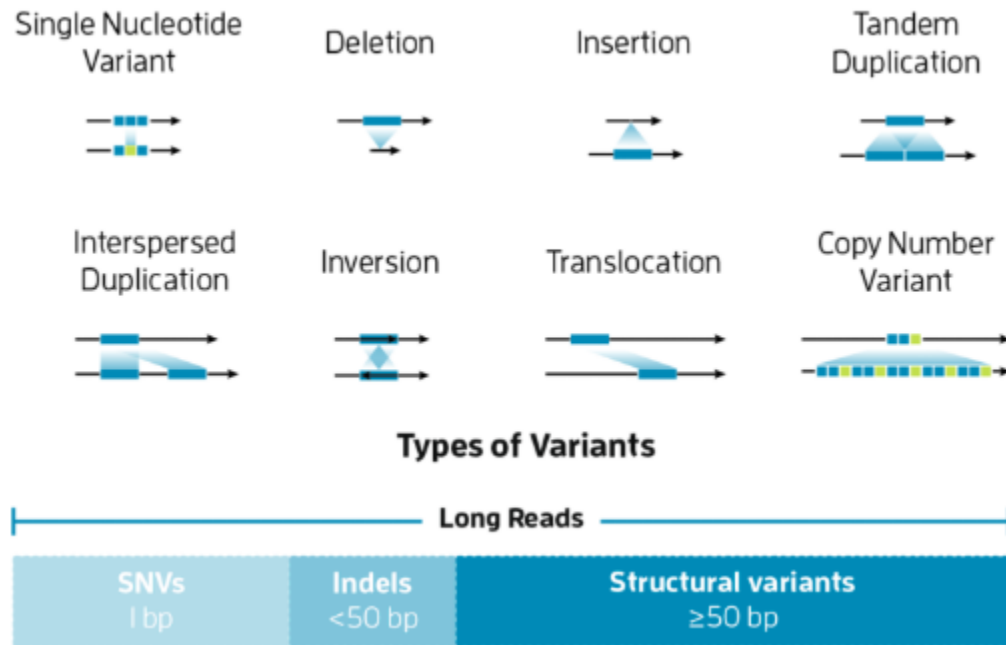
## Genome resequencing



## De novo genome assembly



# Type of variants



<https://www.pacb.com/applications/whole-genome-sequencing/variant-detection/>

# The Angelina effect

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# BRCA1 and BRCA2

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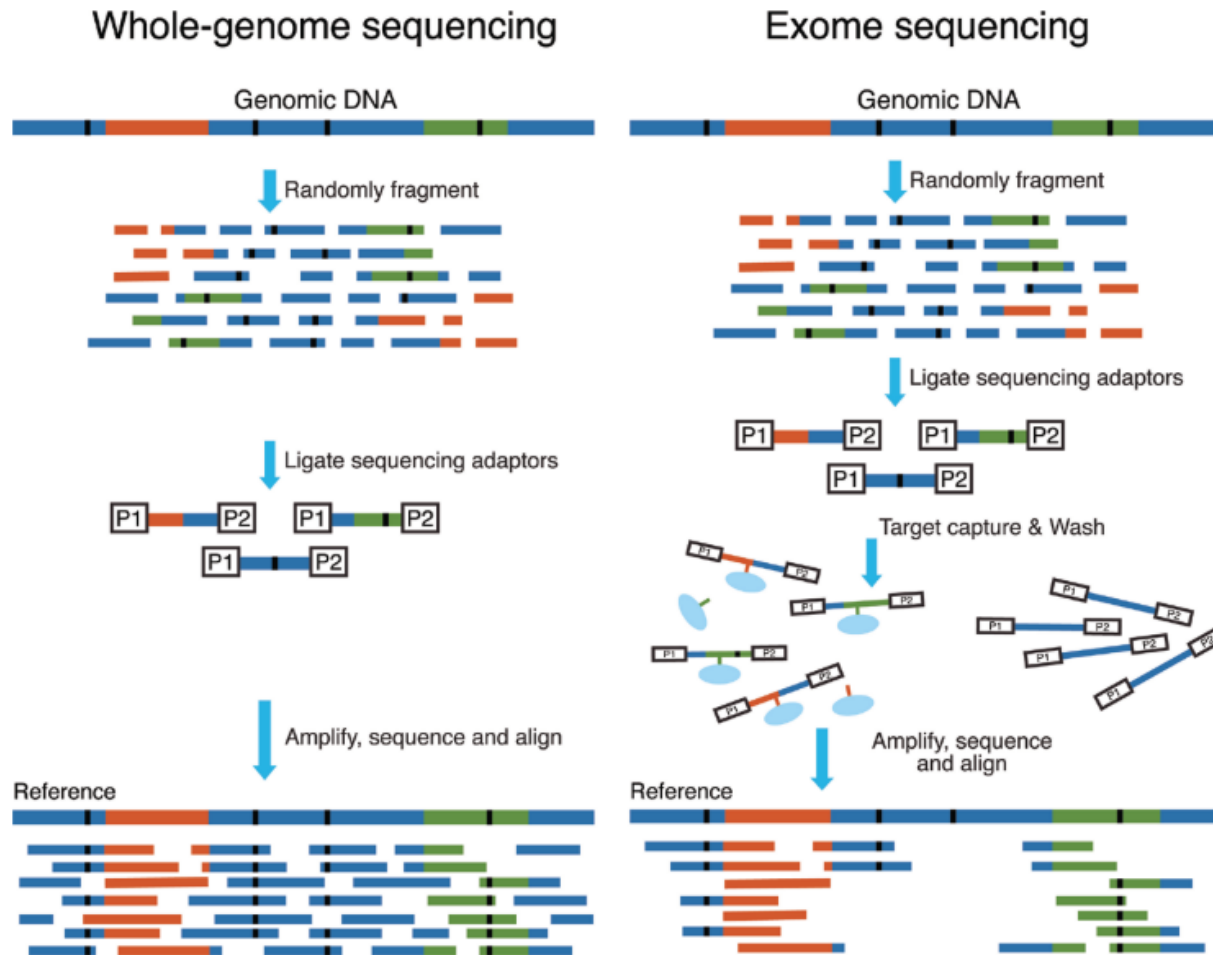
*BRCA1* and *BRCA2* are human genes that produce tumor suppressor proteins. These proteins help repair damaged DNA and, therefore, play a role in ensuring the stability of each cell's genetic material. When either of these genes is mutated, or altered, such that its protein product is not made or does not function correctly, DNA damage may not be repaired properly. As a result, cells are more likely to develop additional genetic alterations that can lead to cancer.

Specific inherited mutations in *BRCA1* and *BRCA2* most notably increase the risk of female breast and ovarian cancers, but they have also been associated with increased risks of several additional types of cancer. People who have inherited mutations in *BRCA1* and *BRCA2* tend to develop breast and ovarian cancers at younger ages than people who do not have these mutations.

**Breast cancer:** About 12% of women in the general population will develop breast cancer sometime during their lives (1). By contrast, a recent large study estimated that about 72% of women who inherit a harmful *BRCA1* mutation and about 69% of women who inherit a harmful *BRCA2* mutation will develop breast cancer by the age of 80 (2).

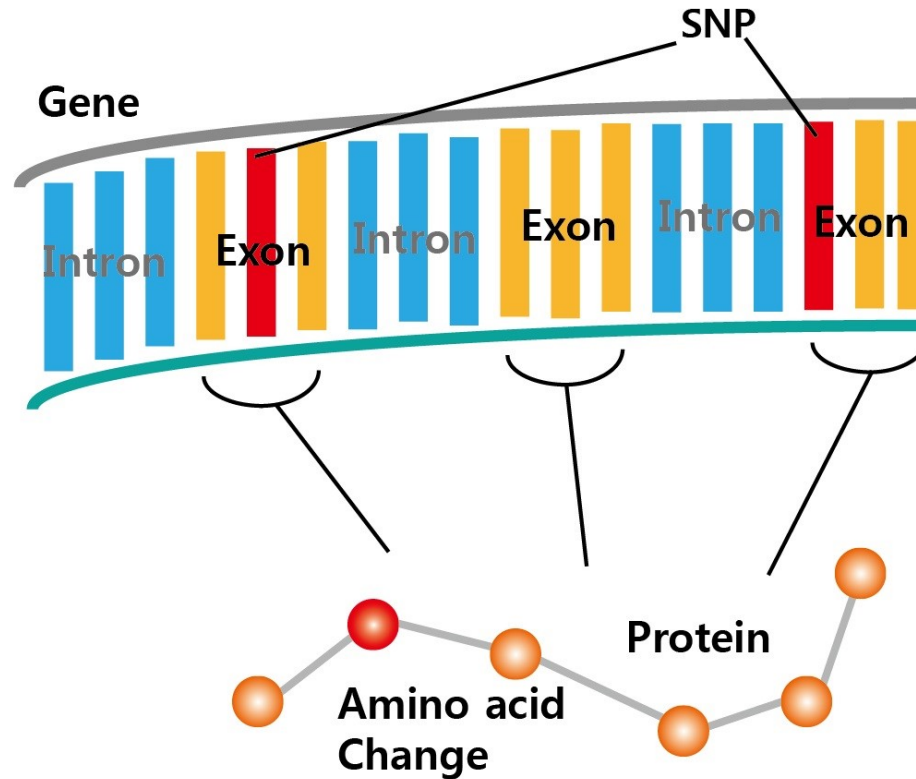
**Ovarian cancer:** About 1.3% of women in the general population will develop ovarian cancer sometime during their lives (1). By contrast, it is estimated that about 44% of women who inherit a harmful *BRCA1* mutation and about 17% of women who inherit a harmful *BRCA2* mutation will develop ovarian cancer by the age of 80 (2).

# Sequencing for resequencing



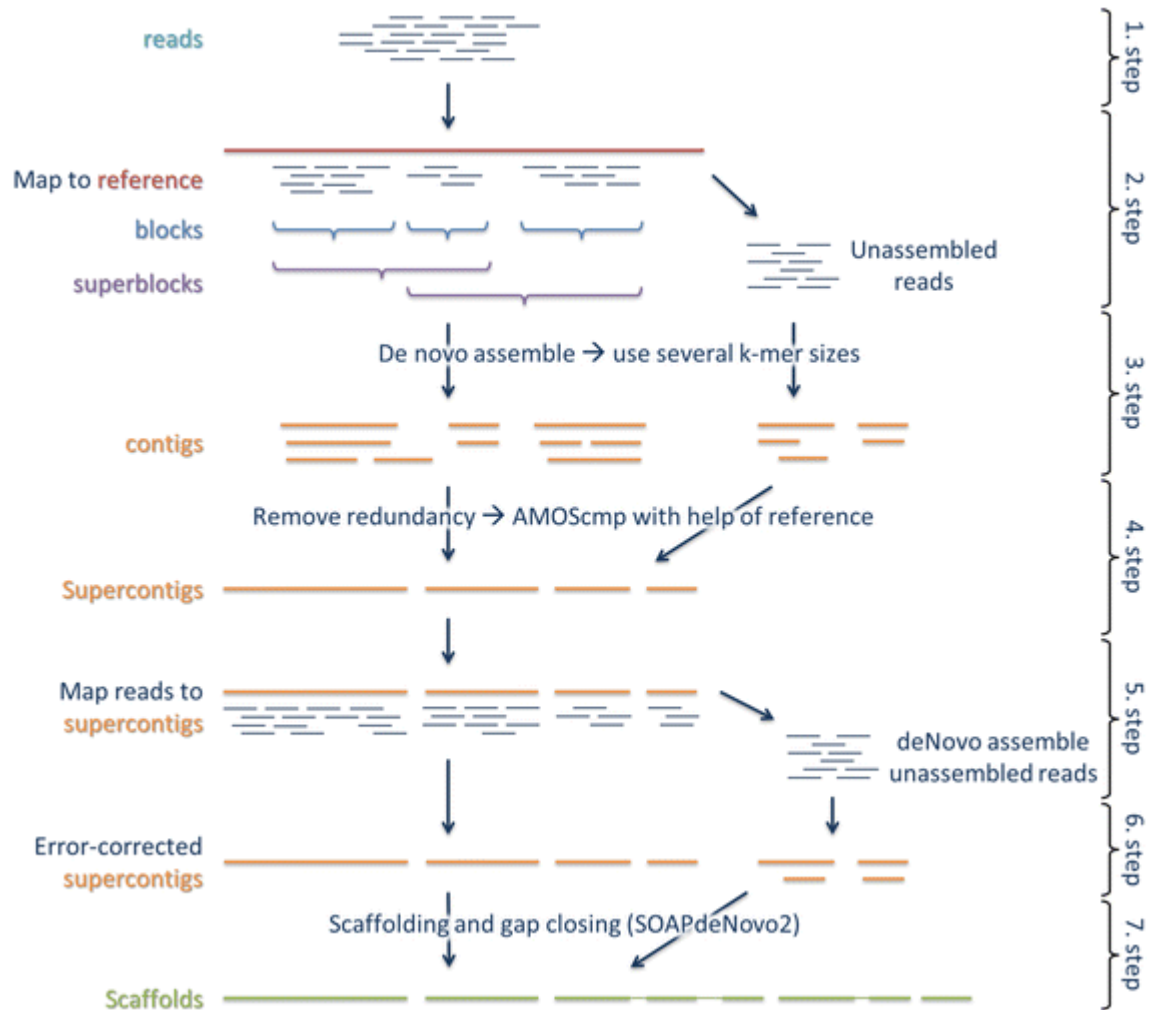
[https://ngxbio.com/pub/media/magefan\\_blog/WES1.jpg](https://ngxbio.com/pub/media/magefan_blog/WES1.jpg)

# Whole Exome Sequencing



[https://ngxbio.com/pub/media/magefan\\_blog/WES1.jpg](https://ngxbio.com/pub/media/magefan_blog/WES1.jpg)

# Steps of De novo genome assembly

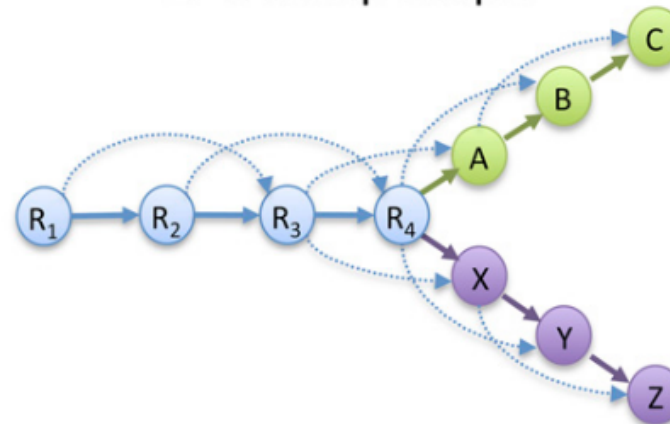


# Genome assembly algorithms

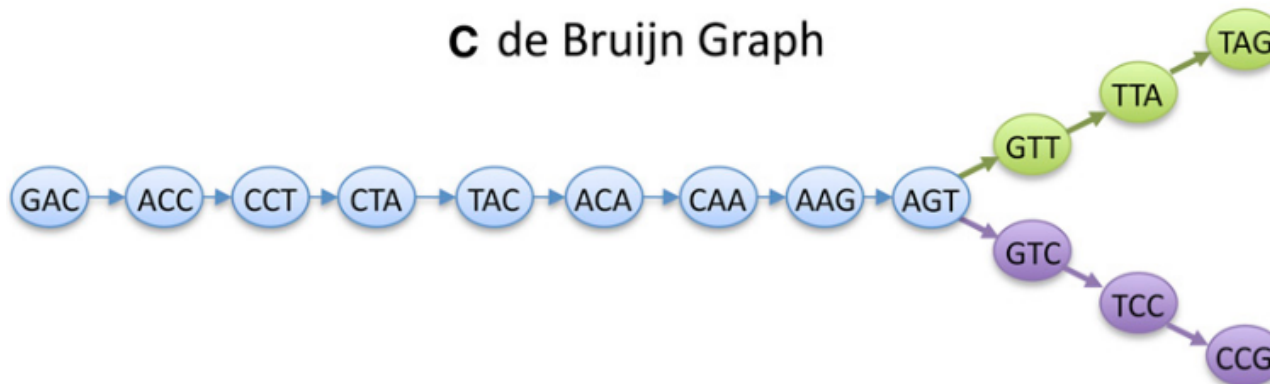
**A Read Layout**

R<sub>1</sub>: GACCTACA  
R<sub>2</sub>: ACCTACAA  
R<sub>3</sub>: CCTACAAG  
R<sub>4</sub>: CTACAAGT  
A: TACAAGTT  
B: ACAAGTTA  
C: CAAGTTAG  
X: TACAAGTC  
Y: ACAAGTCC  
Z: CAAGTCCG

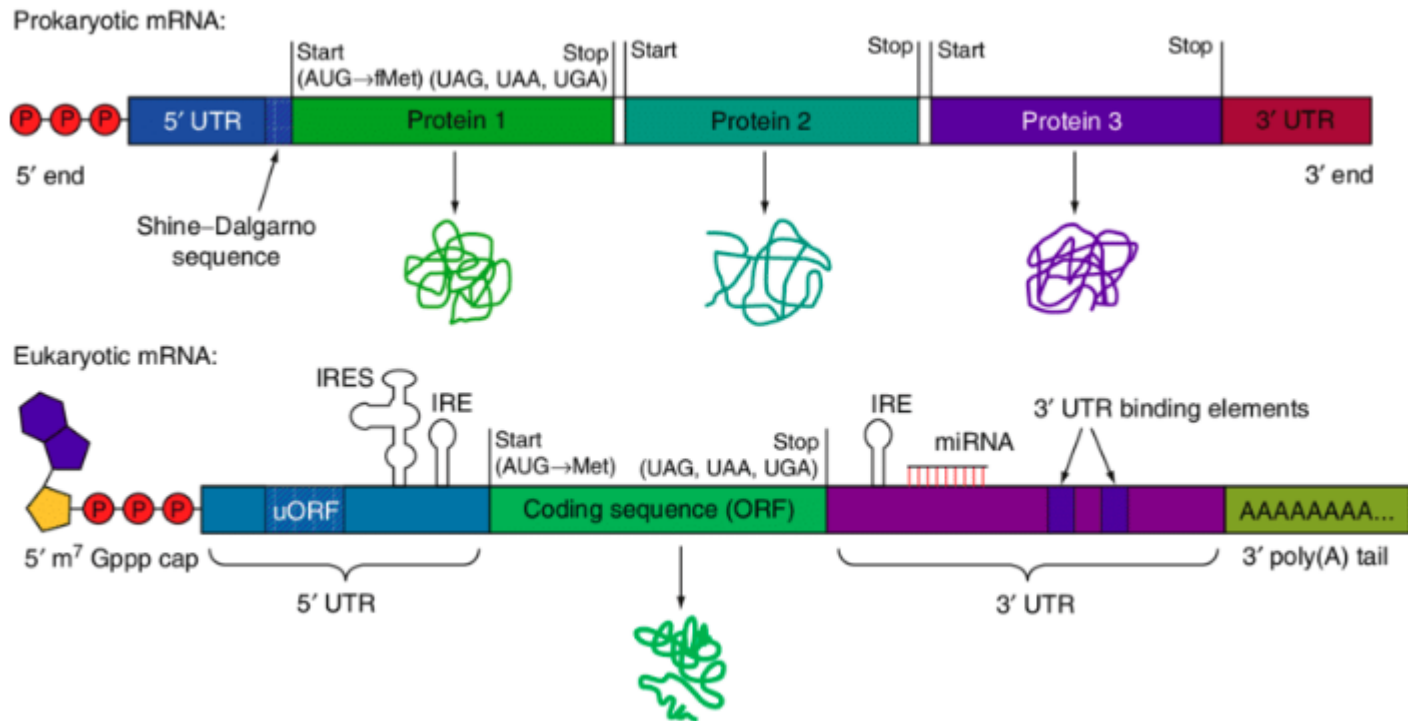
**B Overlap Graph**



**C de Bruijn Graph**

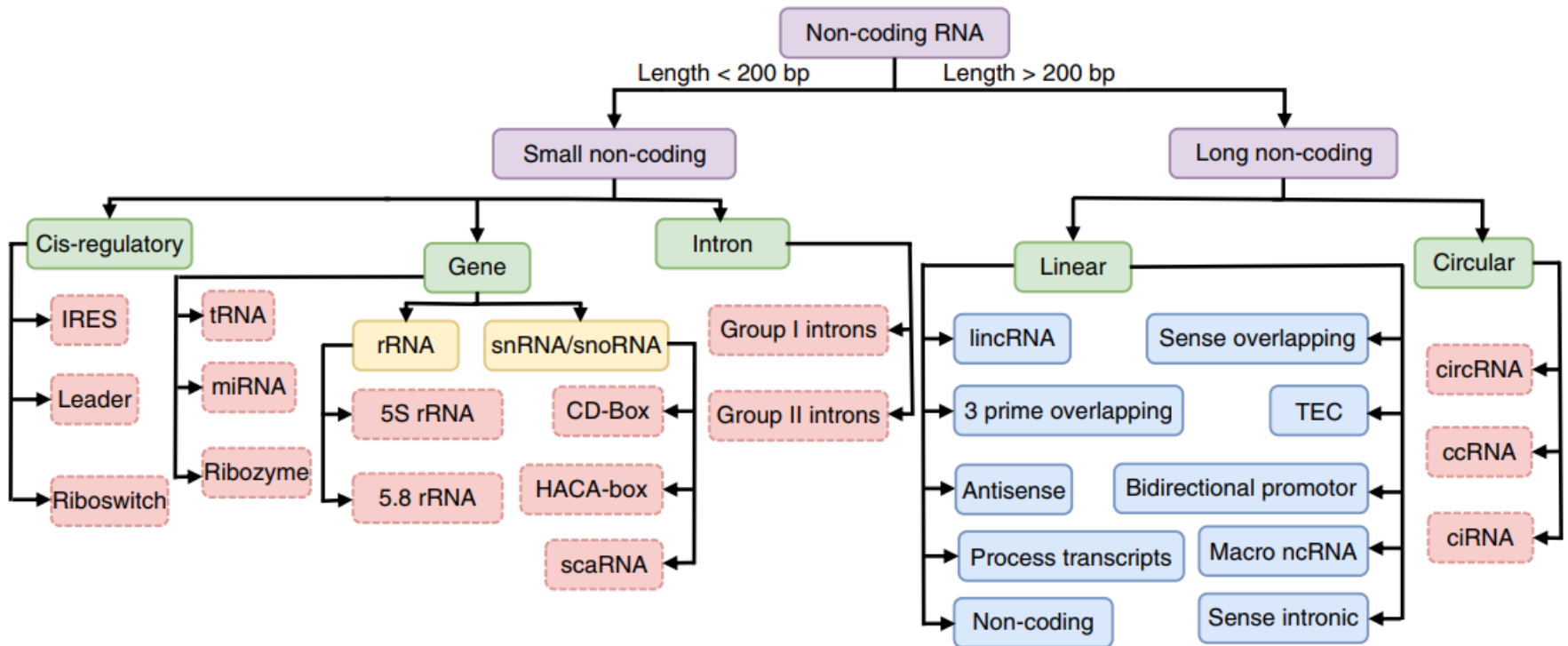


# Messenger RNA

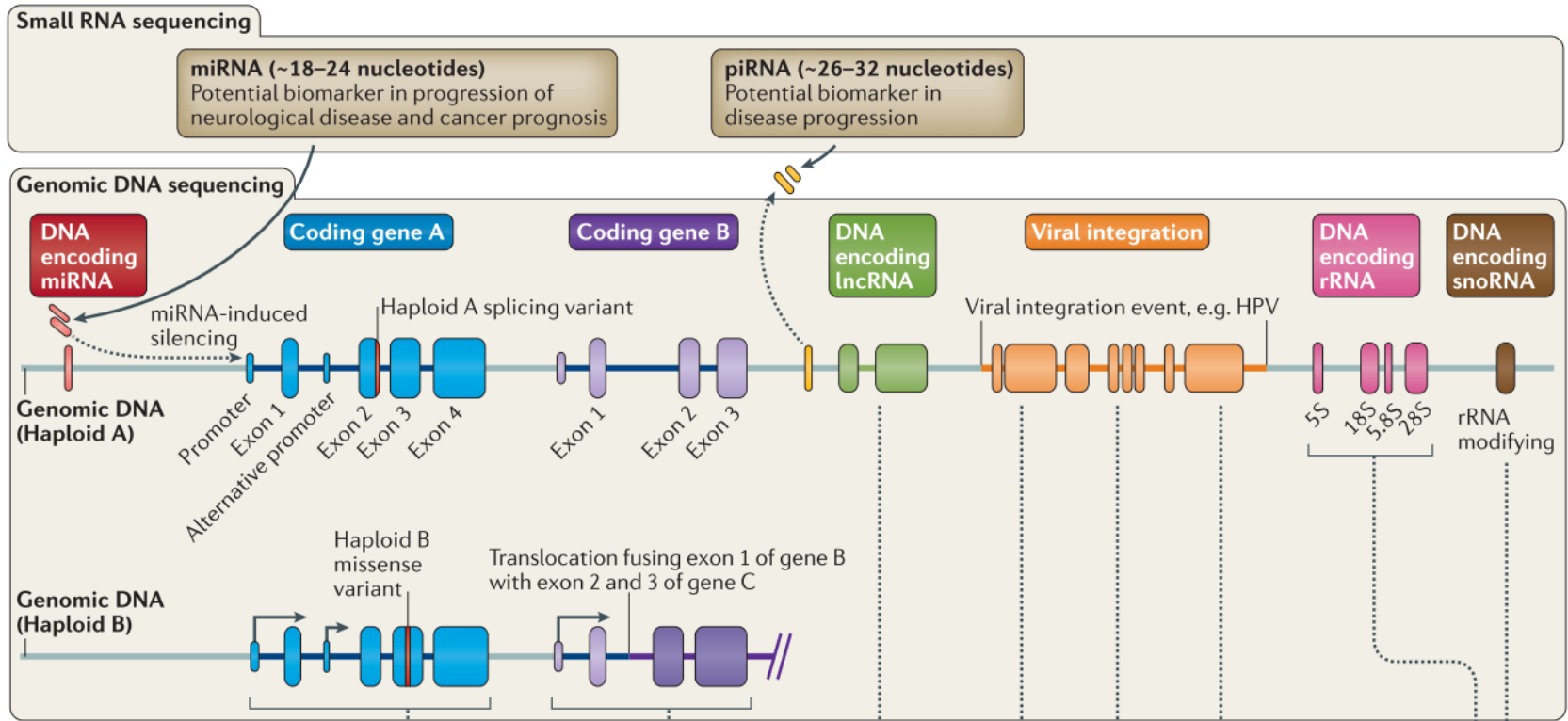


[https://www.researchgate.net/figure/Schematic-diagram-of-prokaryotic-top-and-eukaryotic-bottom-mRNA-Bars-indicate-the\\_fig1\\_305698915](https://www.researchgate.net/figure/Schematic-diagram-of-prokaryotic-top-and-eukaryotic-bottom-mRNA-Bars-indicate-the_fig1_305698915)

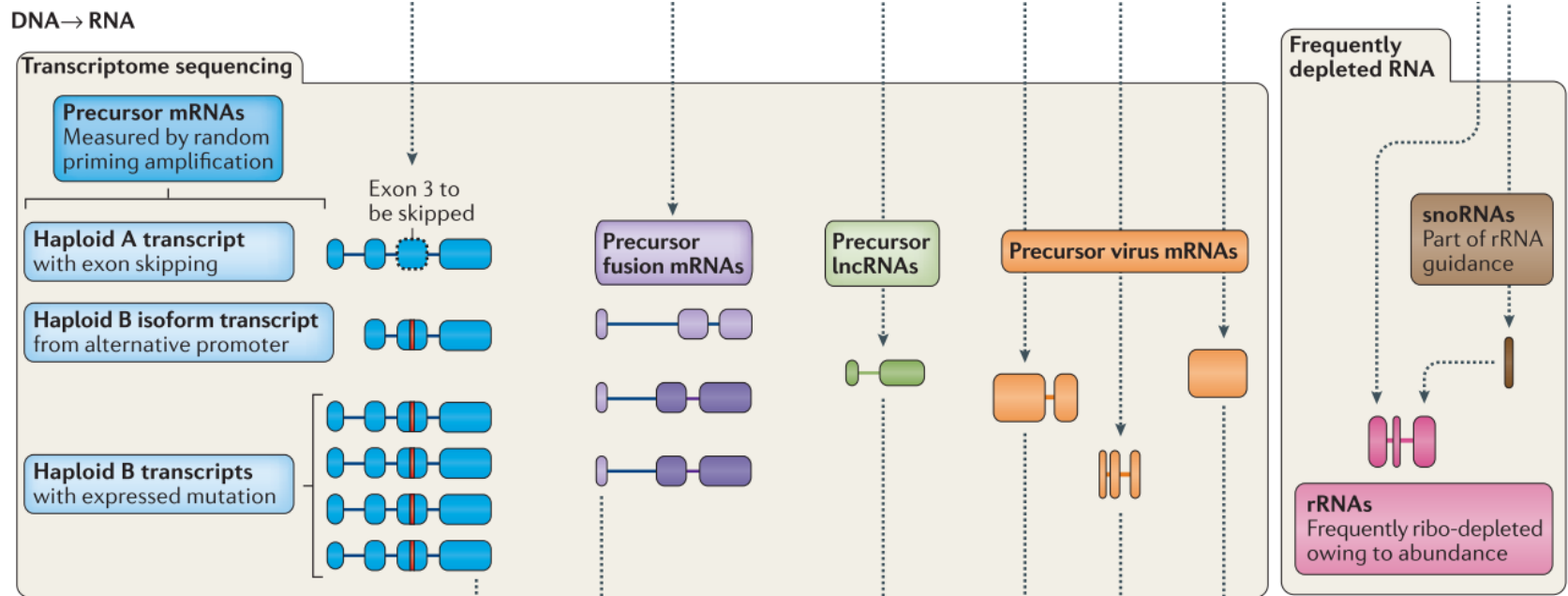
# Non-coding RNA



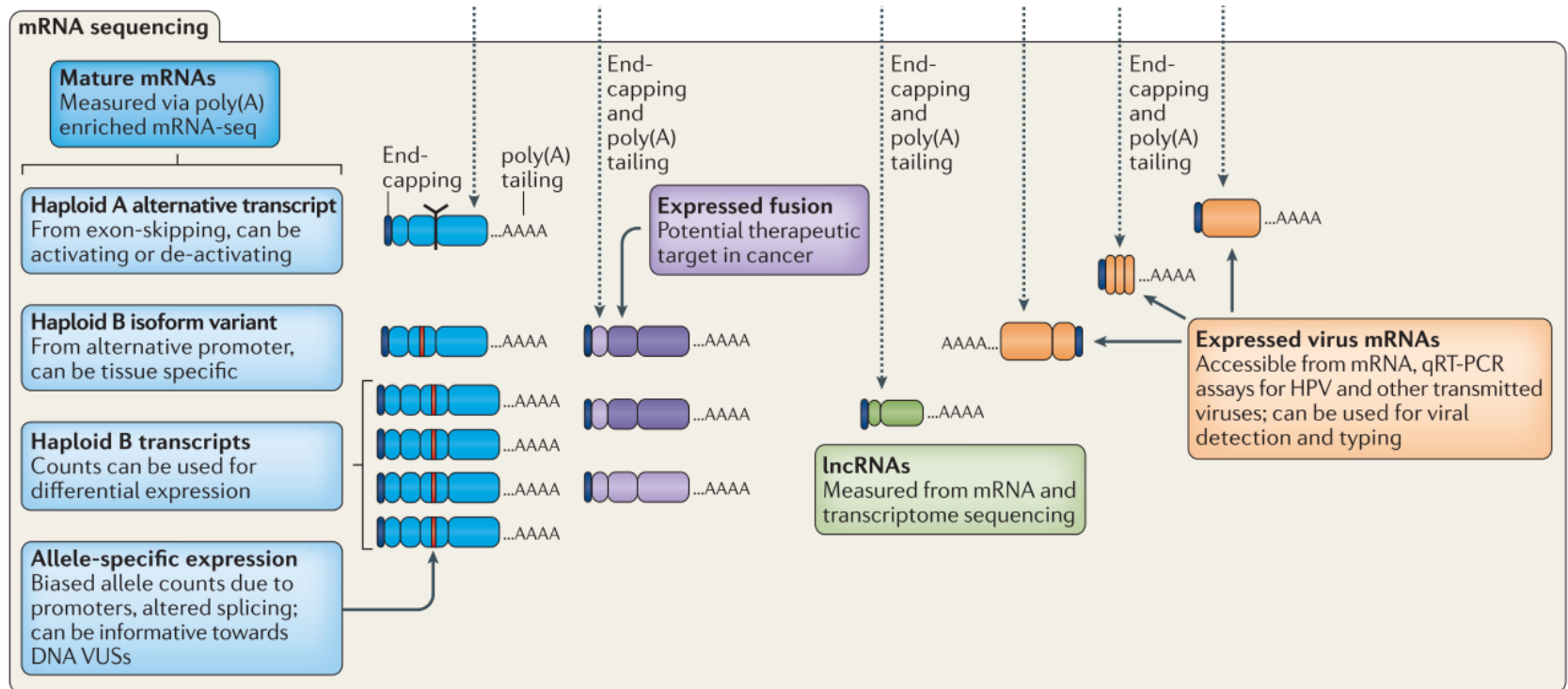
# Type of RNA-seq



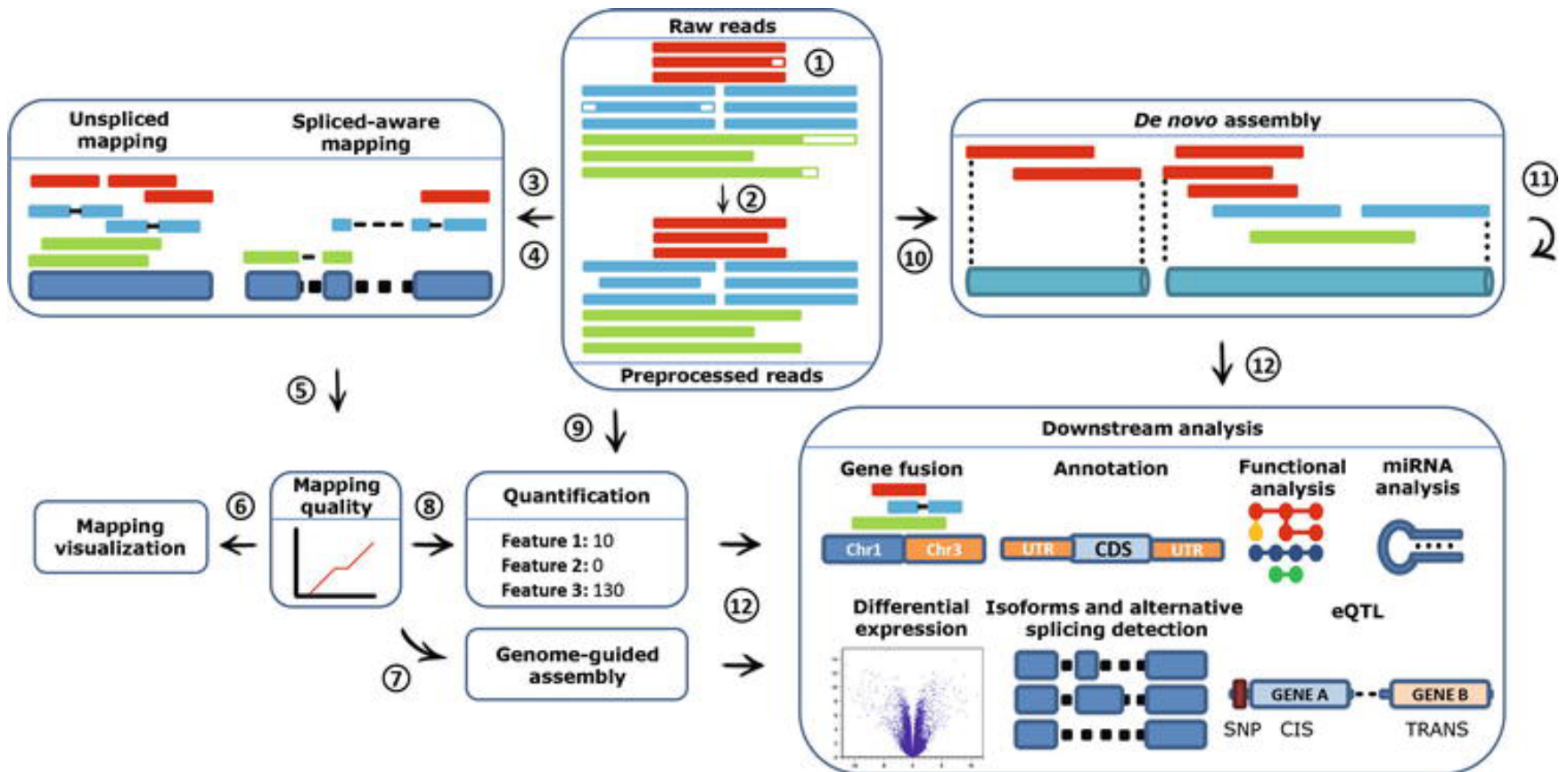
# Type of RNA-seq



# Type of RNA-seq

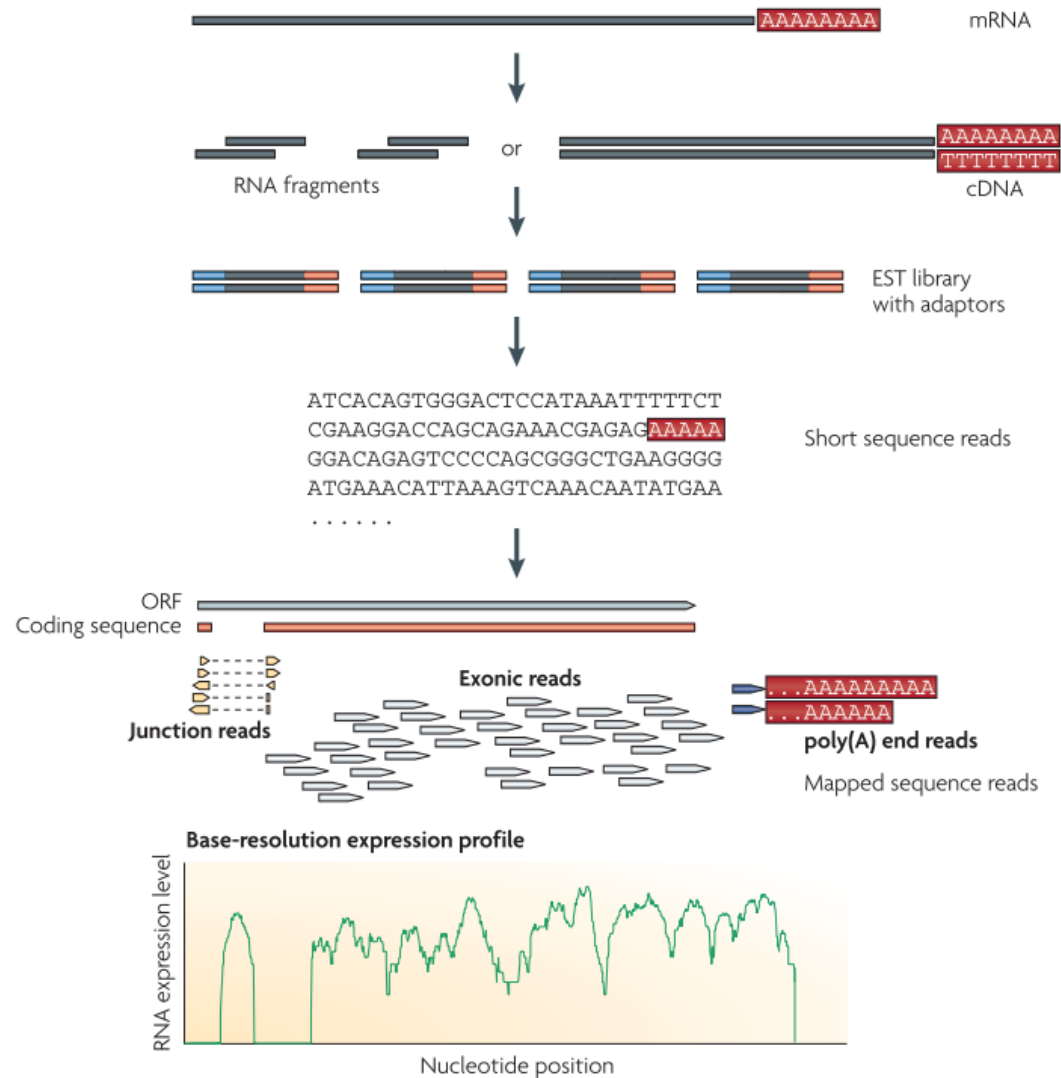


# RNA-seq data analysis



<https://www.intechopen.com/books/applications-of-rna-seq-and-omics-strategies-from-microorganisms-to-human-health/rna-seq-applications-and-best-practices>

# RNA-seq data analysis



*Nature reviews genetics, 2009*

# Nobel prize – RNA Interference

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## The Nobel Prize in Physiology or Medicine 2006

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Photo: L. Cicero

**Andrew Z. Fire**

Prize share: 1/2



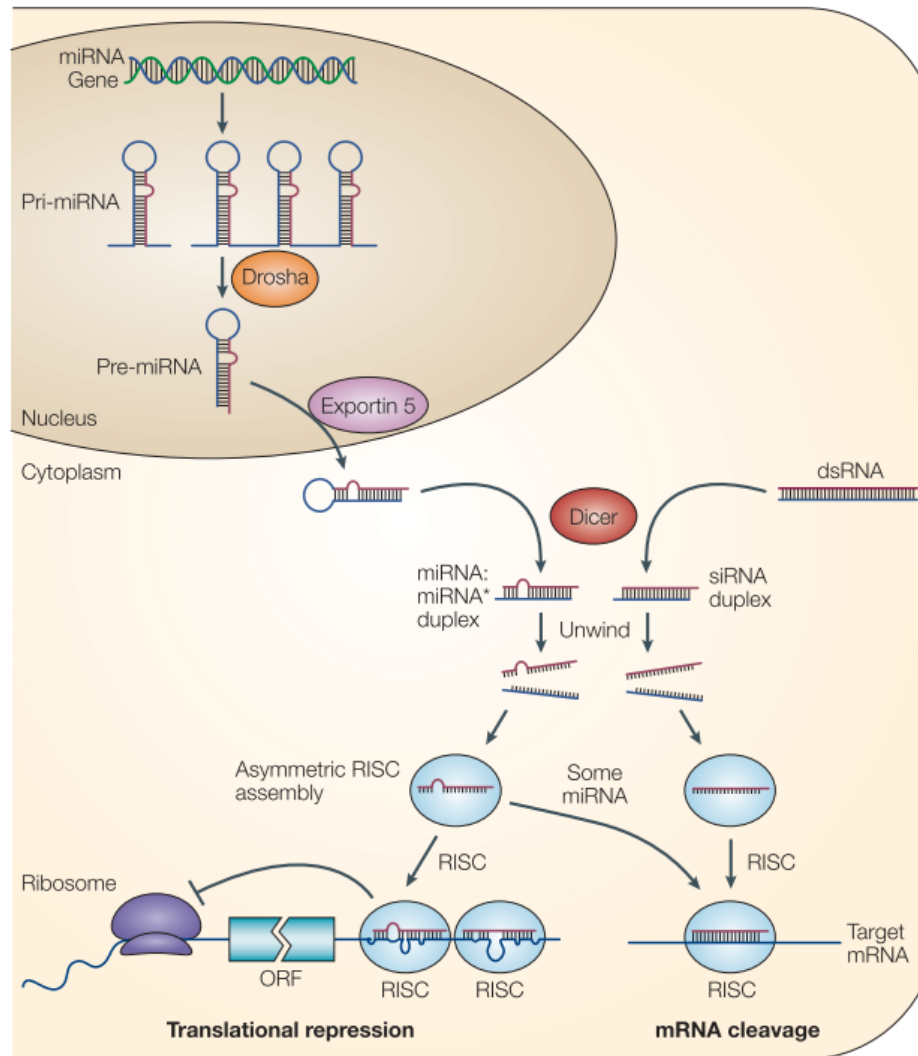
Photo: J. Mottern

**Craig C. Mello**

Prize share: 1/2

<https://www.nobelprize.org/prizes/medicine/2006/summary/>

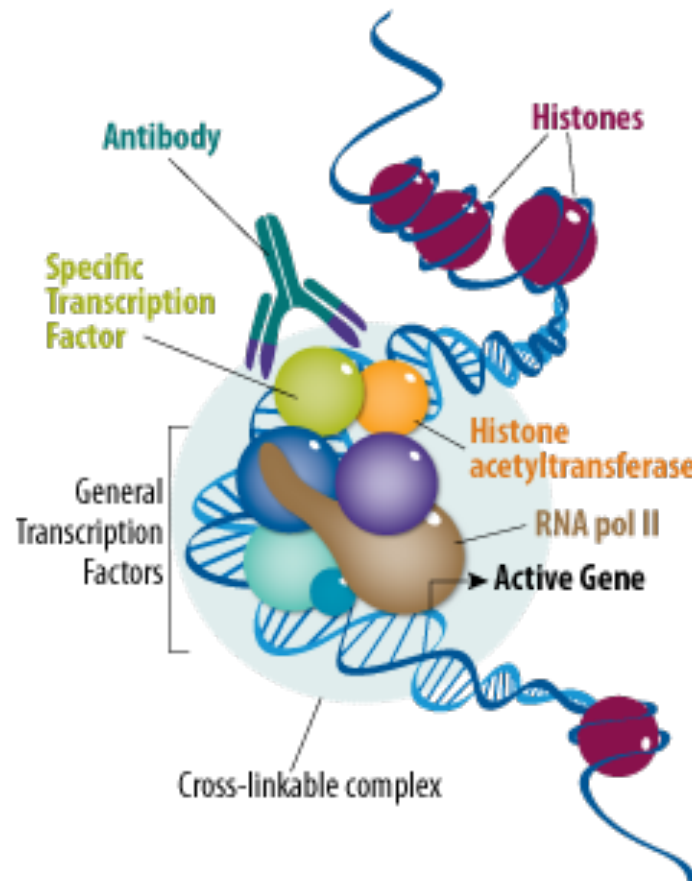
# Micro RNA biogenesis



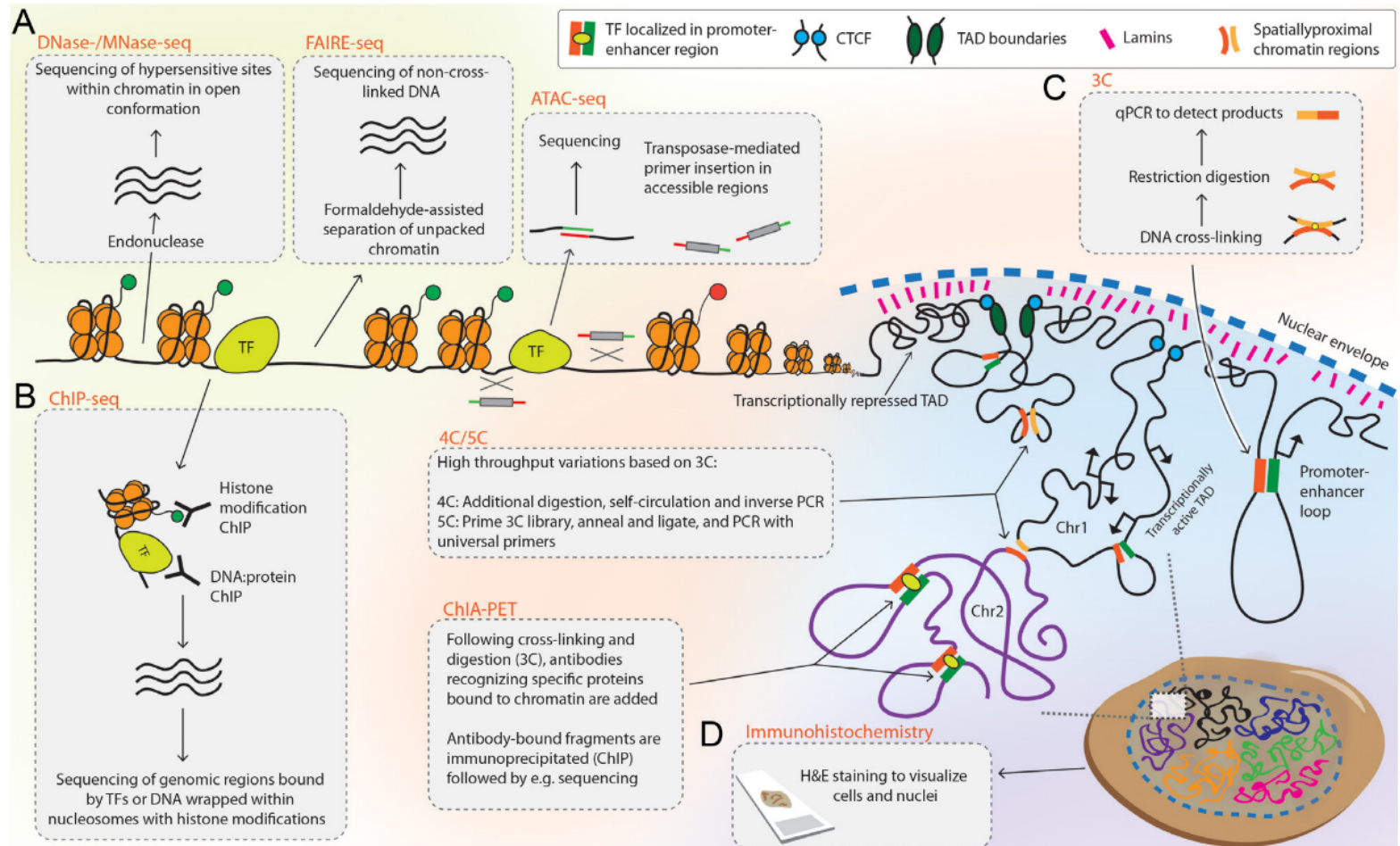
*Nature reviews Genetics, 2004*

# Chromatin and Chromatin immunoprecipitation

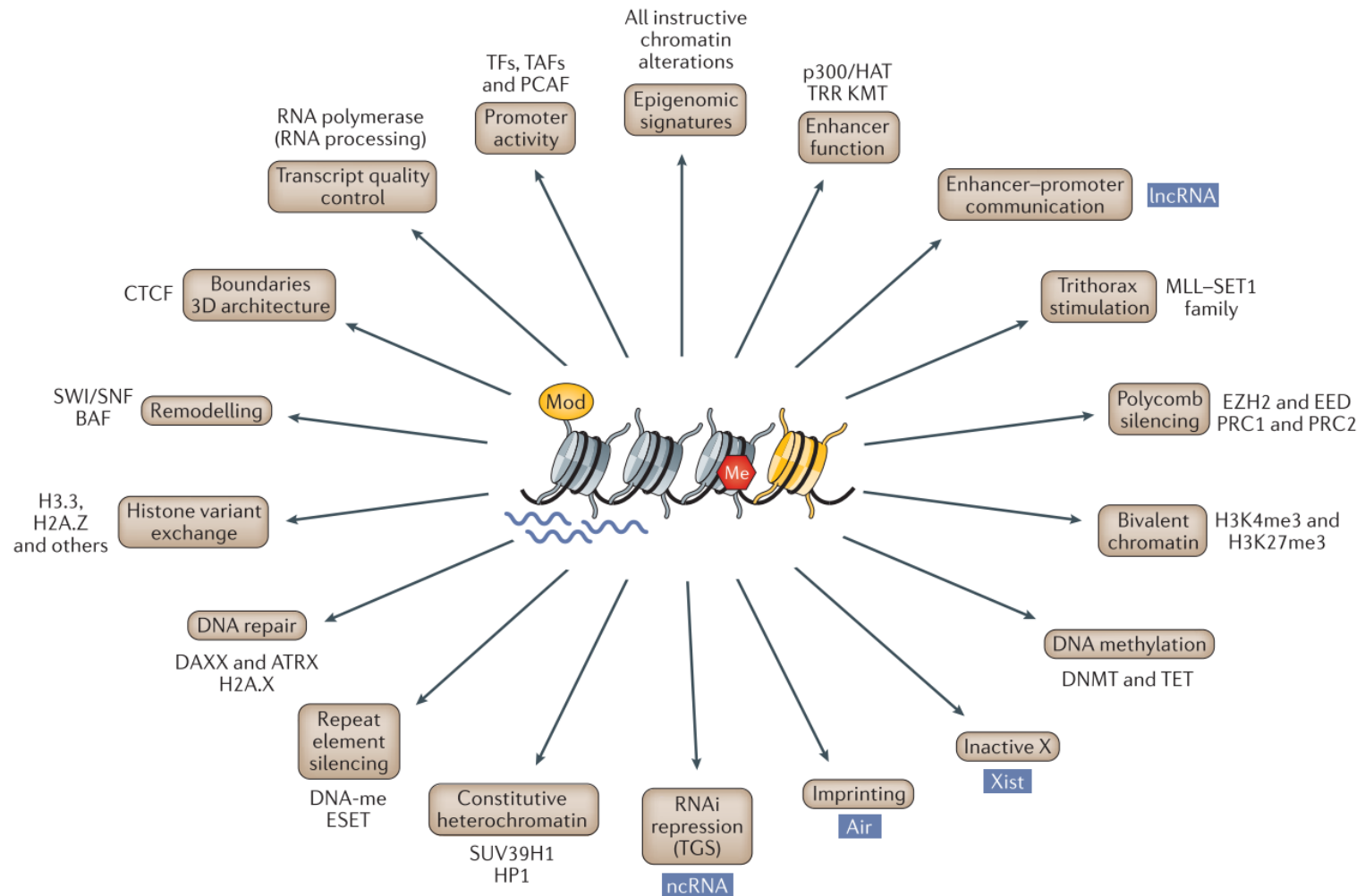
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# Sequencing Application in Epigenome

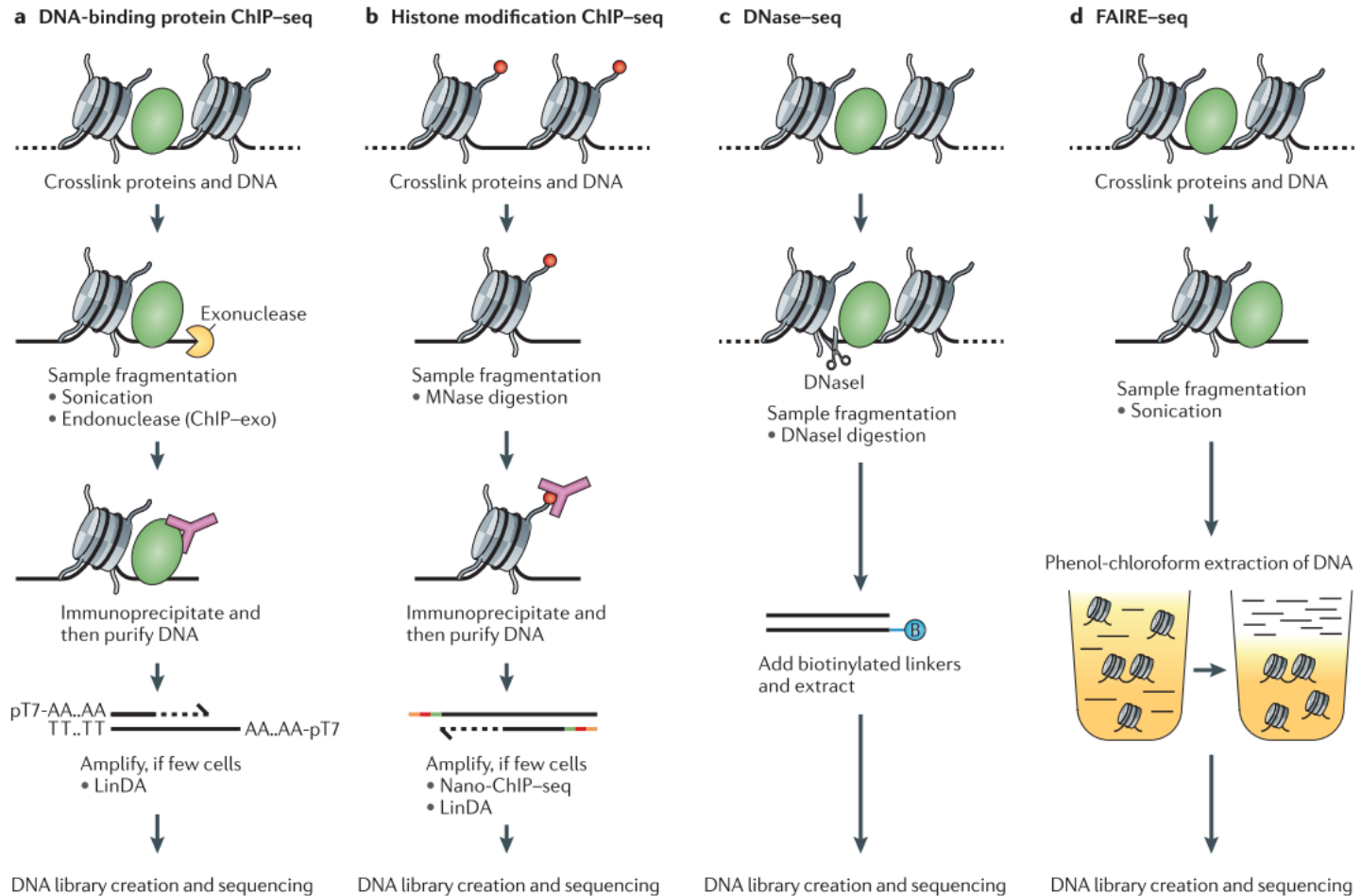


# Epigenome function

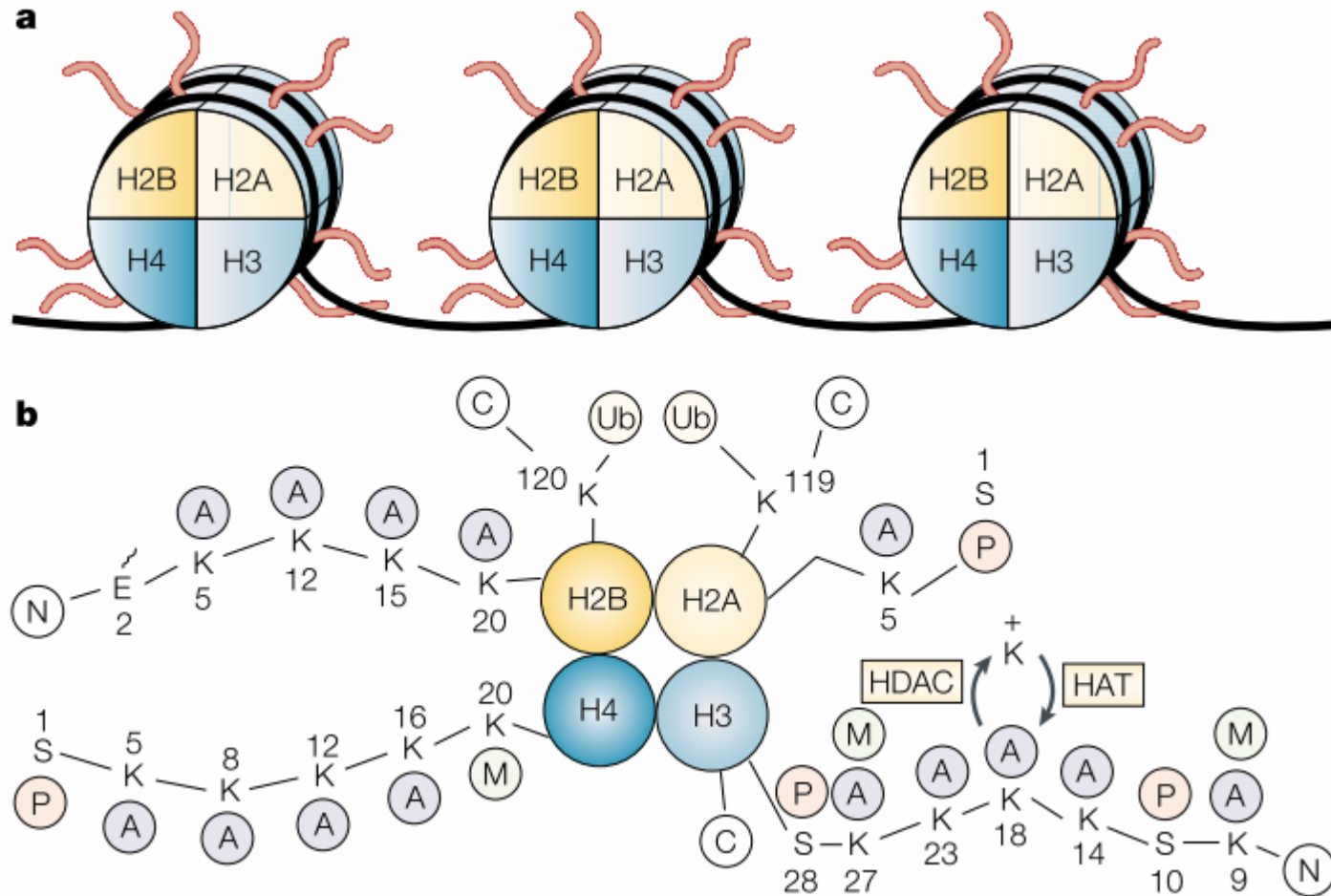


*Nature Reviews Genetics, 2016*

# ChIP-Seq and DNA accessibility sequencing

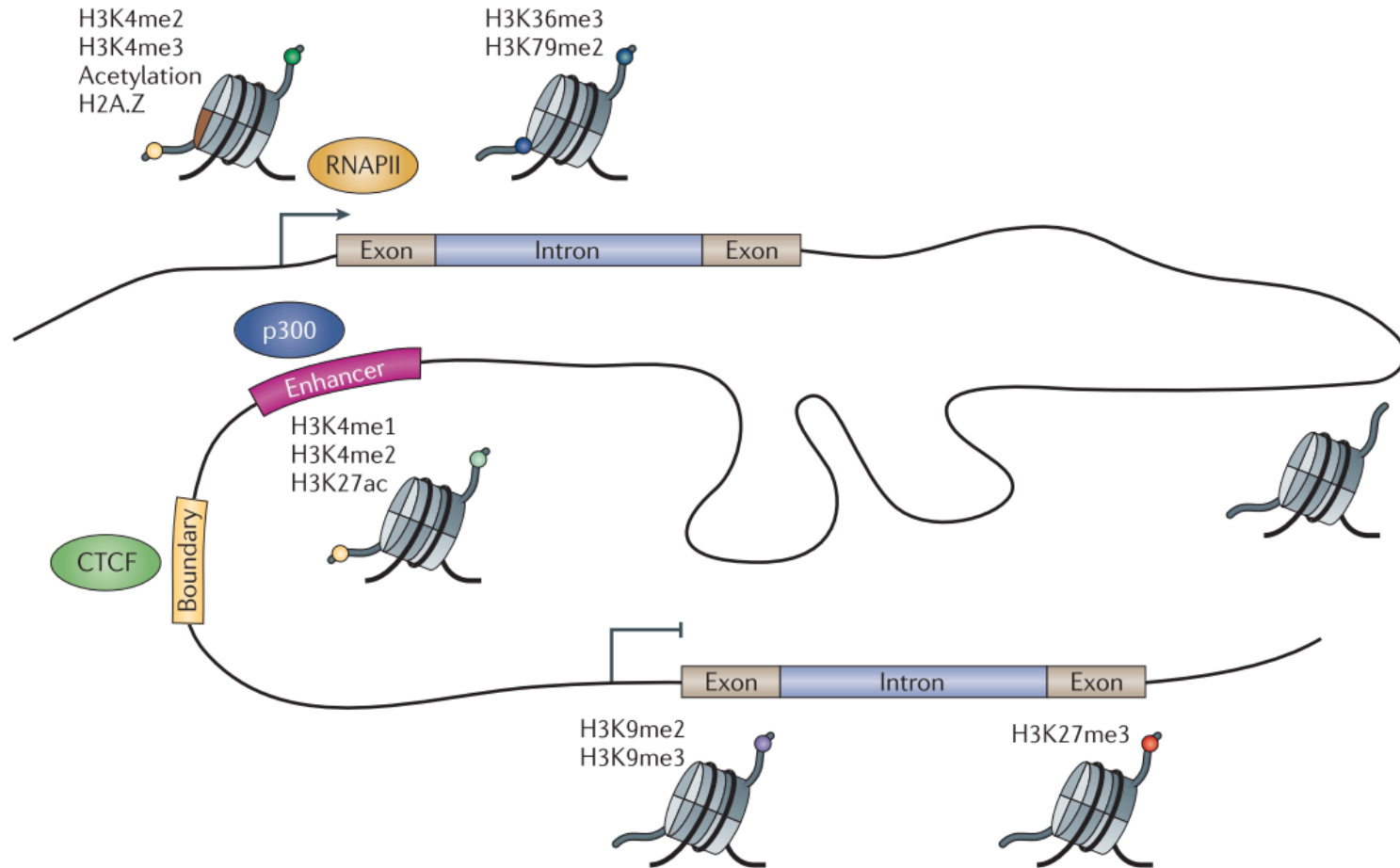


# Histone modification



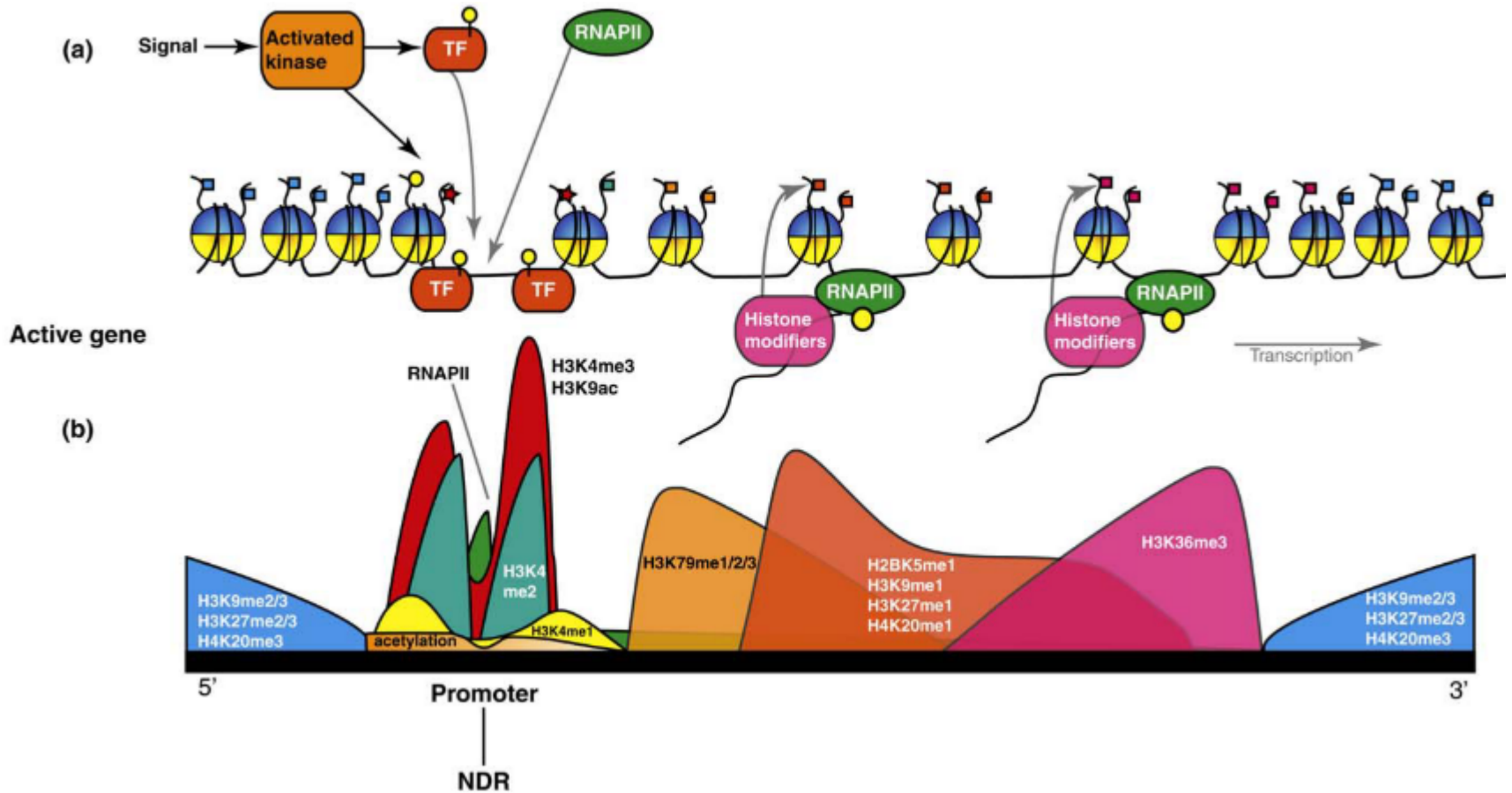
*Nature reviews Cancer, 2001*

# Regulation by histone profile

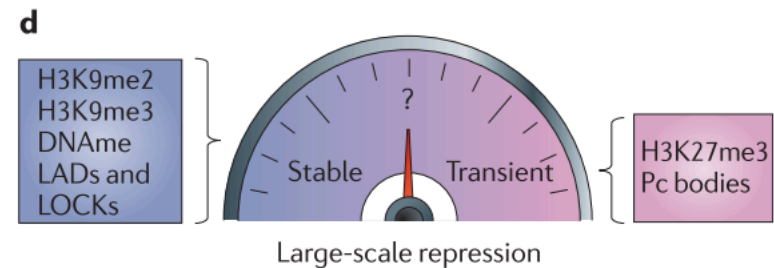
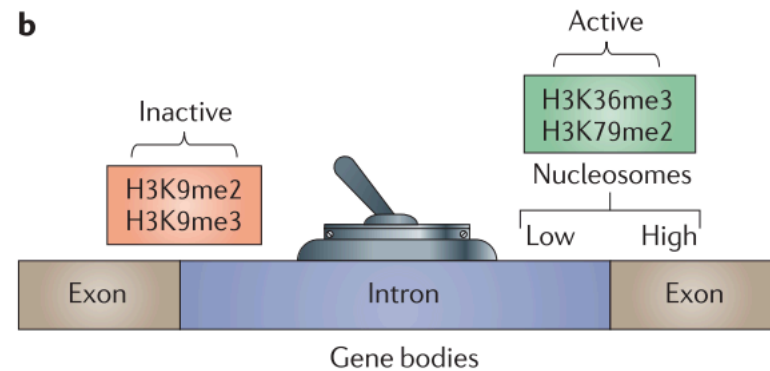
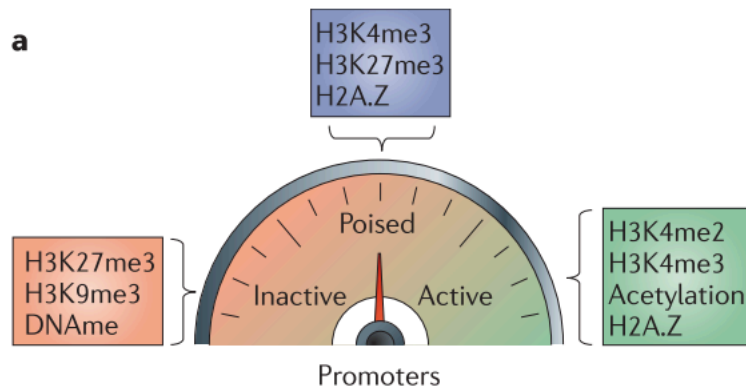


*Nature reviews. Genetics, 2011*

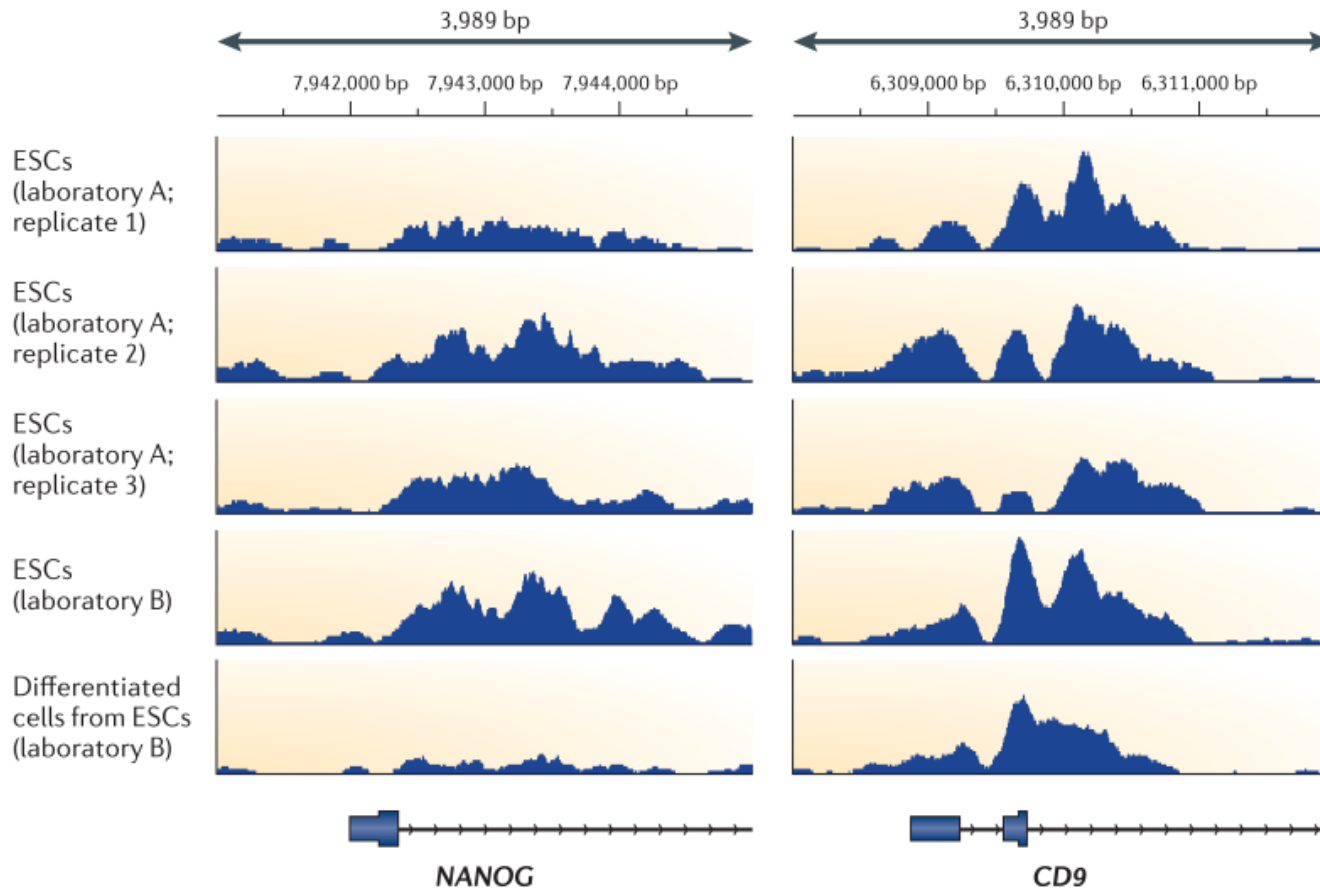
# Active regulation in gene



# 'Dashboard' of histone modifications

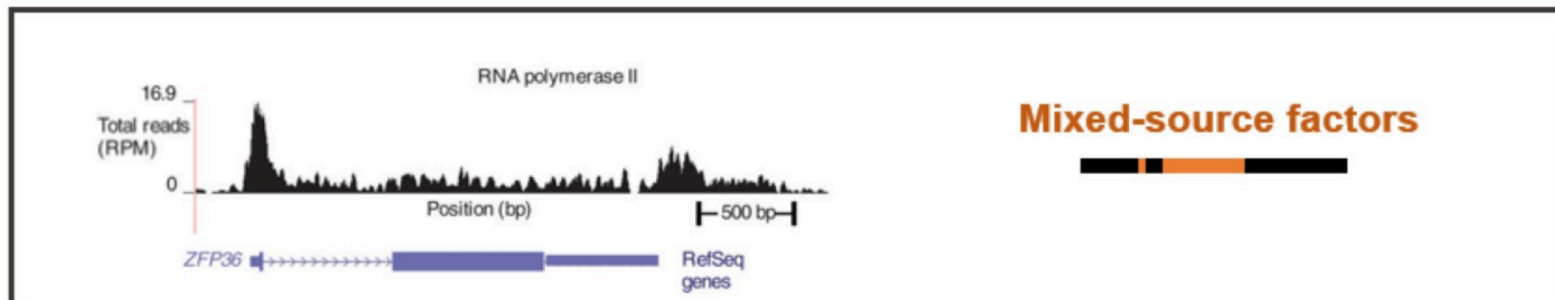
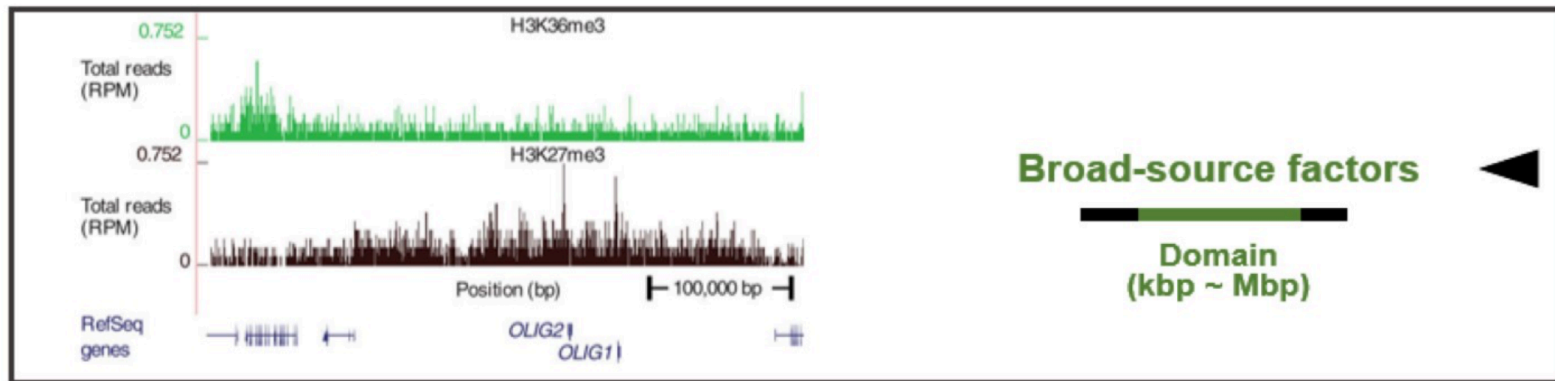
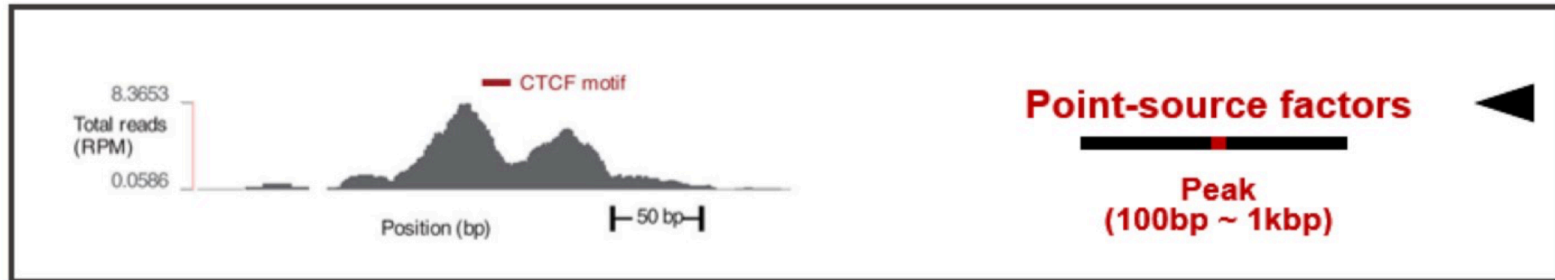


# Histone profile depending on Cell type



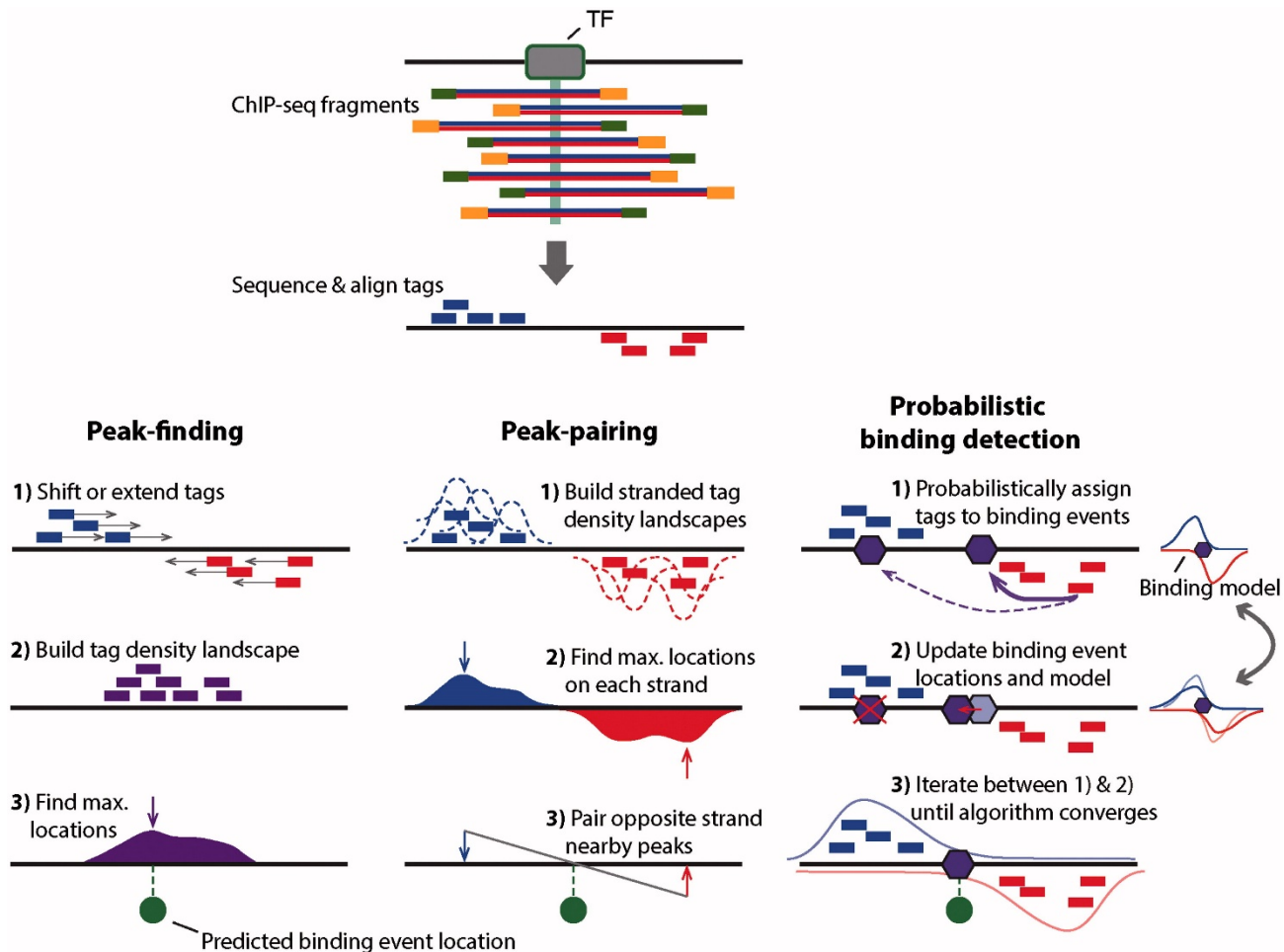
*Nature Reviews Genetics*, 2014

# Classes of DNA-bound proteins



*Nature Reviews Genetics*, 2014

# Principle detection of protein binding region



<https://galaxyproject.org/tutorials/chip/>

# Principle detection of protein binding region

## Design and analysis of ChIP-seq experiments for DNA-binding proteins

Peter V Kharchenko<sup>1-3</sup>, Michael Y Tolstorukov<sup>1,2</sup> & Peter J Park<sup>1-3</sup>

## An integrated software system for analyzing ChIP-chip and ChIP-seq data

Hongkai Ji<sup>1</sup>, Hui Jiang<sup>2</sup>, Wenxiu Ma<sup>3</sup>, David S Johnson<sup>4,8</sup>, Richard M Myers<sup>5</sup> & Wing H Wong<sup>6,7</sup>

## FindPeaks 3.1: a tool for identifying areas of enrichment from massively parallel short-read sequencing technology

Anthony P. Fejes<sup>1,\*</sup>, Gordon Robertson<sup>1</sup>, Mikhail Bilenky<sup>1</sup>, Richard Varhol<sup>1</sup>, Matthew Bainbridge<sup>2</sup> and Steven J. M. Jones<sup>1,\*</sup>

## HPeak: an HMM-based algorithm for defining read-enriched regions in ChIP-Seq data

Zhaohui S Qin<sup>1,2,3</sup>, Jianjun Yu<sup>3,4</sup>, Jincheng Shen<sup>1</sup>, Christopher A Maher<sup>2,3,4</sup>, Ming Hu<sup>1</sup>, Shanker Kalyana-Sundaram<sup>3,4</sup>, Jindan Yu<sup>3</sup> and Arul M Chinnaiyan<sup>2,3,4,6,7,8</sup>

## Model-based Analysis of ChIP-Seq (MACS)

Yong Zhang<sup>\*,†</sup>, Tao Liu<sup>\*,†</sup>, Clifford A Meyer<sup>\*</sup>, Jérôme Eeckhoutte<sup>†</sup>, David S Johnson<sup>‡</sup>, Bradley E Bernstein<sup>§¶</sup>, Chad Nusbaum<sup>¶</sup>, Richard M Myers<sup>‡</sup>, Myles Brown<sup>†</sup>, Wei Li<sup>#</sup> and X Shirley Liu<sup>\*</sup>

## PeakSeq enables systematic scoring of ChIP-seq experiments relative to controls

Joel Rozowsky<sup>1</sup>, Ghia Euskirchen<sup>2</sup>, Raymond K Auerbach<sup>3</sup>, Zhengdong D Zhang<sup>1</sup>, Theodore Gibson<sup>1</sup>, Robert Bjornson<sup>4</sup>, Nicholas Carriero<sup>4</sup>, Michael Snyder<sup>1,2</sup> & Mark B Gerstein<sup>1,3,4</sup>

## BayesPeak: Bayesian analysis of ChIP-seq data

Christiana Spyrou<sup>\*1,3</sup>, Rory Stark<sup>3</sup>, Andy G Lynch<sup>4</sup> and Simon Tavaré<sup>2,4</sup>

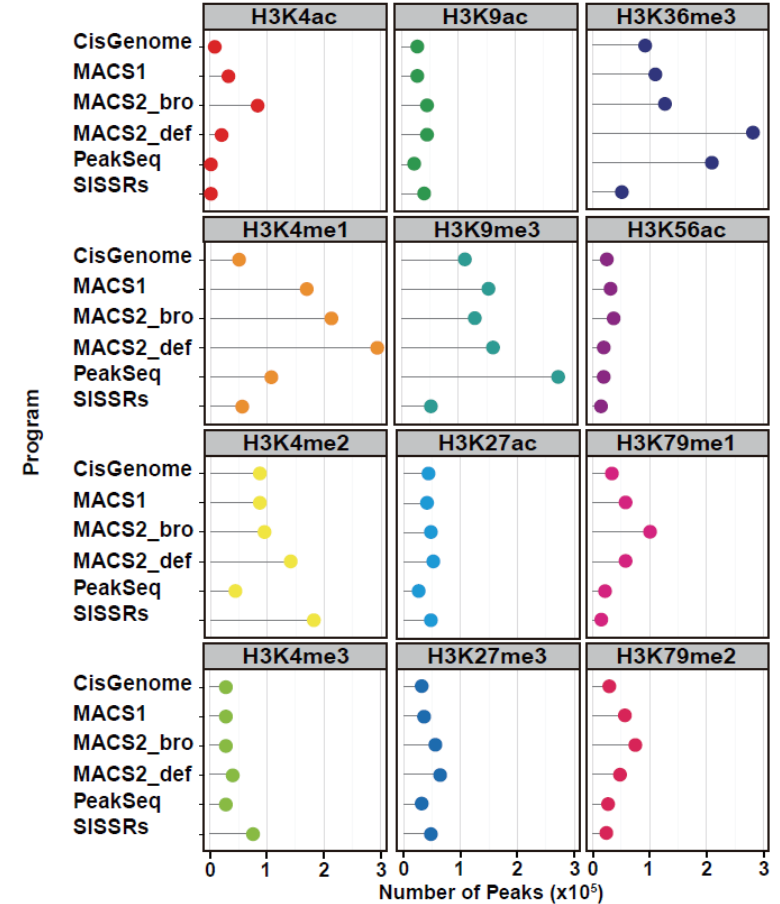
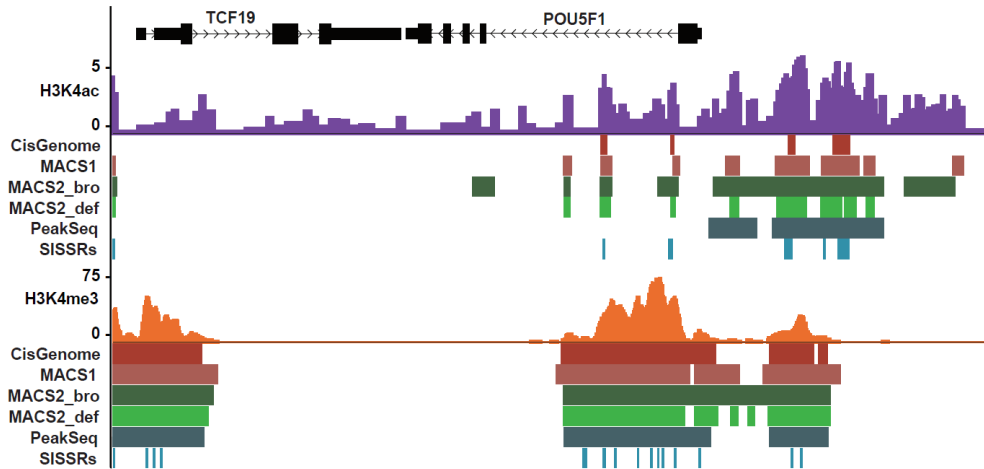
## Genome-wide identification of *in vivo* protein-DNA binding sites from ChIP-Seq data

Raja Jothi, Suresh Cuddapah, Artem Barski, Kairong Cui and Keji Zhao<sup>\*</sup>

## Sole-Search: an integrated analysis program for peak detection and functional annotation using ChIP-seq data

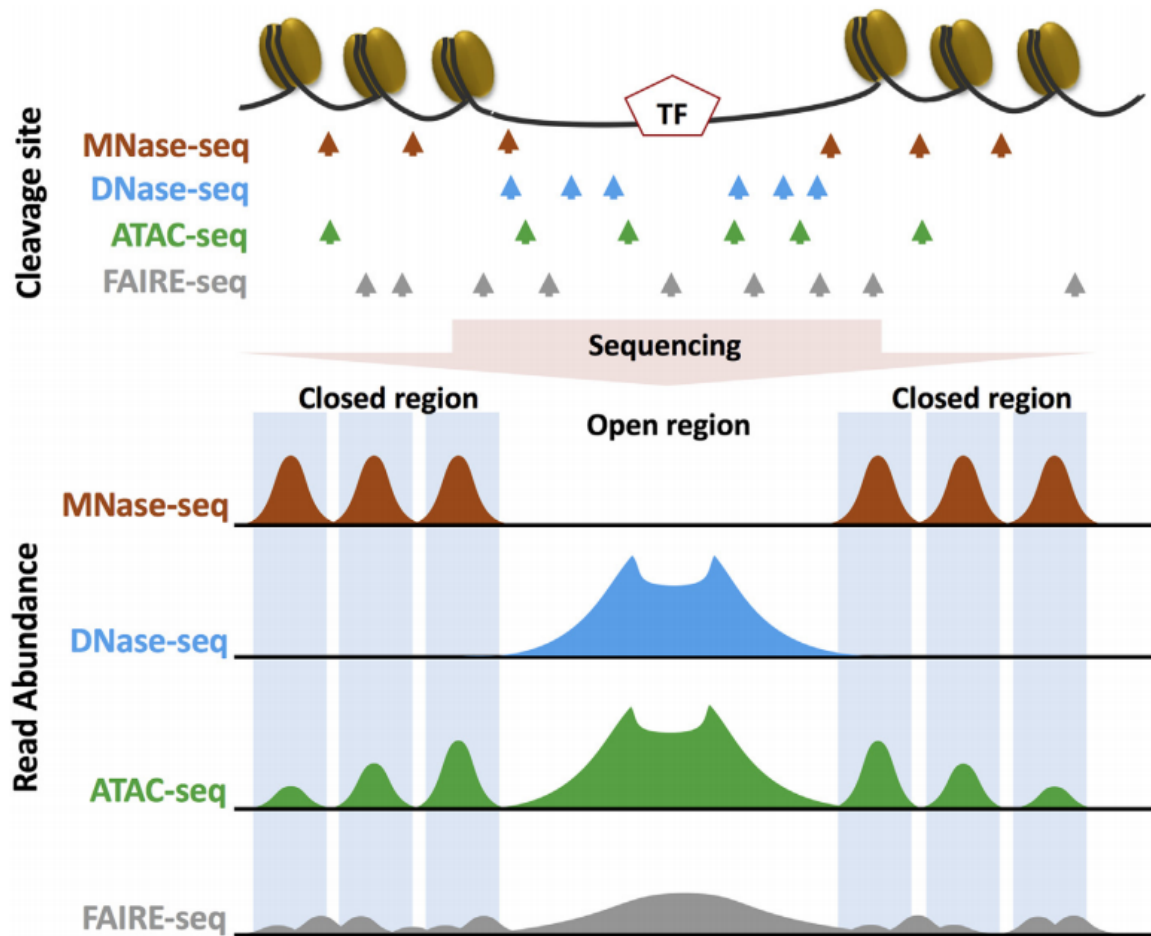
Kimberly R. Blahnik<sup>1</sup>, Lei Dou<sup>1</sup>, Henriette O'Geen<sup>1</sup>, Timothy McPhillips<sup>1</sup>, Xiaoqin Xu<sup>1</sup>, Alina R. Cao<sup>1</sup>, Sushma Iyengar<sup>1</sup>, Charles M. Nicolet<sup>1</sup>, Bertram Ludäscher<sup>1,2</sup>, Ian Korf<sup>1,3</sup> and Peggy J. Farnham<sup>1,4,\*</sup>

# Difference of peak result depending on program

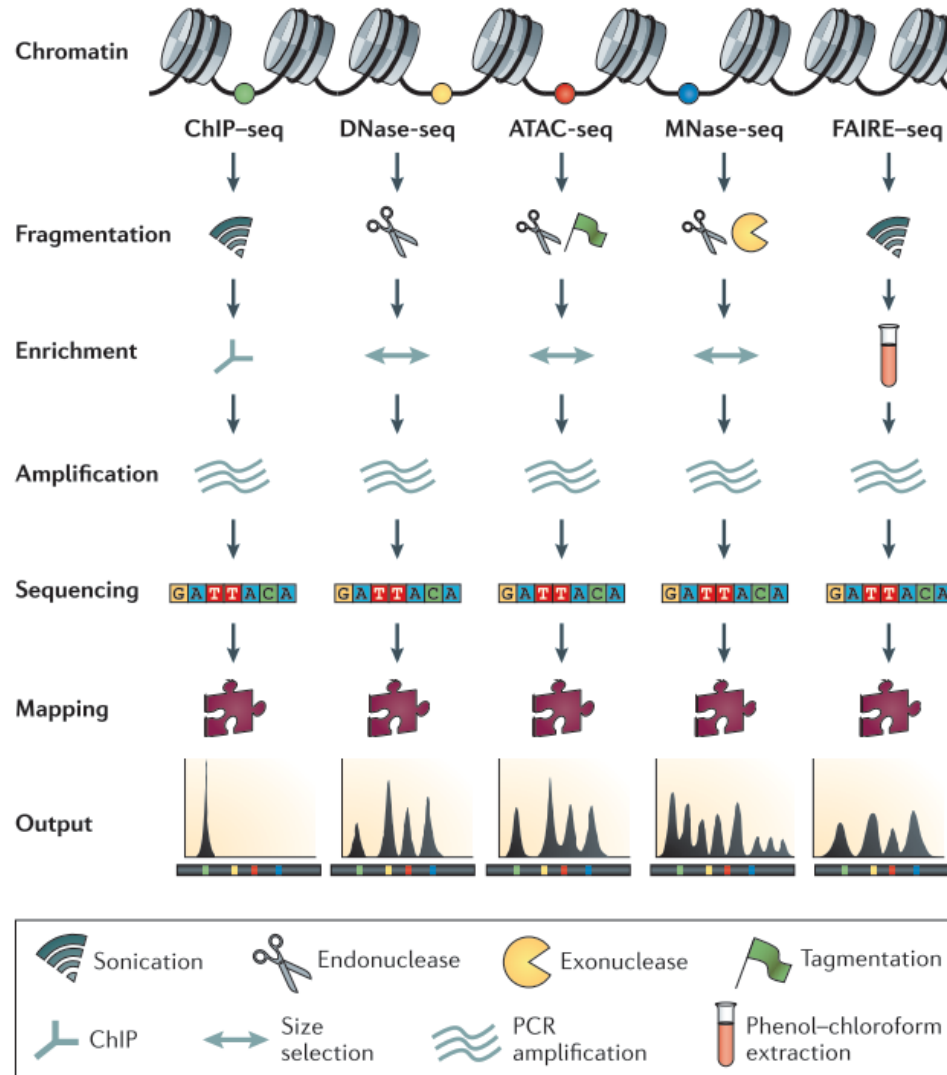


Unpublished

# Active regulation in gene

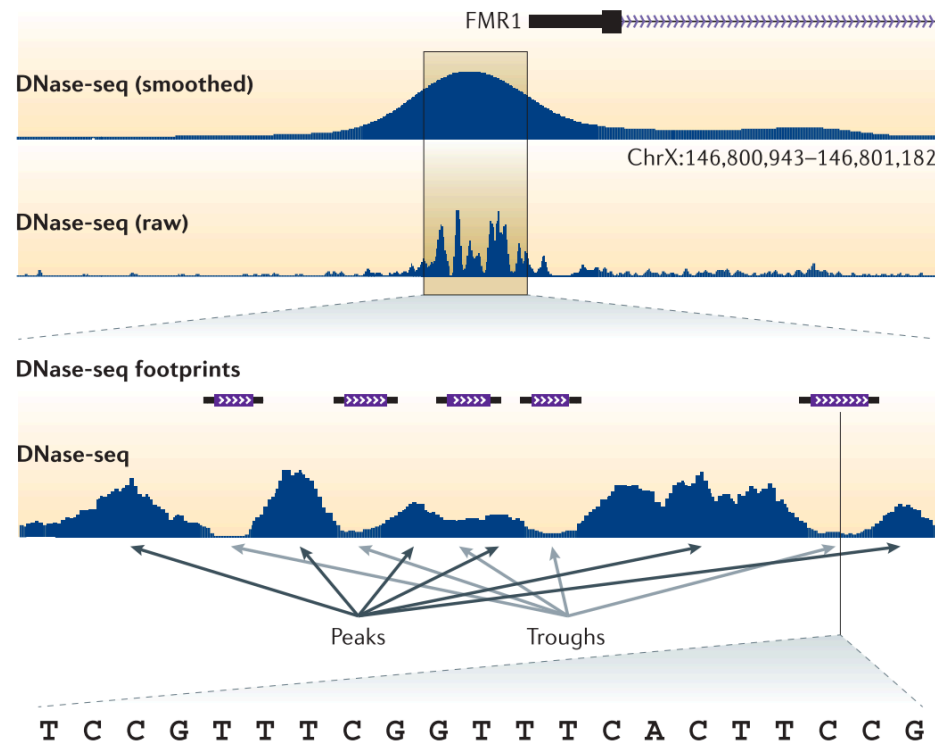


# Sequencing to investigate DNA accessibility



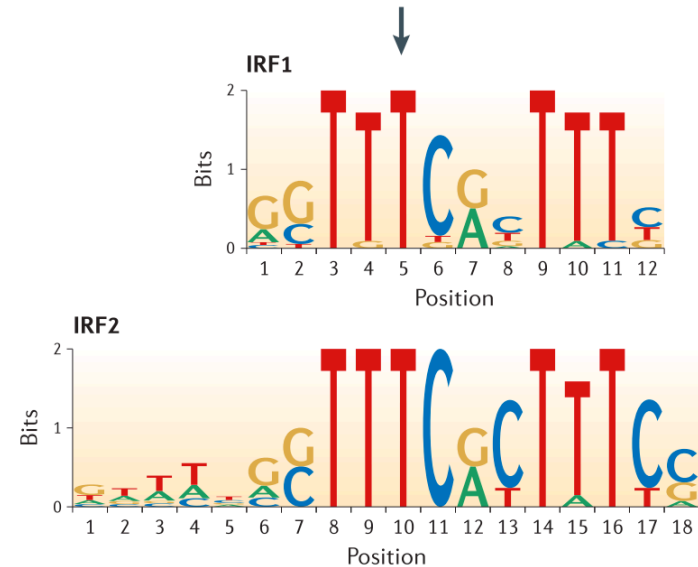
*Nature Reviews Genetics, 2014*

# Sequencing to investigate DNA accessibility



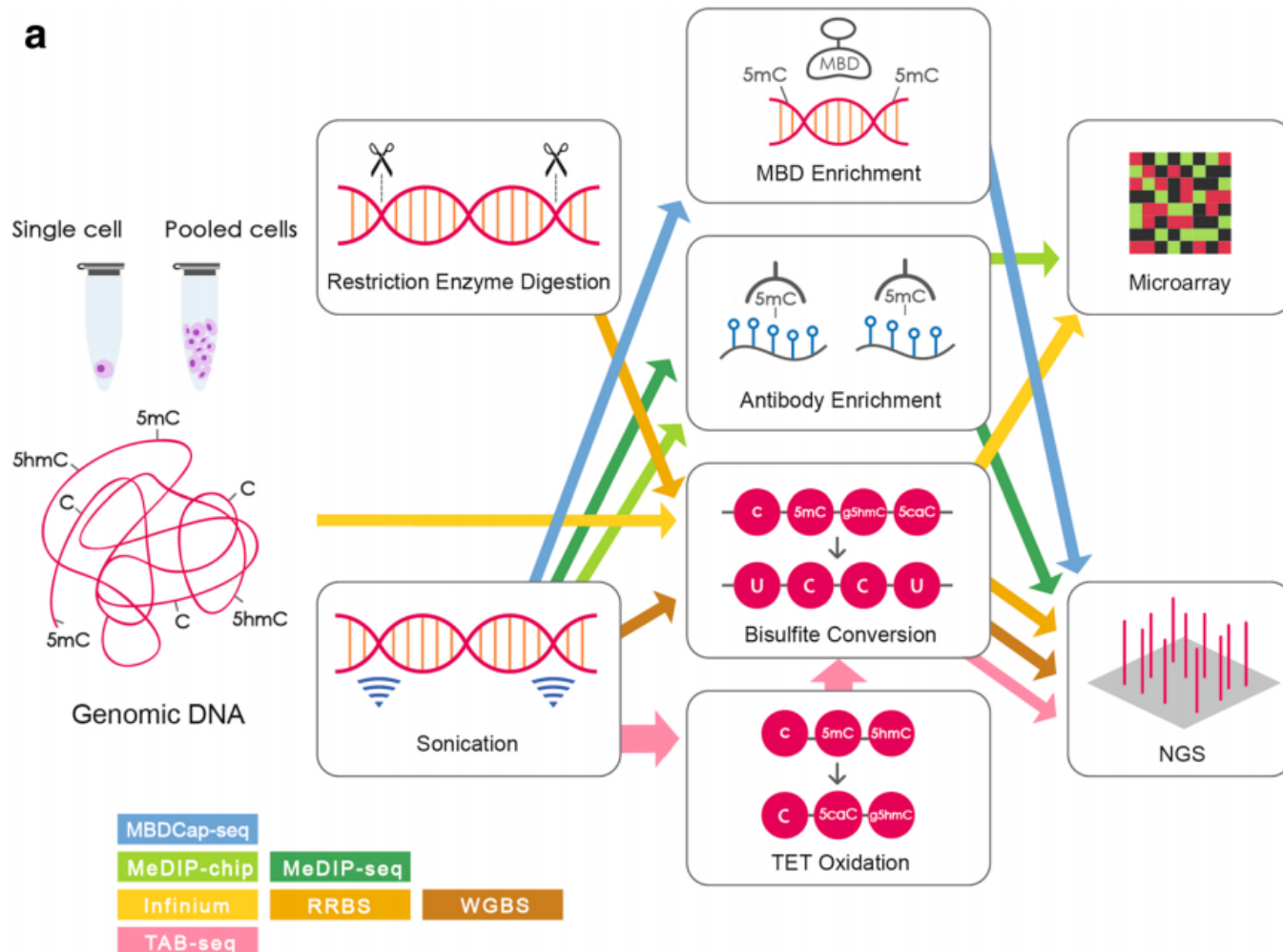
Motifs from JASPAR database

Model name	Score	Relative score	Start	End	Strand	Predicted site sequence
IRF1	12.986	0.904279917181229	3	14	-1	GAAACCGAAACG
IRF2	17.216	0.907706906384892	4	21	-1	CGGAAGTGAAACCGAAAC
SPIB	4.820	0.806987596140569	5	11	-1	ACCGAAA
BRCA1	4.228	0.802287513481405	8	14	-1	GAAACCG



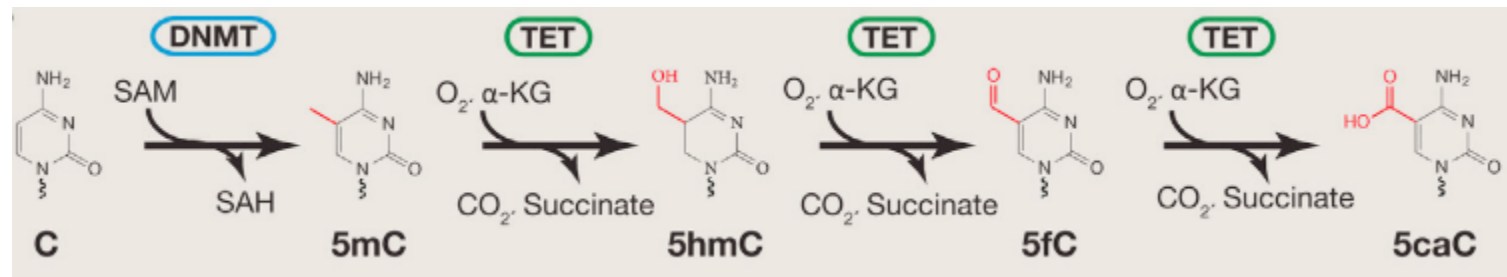
*Nature Reviews Genetics, 2014*

# Sequencing method to measure methylation

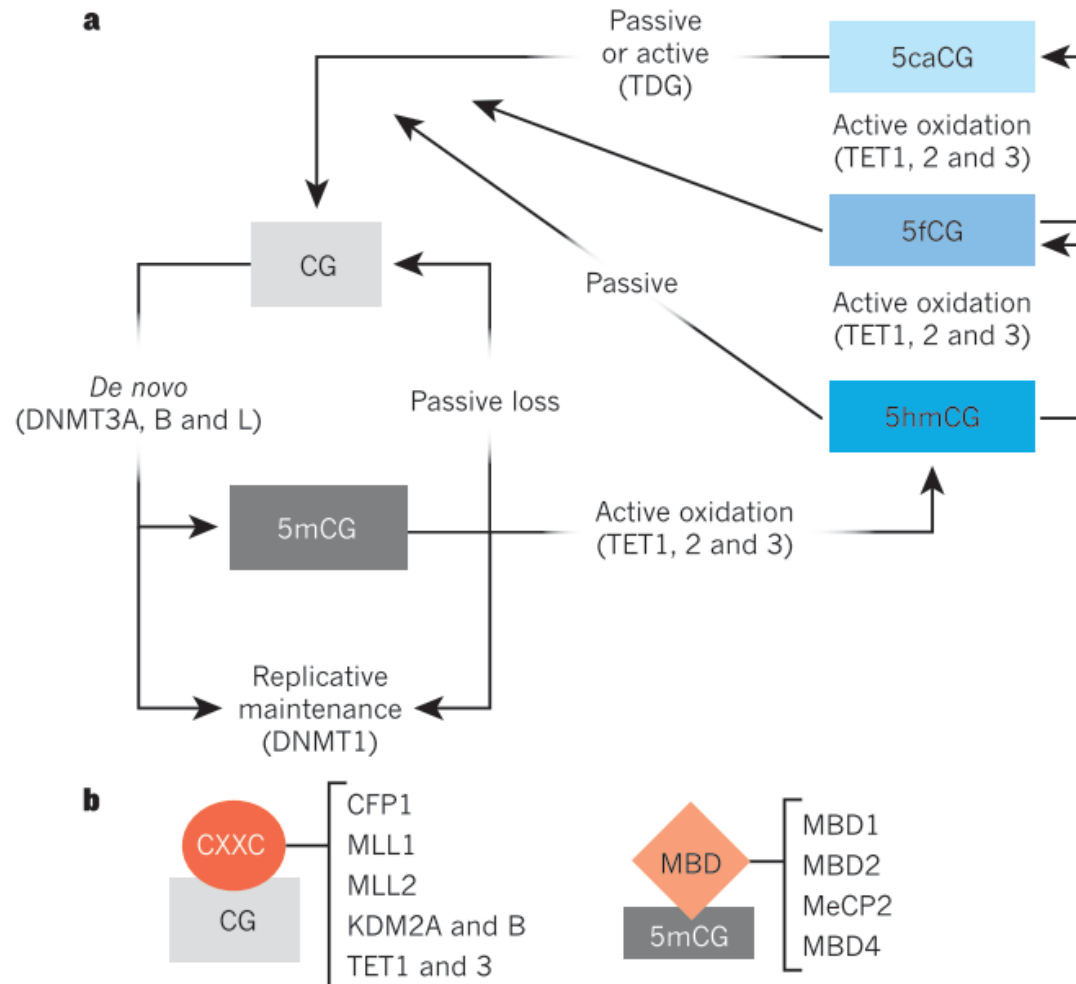


*Cell, 2014*

# DNA methylation mechanism

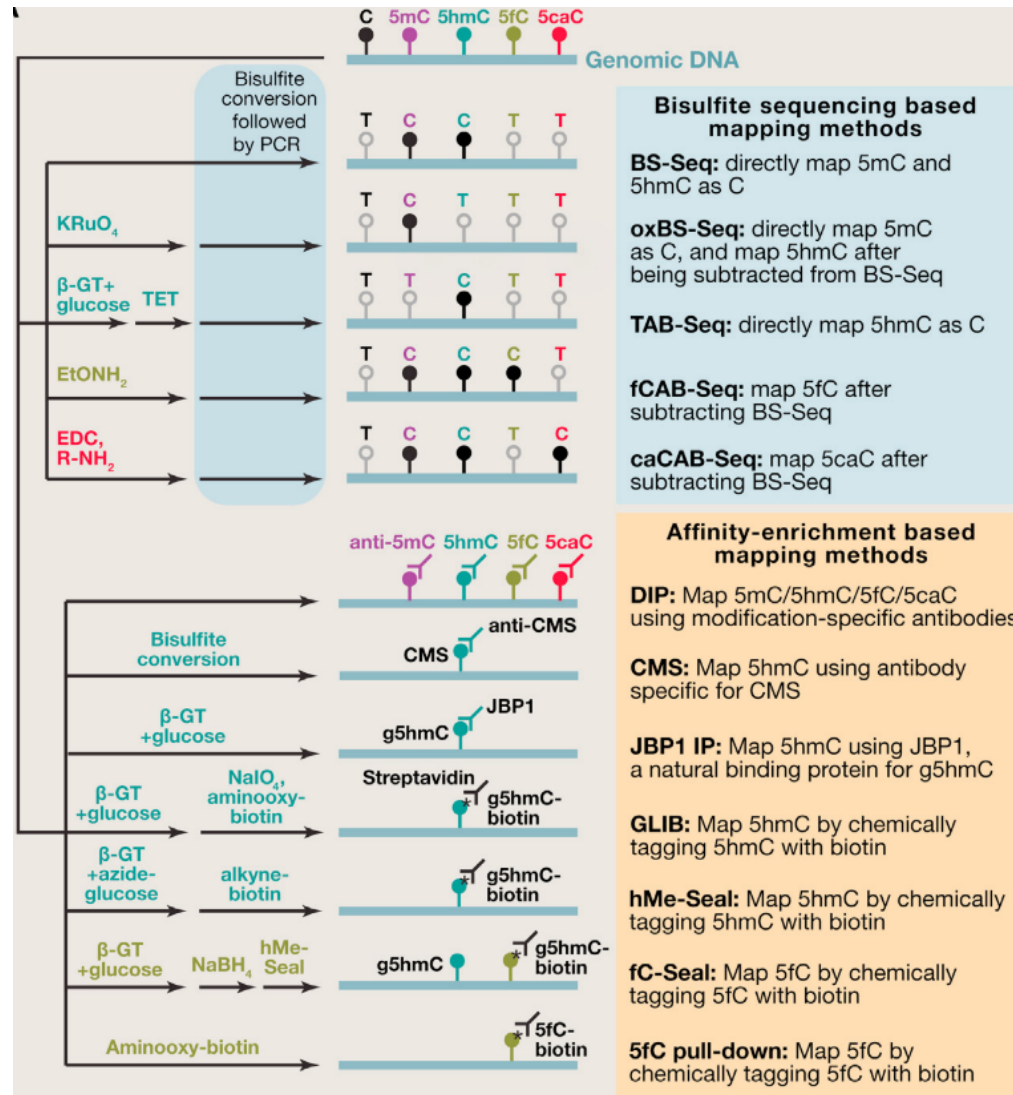


# DNA methylation mechanism

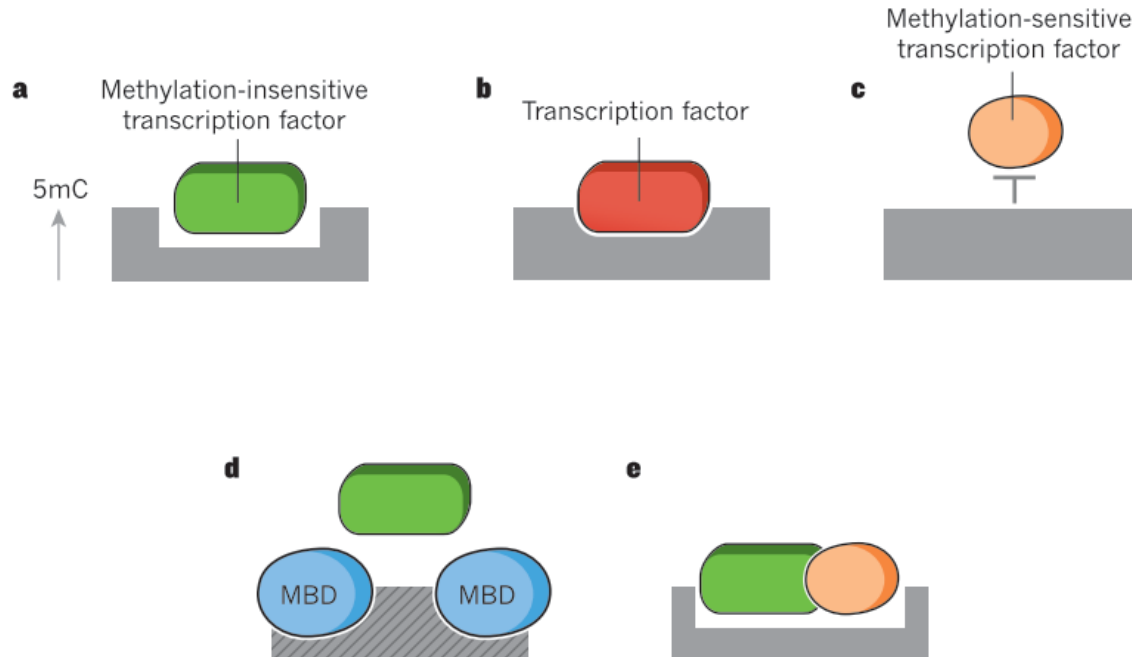


*Nature, 2015*

# Sequencing for methylation



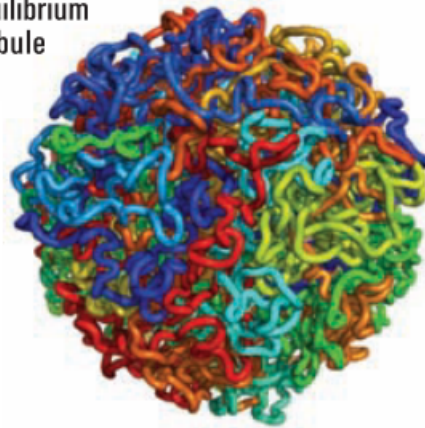
# DNA methylation function



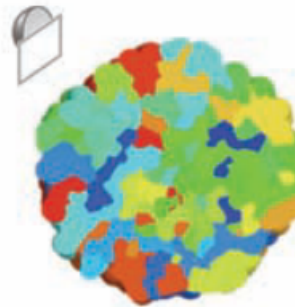
# Chromatin structure

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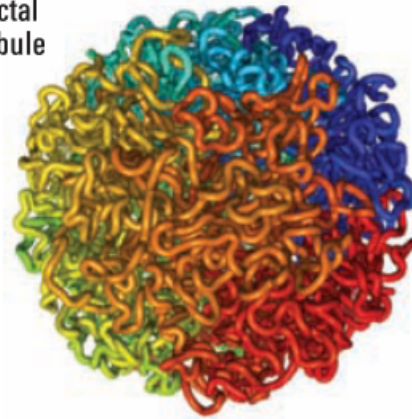
Equilibrium  
globule



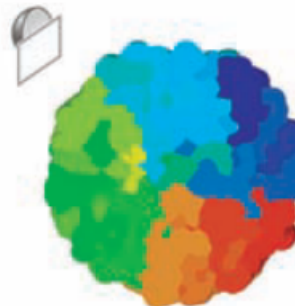
Cross-section view



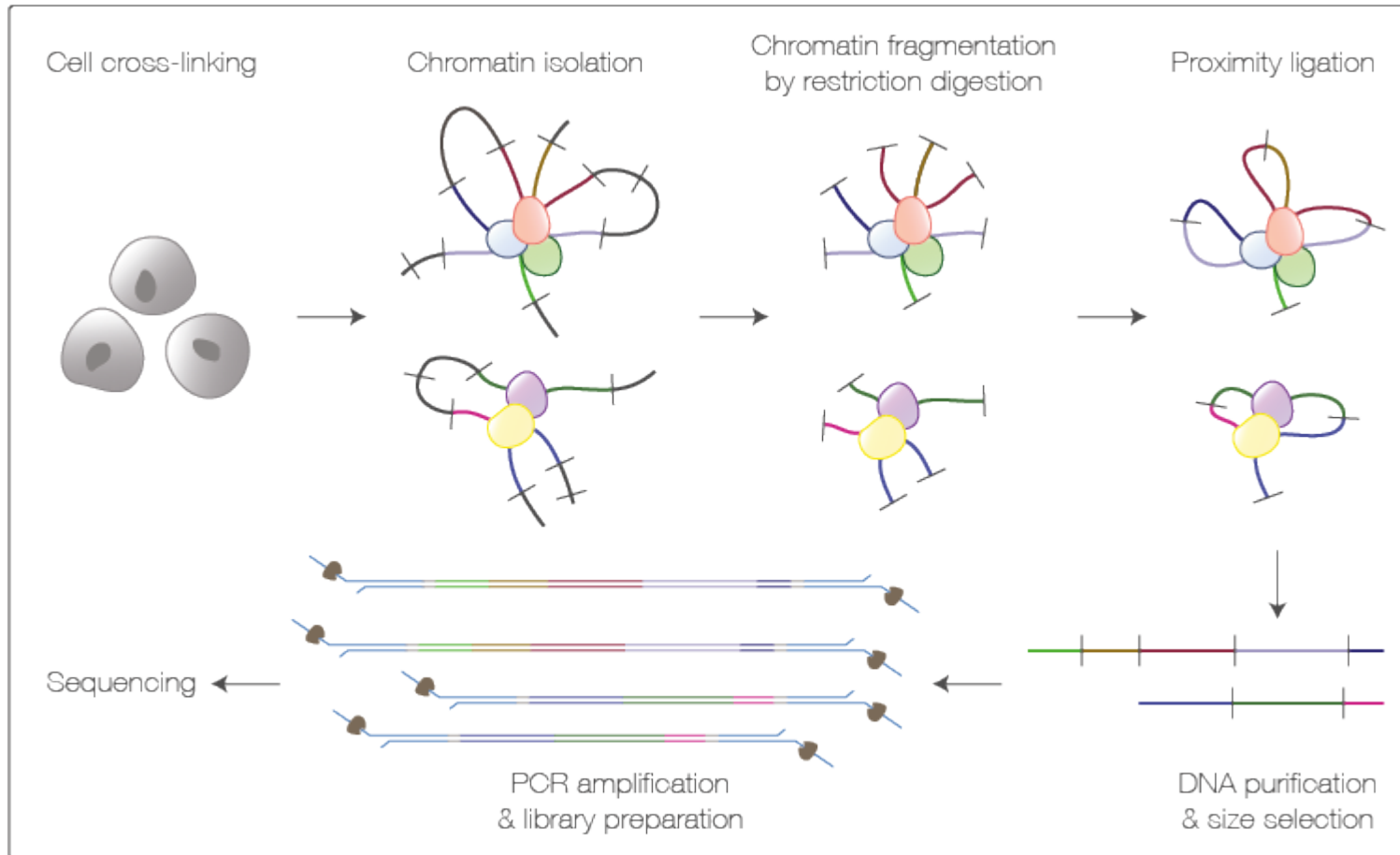
Fractal  
globule



Cross-section view

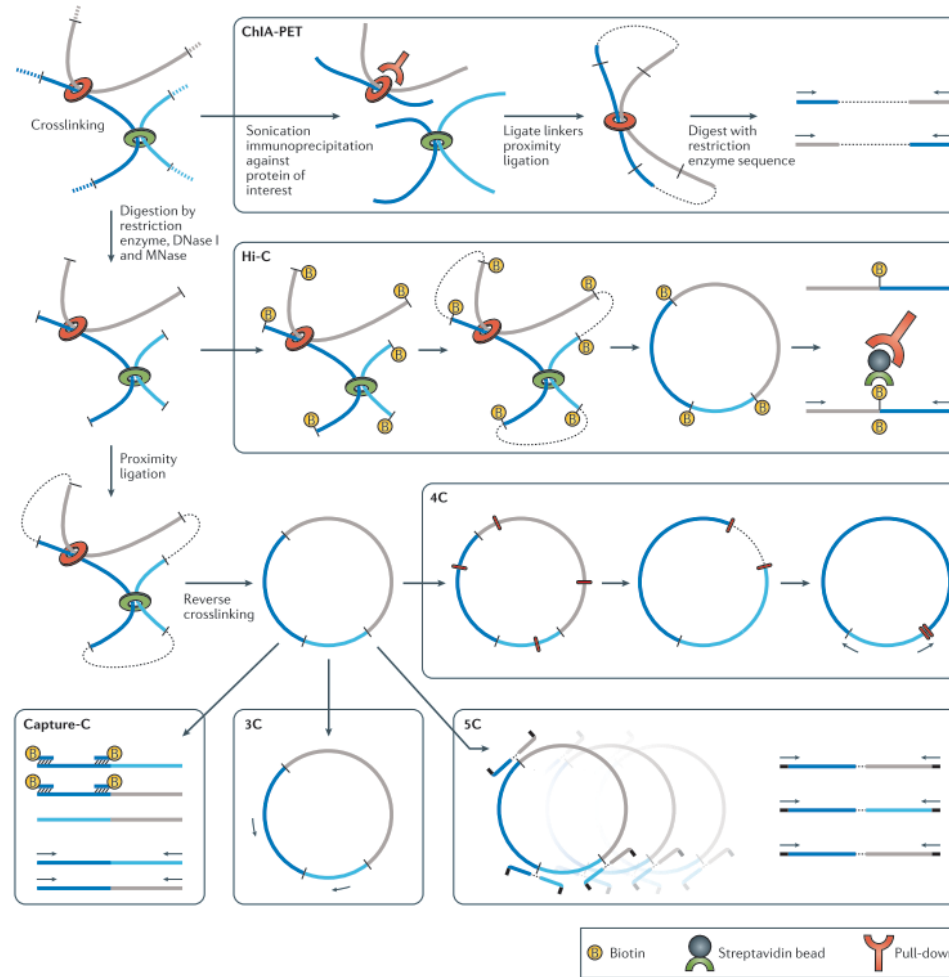


# Chromatin Conformation Capture(3C)



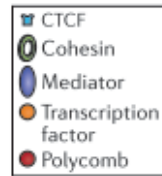
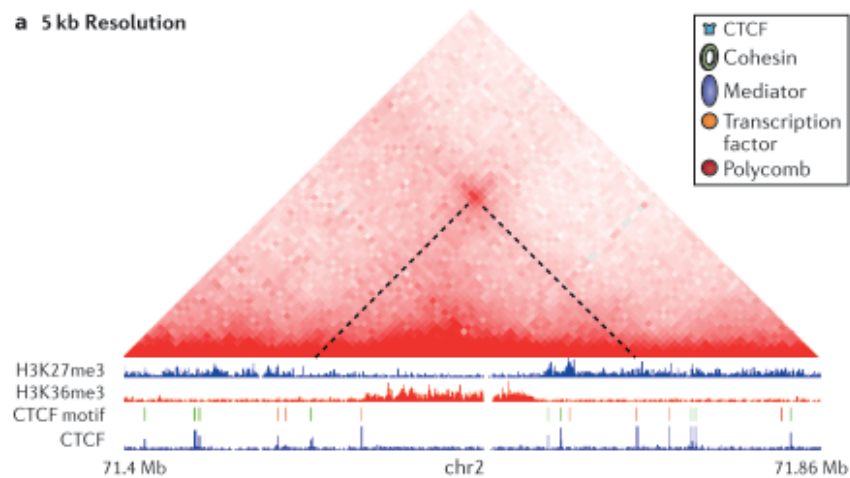
<https://nanoporetech.com/resource-centre/pore-c-using-nanopore-reads-delineate-long-range-interactions-between-genomic-loci#image1&>

# 3C-based approaches

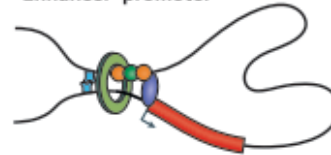


# Conformation in genome

a 5 kb Resolution



Enhancer-promoter



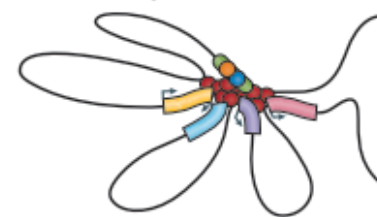
Gene loop



Architectural loop

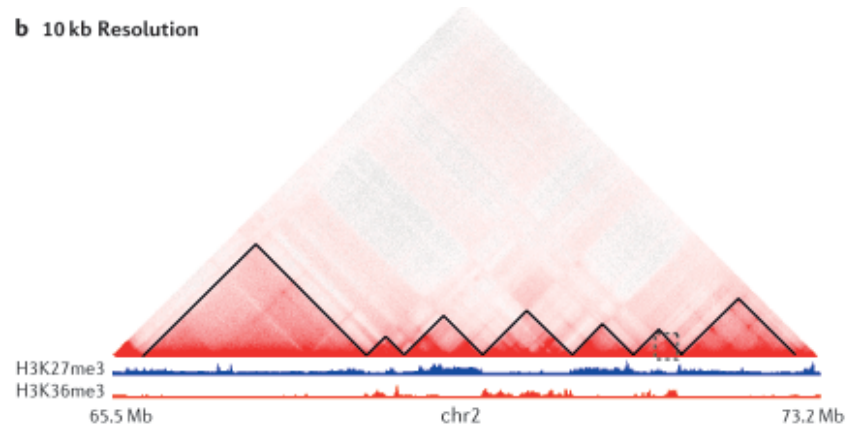


Polycomb-mediated

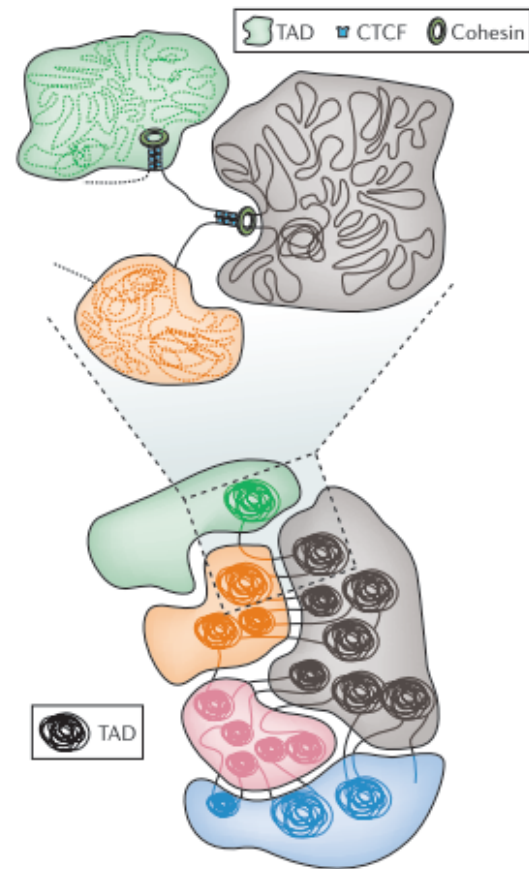
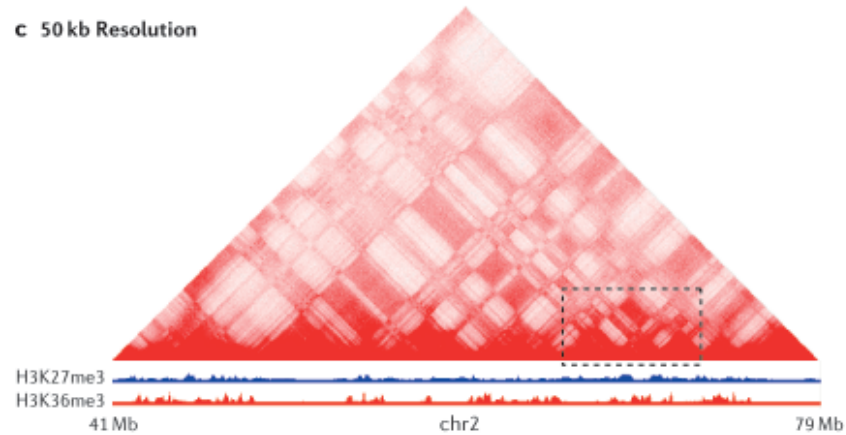


# Conformation in genome

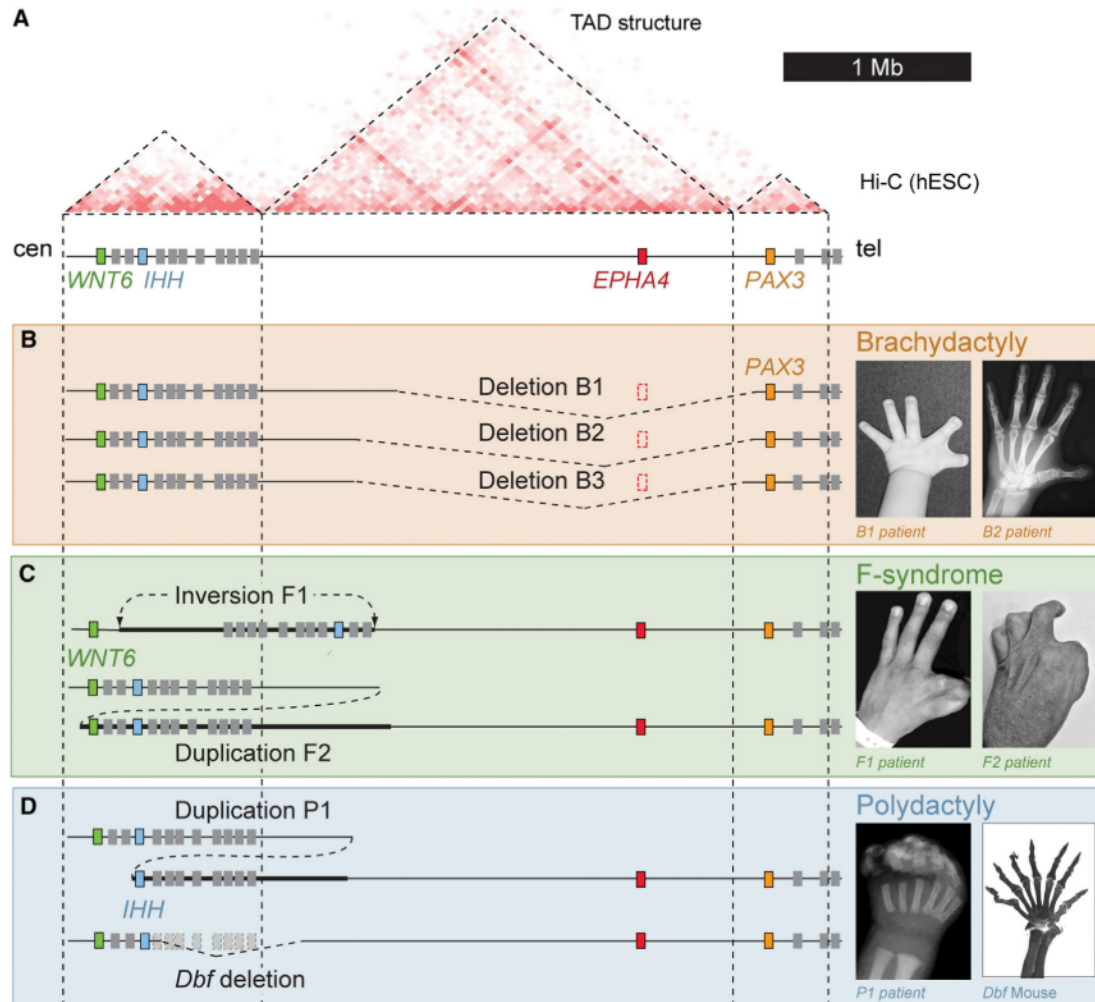
**b** 10 kb Resolution



**c** 50 kb Resolution

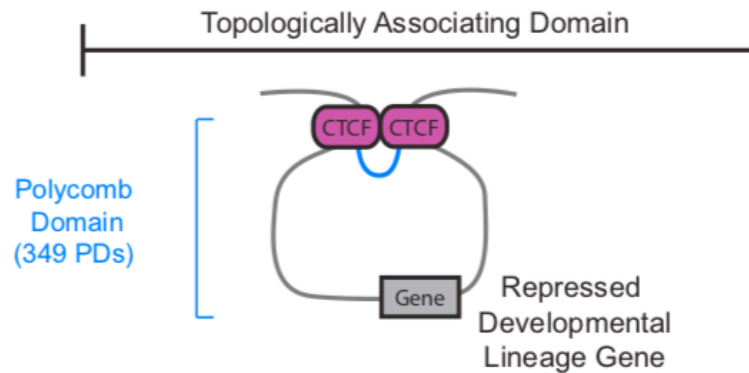
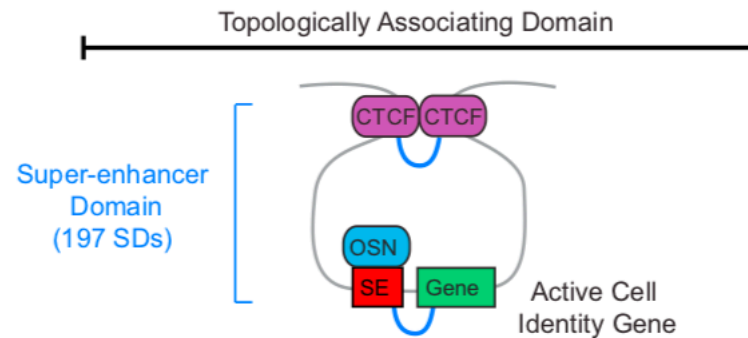


# Diseases associated to chromatin conformation

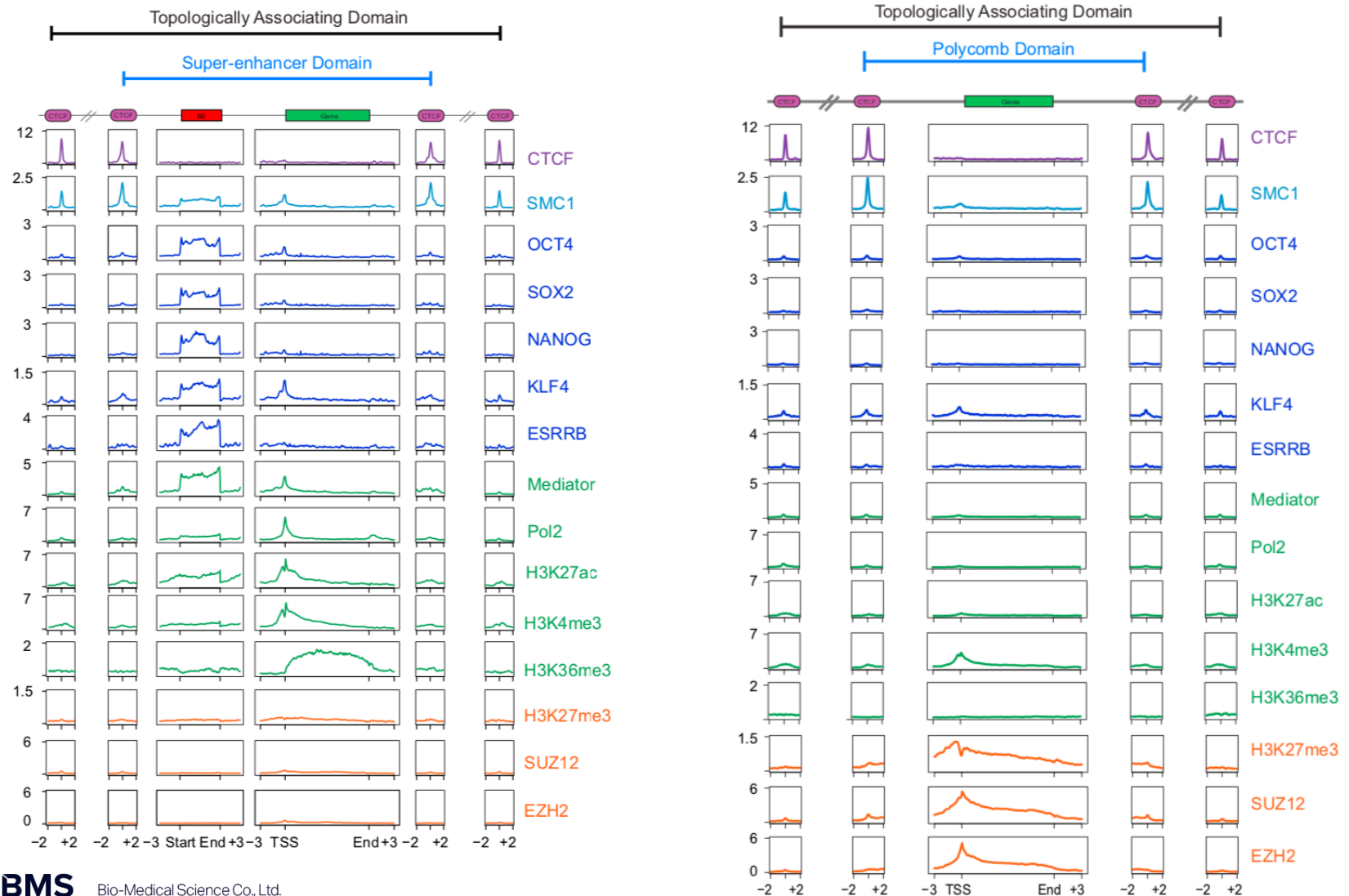


Cell, 2015

# Interplay among epigenome feature



# Interplay among epigenome feature



# Interplay among epigenome feature

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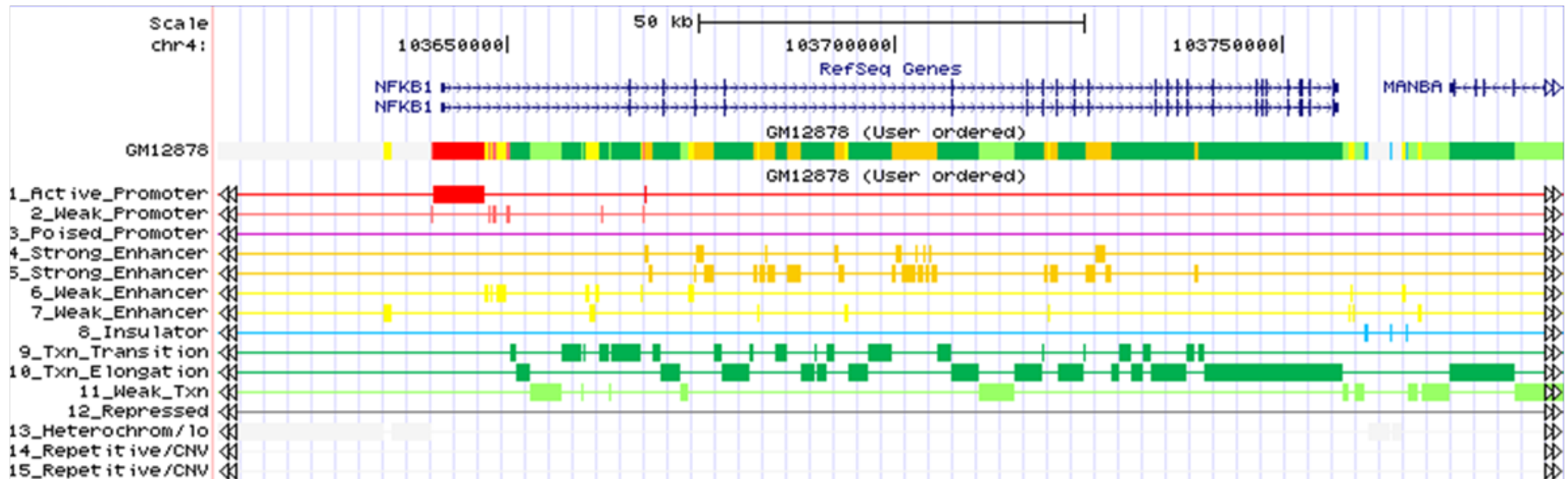
Cell Type A



Cell Type B



# Multi-omics analysis



THANK YOU.

**BMS**